

Review

Guang Gao*, James Grant Burgess, Min Wu, Shujun Wang and Kunshan Gao*

Using macroalgae as biofuel: current opportunities and challenges

<https://doi.org/10.1515/bot-2019-0065>

Received 9 September, 2019; accepted 18 February, 2020

Abstract: The rising global demand for energy and the decreasing stocks of fossil fuels, combined with environmental problems associated with greenhouse gas emissions, are driving research and development for alternative and renewable sources of energy. Algae have been gaining increasing attention as a potential source of bio-renewable energy because they grow rapidly, and farming them does not, generally, compete for agricultural land use. Previous studies of algal biofuels have focused on microalgae because of their fast growth rate and high lipid content. Here we analyze the multiple merits of biofuel production using macroalgae, with particular reference to their chemical composition, biomass and biofuel productivity, and cost-effectiveness. Compared to microalgae, macroalgae have lower growth rates and energy productivity but higher cost-effectiveness. A biomass productivity of over 73.5 t dry mass ha⁻¹ year⁻¹ with a methane yield of 285 m³ t⁻¹ dry mass would make electricity production from macroalgae profitable, and this might be achieved using fast-growing macroalgae, such as *Ulva*. Taking into account the remediation of eutrophication and CO₂, exploring macroalgae for a renewable bioenergy is of importance and feasible.

Keywords: biofuel; biogas; bioremediation; cultivation; macroalgae; photosynthesis.

***Corresponding authors:** Guang Gao, State Key Laboratory of Marine Environmental Science, Xiamen University, Xiamen 361005, China; and Jiangsu Key Laboratory for Marine Bioresources and Environment, Jiangsu Ocean University, Lianyungang 222005, China, e-mail: guang.gao@xmu.edu.cn; and Kunshan Gao, State Key Laboratory of Marine Environmental Science, Xiamen University, Xiamen 361005, China, e-mail: ksgao@xmu.edu.cn

James Grant Burgess: School of Natural and Environmental Sciences, Newcastle University, Newcastle upon Tyne NE1 7RU, UK

Min Wu and Shujun Wang: Jiangsu Key Laboratory for Marine Bioresources and Environment, Jiangsu Ocean University, Lianyungang 222005, China

Introduction

Today, approximately 85% of the total energy consumed worldwide is provided by fossil fuels (Dudley 2018). Stocks of fossil fuels are declining, and there is growing concern regarding the serious environmental problems associated with their consumption (Ravi et al. 2018, Saratale et al. 2018). Consequently, it is of general importance to search for renewable and cost-effective energy sources with low or zero greenhouse gas emissions (Ravi et al. 2018). Biofuels, mainly extracted from plants, can serve as an attractive source of energy to meet some of the present and future fuel demand (Robertson et al. 2017). Currently, bio-ethanol and biodiesel, which are produced primarily from food crops such as grain, sugar cane and vegetable oils, are the most widely available forms of biofuel (Landis et al. 2017, Adeniyi et al. 2018). However, these first generation biofuels suffer from concerns regarding their possible impact on food supply and security (Herrmann et al. 2018). For instance, the UK consumed an estimated 47 billion liters of transport fuel in 2008, 53% of which was diesel. More than half the land area of the UK would be needed if this diesel were to be produced from oilseed rape (Stephenson et al. 2008). As a result, increasing attention is being focused on non-food based biofuels and feedstocks. In comparison to other feedstocks, such as sugarcane, algae can provide a high-yield source of biofuels without competing with food production (Chisti 2007, Adeniyi et al. 2018). For instance, to meet the energy requirement for transport in the UK in 2008, only 2.17% and 6.63% land area of the UK is needed if diesel is produced from microalgae with 30% lipid content (Chisti 2007) and from *Ulva lactuca* Linnaeus (Bruhn et al. 2011), respectively. In addition, algae also display a potential for CO₂ sequestration (Gao and McKinley 1994, Duarte et al. 2013, 2017).

The first consideration when developing algal biofuels is the choice of species or strains that contain high oil-like precursors. Algae are aquatic photosynthetic organisms which can generally be divided into two groups: microalgae (unicellular plants) and macroalgae (or seaweeds). Microalgae appear to be the only source of biodiesel that has the potential to completely displace fossil diesel

because of their fast growth rate and rich lipid content (Singh et al. 2011, Jiang et al. 2016, Dickinson et al. 2017). Biomass doubling times for microalgae are generally less than 24 h. In addition, the lipid content of some microalgae is close to 80% dry weight (Spolaore et al. 2006, Chisti 2007, Bwapwa et al. 2017). Thanks to their fast growth rate and high lipid content, biodiesel productivity from microalgae with 30% (w/w) lipid content in algal biomass could be up to 342 or 92 times more when compared to corn or soybean, respectively (Mata et al. 2010). However, based on previous studies, there is still a long way to go to bring biodiesel from microalgae to the market because of the high cost of biomass production and extraction (Sheridan 2013, Ajjawi et al. 2017, Borowitzka and Vonshak 2017, Posewitz 2017, Remmers et al. 2018).

The use of macroalgae for CO₂ bioremediation has been explored previously (Gao and McKinley 1994, Chung et al. 2011, Sondak et al. 2017). Nevertheless, macroalgae have been less studied as a source of biofuel, although there was a pilot project during the 1980s in the USA (North 1987). However, macroalgae are commonly known to have high levels of carbohydrates (Kraan 2010, Gao et al. 2017a, 2018a), which can be fermented into biogas. They are also more cost-efficient with respect to farming and processing compared to

microalgae (see the section “Economics of biofuels from algae”). We therefore set out to examine the feasibility of using macroalgae as biofuel by analyzing their chemical composition, biomass productivity and biofuel productivity, and cost-effectiveness. In terms of the macroalgal biofuels, we focus on biomethane as it is deemed to have the highest energy yield in comparison to biodiesel and bioethanol (Harun et al. 2011, El-Mashad 2015, Wu et al. 2019). The literature was obtained from Web of Science for the period from 1980 to 2019 when searching with related keywords. The availability of the data was also considered during this selection of literature. While there are a great number of scientific articles about algal biofuels, we only analyzed and cited those that are most relevant to the topic of this review.

Chemical composition

The chemical composition of plant biomass is a primary factor to be considered for generation of biofuels as it determines not only which method should be employed to process the biomass but also the production yield of biofuel. As shown in Figure 1A and Supplementary Table S1, carbohydrates dominate macroalgal chemical

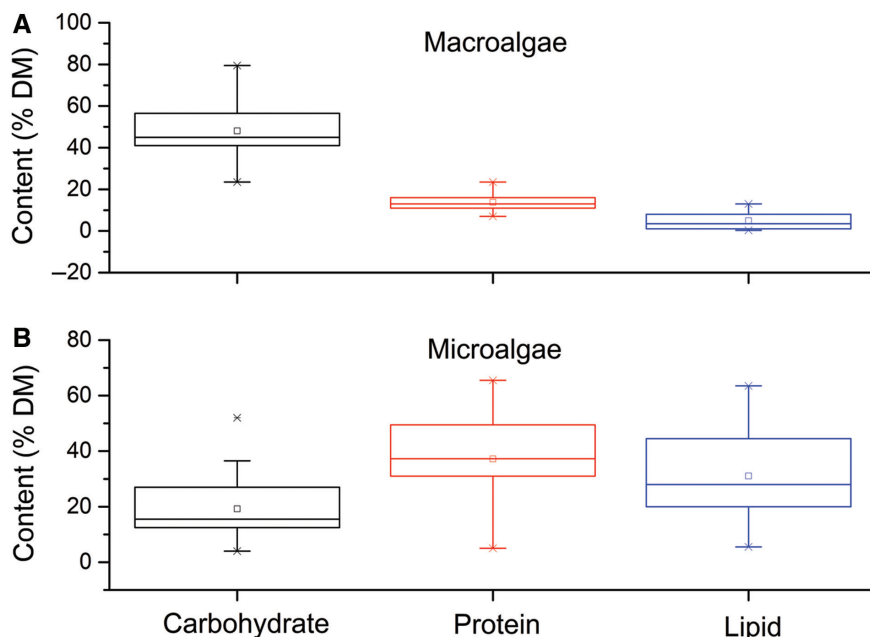


Figure 1: Box charts of main biochemical composition of macroalgae (A) and microalgae (B).

DM, Dry mass. Data were based on Renaud et al. (1994), Horn (2000), Pádua et al. (2004), Chisti (2007), Demirbas (2010), Yoon et al. (2010), Kim et al. (2011), Satyanarayana et al. (2011), Jang et al. (2012), Hong et al. (2014), Khattoon et al. (2014), Tibbetts et al. (2015), Jiang et al. (2016), Zheng et al. (2016), Chan and Matanjun (2017), Cheng et al. (2017), Diprat et al. (2017), Gao et al. (2017a, 2019), Kalita et al. (2017), Shakya et al. (2017), Wang et al. (2017), Abomohra et al. (2018), Gao et al. (2018b), Ishika et al. (2018), Shuba and Kifle (2018), Uribe et al. (2018). Please see Supplementary Table S1 for details.

composition, making up about half of their dry mass in some species, while the lipid content is usually very low (<10% dry mass). Based on the chemical composition, the preferential approach with macroalgae has therefore been to produce biogas by anaerobic digestion or bioethanol by fermentation. In contrast, the major components of microalgae are usually protein and lipid (Figure 1B). The lipid content of microalgae ranges from 1 to 77% of the dry mass and a level of 20–40% is quite common. The high lipid content in microalgae indicates that extracting biodiesel would be the most appropriate for biofuel production.

Despite species and environmental differences, the reasons for a substantially different chemical composition between macroalgae and microalgae are many. Firstly, macroalgae have substantially more compounds with a mechanical function to support the thallus structure (especially carbohydrates such as phycocolloids) leading to a relatively low abundance of lipids (Lee et al. 2017, Rhein-Knudsen et al. 2017). Secondly, microalgae have a larger proportion of chloroplast and thylakoid membranes compared to macroalgae. This is because microalgae are unicellular with little functional differentiation, leading to a larger proportion of photosynthetic membranes over the total cell volume (Han et al. 2003, Gao et al. 2017b). Phospholipid is the main component of cell membranes and a larger proportion of cell membranes in microalgae leads to a higher lipid content (Mooy et al. 2009, Yu et al. 2018). Thirdly, microalgae have a higher percentage of cell membrane by weight and hence a higher lipid content because of their higher surface to volume ratio (Hein et al. 2014). Finally, many microalgal species may have proportionately more lipids to increase their buoyancy and decrease their settling (Khanam et al. 2017, Pančić and Kjørboe 2018).

Biomass production

High biomass productivity is also an important factor when searching for potential bio-renewable resources. Algae possess remarkable advantages over terrestrial plants in terms of their biomass productivity since they grow in aquatic environments, saving land space and offering the potential for continuous cultivation (Jung et al. 2016). In addition, algae have a higher conversion efficiency of light energy to biomass compared to terrestrial plants, up to 5–10% vs. 0.5–3% (Wassink 1959, Laws et al. 1986, 1988, Melis 2009). This could be due to their fast growth with lower photorespiratory carbon loss because most algae operate CO₂ concentrating mechanisms to facilitate

carboxylation and suppress photorespiration (Melis 2009, Stephenson et al. 2011).

In addition to the culture conditions, the culture systems determine, to a large extent, the algal biomass yield. Daily biomass productivity of most macroalgae in tank culture is commonly not more than 30 g DM m⁻² land d⁻¹ (Figure 2A and Supplementary Table S2), which is comparable to microalgal productivity in open ponds (OP) and horizontal photobioreactors (HPB) but lower than in inclined photobioreactors (IPB; Figure 2B). Annual biomass productivity of macroalgae in nearshore farms is usually lower than 60 t DM ha⁻¹ land year⁻¹, and lower than that in tank culture (Figure 2C). Tank culture of *Ulva* species could reach higher biomass productivity (138 t DM ha⁻¹ land year⁻¹) but is still lower than the maximum from microalgae (Figure 2C, D). Microalgae are usually single-celled organisms, with a diameter of 1–200 μm (Madhu et al. 2017), while macroalgae, in most life stages, are more than 1 cm in length, and some species such as giant kelp can reach up to 90 m in length (Setchell 1908). The smaller microalgae have a higher surface to volume ratio and as a result can benefit from faster access to light and nutrients (Hein et al. 2014). In addition, microalgae do not have nonproductive cells, and all cells of microalgae are identically productive, while some tissues in the basal parts of macroalgae generally grow much slower than apical cells (Gao et al. 2017a). Due to their single-celled construction, microalgae do not utilize energy in producing structural biopolymers as backbones for their multicellular tissue. In addition, photobioreactors are more productive by providing optimal light and temperature conditions compared to ponds or near shore cultivation of macroalgae (Narala et al. 2016).

Biofuel production

Conversion of biomass to energy can be categorized into three main technologies: thermo-chemical, biochemical/biological and physical extraction (with esterification) as shown in Figure 3. Thermo-chemical conversion encompasses four process options: combustion, pyrolysis, liquefaction and gasification. There are two main process techniques in biochemical conversion: anaerobic digestion and ethanol fermentation. Several techniques have been utilized to produce biofuel from macroalgae; ethanol fermentation, anaerobic digestion and liquefaction will be briefly introduced in the following context.

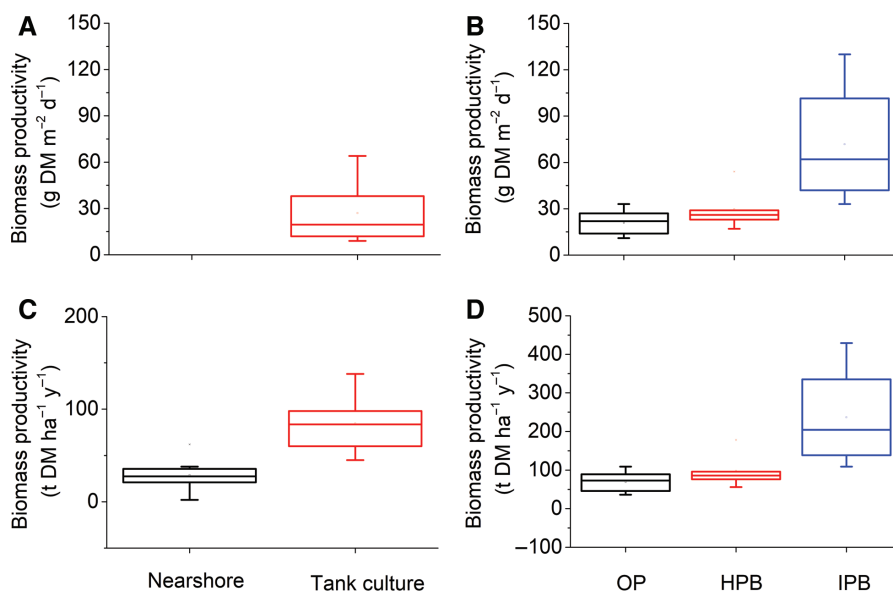


Figure 2: Box charts of biomass productivity of macroalgae (A, C) and microalgae (B, D) using different culture systems. OP, Open pond; HPB, horizontal photobioreactor; IPB, inclined photobioreactor; DM, dry mass. Data were based on Bidwell et al. (1985), Torzillo et al. (1986), Chaumont et al. (1988), Richmond et al. (1990), Lee and Low (1991), Richmond et al. (1993), Buschmann et al. (1994), Lee et al. (1995), Hu et al. (1996), Chynoweth (2002), Jiménez et al. (2003), Moreno et al. (2003), Doucha et al. (2005), Doucha and Lívanský (2006), Bruhn et al. (2011), Chen et al. (2013), Al-Hafedh et al. (2015), Correa et al. (2016), de Mooij et al. (2016), Mata et al. (2016), CFSY (2017), Benavides et al. (2017), Camus et al. (2018), Gao et al. (2018b), Romero-Villegas et al. (2018) and Magnusson et al. (2019). Please see Supplementary Table S2 for details.

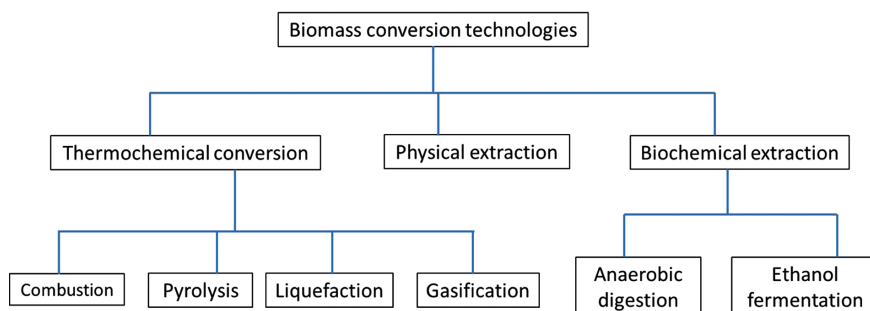


Figure 3: Technologies for conversion of biomass to energy.

Ethanol fermentation

Bioethanol from first generation feedstock, such as corn and sugarcane, has been the preferred choice as an automotive co-fuel and is now widely produced and used in many countries (Khuong et al. 2017). However, this first-generation bioethanol consumes a large amount of food crops and impacts food security. This has led to the development of second-generation bioethanol from lignocellulose biomass. Unfortunately, bioethanol from second-generation feedstock encountered huge resistance due to the difficulties in processing technology and scaling up (Gressel 2008, Ramachandra and Hebbale 2020). One

main technological challenge is that biodegradation of cellulose and hemicelluloses by cellulases can be inhibited by lignin (Gressel 2008, Ramachandra and Hebbale 2020). Macroalgae do not usually contain much lignin and therefore polysaccharides in macroalgae can be more easily converted to ethanol (Dave et al. 2019). Accordingly, macroalgae-derived bioethanol has been gaining increasing attention.

Bioethanol has been produced from all kinds of macroalgae, including brown, green and red macroalgae (Ramachandra and Hebbale 2020). Brown macroalgae seem to be the principal feedstock for bioethanol production due to their high polysaccharide content and

successful mass-cultivation (Jung et al. 2013, Enquist-Newman et al. 2014). For instance, ethanol production from brown macroalgal sugars by a synthetic yeast platform yields up to 83% of the theoretical maximum (Enquist-Newman et al. 2014), although ethanol conversion up to of 50% of the theoretical maximum from macroalgae is considered ambitious (Roesijadi et al. 2010a). Red macroalgae have also been fermented and yielded 45% of the theoretical maximum (Meinita et al. 2013). Bioethanol yields from the fermentation of macroalgae commonly range from 0.08 to 0.12 kg · kg⁻¹ dry mass. However, Wargacki et al. (2012) has reported higher experimental ethanol yields of up to 0.281 kg · kg⁻¹ dry mass from brown macroalgae by an engineered microbial platform. Different bioethanol yields can be attributed to species differences and processing methods. The bioethanol production from macroalgal biomass can be divided into two processes: pretreatment and microbial fermentation. The pretreatment step is very crucial and can determine the saccharide generation efficiency for efficient bioethanol production (Dave et al. 2019). The energy-intensive pretreatment leads to the failure of large-scale utilization of bioethanol from macroalgae to a large extent. Therefore, future work should pay more attention to developing a low-cost and scalable method for bioethanol production from macroalgae.

Anaerobic digestion

Anaerobic digestion (AD) technologies have a long history and the first industrial digestion plant was initiated in Bombay in 1859 (Kiyasudeen 2016). The first usage of biogas recovered from a sewage treatment facility was reported for street lamps in Exeter, England in 1895 (Kiyasudeen et al. 2016). Cellulosic materials such as dung and straw have been converted into methane for cooking in China for a long time (Buysman 2009). Anaerobic digestion is also an effective technique to treat sewage bio-solids, livestock manure, and concentrated wastes from the food industry and industrial wastewater (Nasir et al. 2012). Rising fossil fuel prices combined with increasing concerns for greenhouse gas emissions and global warming have prompted interest in further AD research and industrial applications. The total biogas production in the world has almost doubled from 1990 to 2016 (Energy Statistics Database 2019). AD occurs in four stages: hydrolysis, acidogenesis, acetogenesis and methanogenesis, among which, hydrolysis is the rate-limiting step of AD and also determines the final yield of biomethane to a great extent (Montingelli et al. 2016, Thompson et al. 2019).

Compared to biodiesel extraction and bioethanol fermentation, AD has a larger energy output because all macromolecular substances (protein, lipid and carbohydrate) can be utilized for biomethane production (Harun et al. 2011). Meanwhile, the energy input for biogas production from some crops is lower than that for bioethanol production, which results in a higher energy output-to-input ratio for AD (Börjesson and Mattiasson 2008). However, Patterson et al. (2008) demonstrated that the input energy for biomethane from sugar beet or fodder maize was higher than that for biodiesel from oilseed rape and bioethanol from wheat grain due to extra energy requirement for gas upgrading and compression. Another main problem for AD of algae is that biomethane yields from many algae are substantially below the theoretical maximum. Typical methane yields of seaweeds (~200 l CH₄ kg⁻¹ volatile solids, VS) are less than 50% of those from common commercially exploited feedstocks (Astals et al. 2015, Chen et al. 2015) although an extremely high yield (480 l CH₄ kg⁻¹ VS) was reported from a mixture of *Ulva*, *Cladophora* and *Chaetomorpha* species (Hansson 1983). The relatively low yield is related to high nitrogen, sulfur and salt contents of algae that can lead to potential inhibition for biomethanation (Tedesco et al. 2014). Therefore, there is noticeable room to improve biomethane yield by developing robust AD technology.

Liquefaction

Liquefaction used to involve employing carrier gases such as hydrogen or carbon monoxide to produce liquid fuels from solid material at moderate temperature (573–673 K) and high pressure, and it was initially used for coal liquefaction (Balat 2008). Liquefaction now refers to any thermochemical conversion process that primarily yields liquid products (Balat 2008, Ghadiryanfar et al. 2016). Liquefaction can process materials with any level of moisture content. Thus, this method is particularly suitable for algae since they contain a high level of water. The water may play a positive role in the liquefaction process as the physical and chemical properties of water change when heated near or above the critical point (374°C, 221 bar). The solubility of nonpolar hydrocarbons increases and, therefore, the decomposition of the biomass is improved (Kruse and Dinjus 2007). Current liquefaction processes involve high temperature and high pressure liquefaction, direct catalyst liquefaction, and supercritical liquefaction (Zhuang et al. 2012, Raikova et al. 2019).

Due to their high alkali content, macroalgae are suitable for liquefaction because high alkali has been

showed to have a catalytic function on bio-oil production during liquefaction (Anastasakis and Ross 2015). Many macroalgal species in all groups have been examined for bio-crude production via liquefaction and, up to now, the most intensively studied macroalgal species have been from the genera *Laminaria* and *Ulva* (Raikova et al. 2019). A maximum bio-oil yield of 19.3% dry weight with a higher heating value (HHV) of 36.5 MJ kg⁻¹ was obtained from *Laminaria saccharina* Linnaeus at 350°C without the presence of the catalyst (Anastasakis and Ross 2011). Zhou et al. (2010) used direct catalyst liquefaction to recover bio-oil from *Ulva prolifera* OF Müller and a maximum yield of bio-crude of 23% dry weight was achieved at 300°C with 5% Na₂CO₃ (w/w) by hydrothermal liquefaction and HHV of bio-oils were approximately 29 MJ kg⁻¹. The yield of bio-oil from *U. prolifera* OF Müller was significantly improved up to 84.81% with heating value of 15.05 MJ kg⁻¹ via microwave-assisted direct liquefaction and it could be further enhanced to 93.17% with HHV of 17.36 MJ kg⁻¹ by optimizing conditions (Zhuang et al. 2012, Liu et al. 2013). Although hydrothermal liquefaction has higher energy output and lower energy input for processing algae compared to gasification and pyrolysis (Vardon et al. 2012, Chen et al. 2014), it is likely that energy requirements for upgrading bio-crudes to usable biofuels would be high because bio-crudes have a higher concentration of heteroatoms oxygen, nitrogen, and sulfur (Mathimani et al. 2019). Few studies have been conducted on upgrading macroalgal bio-crudes to date, and therefore more work should be conducted in the future to improve algal liquefaction.

A comparison of several of the most used technologies for processing biofuels from algae is shown in Table 1. Here we compare methane from macroalgae with biodiesel from microalgae in terms of energy production and cost, since these two methods have been considered to have the

potential to compete with fossil diesel (Milledge et al. 2014, Dickinson et al. 2017, Wu et al. 2019). The biofuel yields from macroalgae and microalgae were 157–480 m³ CH₄ tonne⁻¹ DM and 85–360 kg biodiesel tonne⁻¹ DM, respectively; when converted to energy yield, they were 6.3–19.2 and 3.2–13.6 MJ tonne⁻¹ DM, indicating that macroalgae have a higher energy yield when normalized to dry mass (Figure 4A, B and Supplementary Table S3). Energy productivity (excluding the energy put into biofuel production) from macroalgae is in the range 96–677 GJ ha⁻¹ year⁻¹, which is again comparable to that from microalgae cultured in open ponds but lower than that from those cultured in photobioreactors (Figure 4C, D). This is due mainly to the high biomass productivity in photobioreactors.

Economics of biofuels from algae

Cost-effectiveness has to be considered when algae are used for the production of biofuels. The main costs can be generally split into two parts: cultivation and processing. The cultivation cost of macroalgae can be lower than \$100 t⁻¹ dry mass in Asian countries where the labor cost is low (Chynoweth 2002, Roesijadi et al. 2010b). The cultivation cost of microalgae cultured in open ponds (\$220–5940 t⁻¹ dry mass) is usually lower than for photobioreactors (\$430–7152 t⁻¹ dry mass; Figure 5A). The higher cultivation cost for microalgae is determined by the culture methods. Large scale microalgal cultivation is conducted in open ponds or photobioreactors, in which the supply of water, CO₂ and nutrients is required and mixed using paddle wheels, which also require energy input, which is costly (Hoffman et al. 2017). In addition, the photobioreactor tube system itself represents a significant capital investment accounting for around 80% of the total capital cost (Davis et al. 2011), leading to higher cultivation costs

Table 1: Comparison of methods used for converting algal biomass to biofuels.

Methods	Principle	Advantage	Disadvantage
Anaerobic digestion	Organic matters $\xrightarrow{\text{Anaerobic bacteria}}$ CH ₄ + CO ₂ + H ₂ + NH ₃ + H ₂ S	1. Utilize the whole algal cell 2. High energy output 3. Do not need drying	1. Lack robustness at industrial scale 2. Products need to purify and compress
Ethanol fermentation	(CH ₂ O) _n $\xrightarrow{\text{Yeast}}$ C ₂ H ₅ OH + CO ₂	1. Do not need drying 2. Product is easy to collect	1. Pretreatment is energy-intensive 2. Only utilize polysaccharides 3. Low energy output
Transesterification/ Biodiesel	—————→	1. Product is easy to extract 2. Product is easy to collect	1. Only apply to algae with high lipid content 2. Biomass needs drying
Liquefaction	Organic matters $\xrightarrow{\text{High T \& pressure}}$ Bio-oil + biochar + gas	1. Utilize the whole algal cell 2. Relatively high energy output 3. Do not need drying	1. Products need to be upgraded 2. High energy input

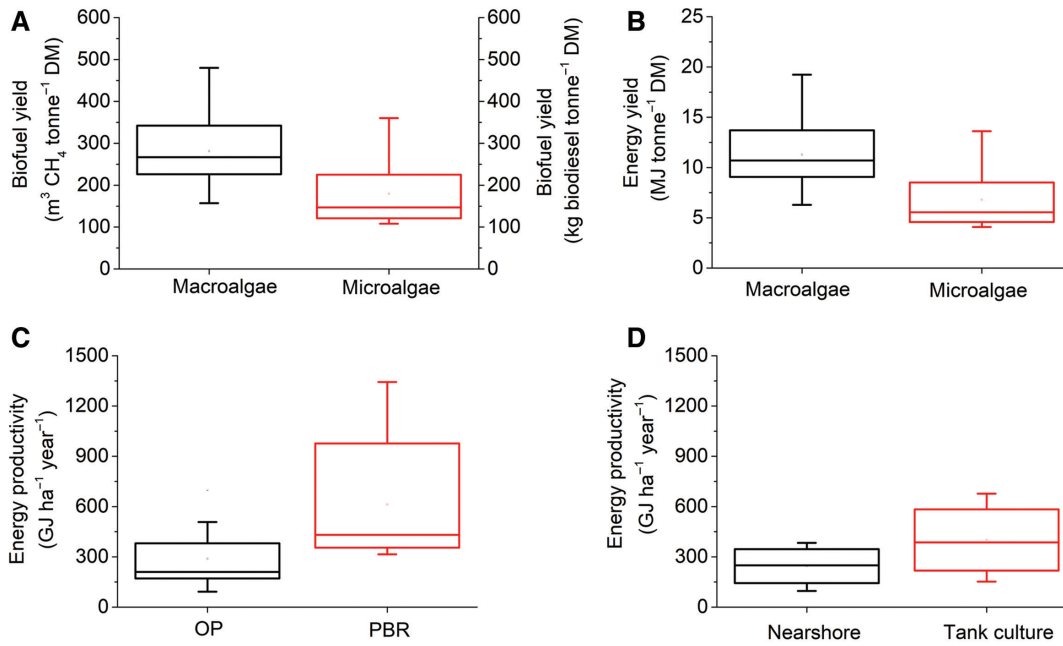


Figure 4: Box charts of yield of biofuel (A) and energy (B) and of gross energy productivity of macroalgae (C) and microalgae (D) in different culture systems.

OP, Open pond; PBR, photobioreactor; DM, dry mass. Data were based on Hansson (1983), Bruhn et al. (2011), Nascimento et al. (2014), Abomohra et al. (2016), Matsumoto et al. (2017), Tabassum et al. (2017), and Gao et al. (2018b). Please see Supplementary Table S3 for details.

compared to open ponds. In contrast, nearshore macroalgal cultivation takes advantage of natural seawater, CO₂, nutrients and mixing, with lower capital costs. Culture systems for macroalgal farming are relatively simple and cheap. In addition, the costs of harvesting and dewatering are more for microalgae compared to macroalgae (Aitken et al. 2014, Fasaie et al. 2018). Processing costs (Figure 5B) are also lower for macroalgae (\$8–18 t⁻¹ dry mass) compared to microalgae (\$74–183 t⁻¹ dry mass) as shown in Supplementary Table S4. This may be due to the chemicals used in oil extraction while aerobic digestion for biogas production from macroalgae does not consume additional chemicals.

To estimate what biomass productivity and methane yield in macroalgae, or lipid content in microalgae, is required to compete with fossil fuels, we have modeled the costs using the data of Dave et al. (2013) to define the relationship between the minimum electricity selling price (MESP), biomass productivity and methane yield for macroalgae, and the data of Davis et al. (2011) and Batan et al. (2016) to define the relationships between the minimum fuel selling price (MFSP), biomass productivity and lipid content for microalgae. This model excludes co-products, as there is currently no successful commercial operation that combines biofuels and co-products. The price of co-products is also highly variable and was

therefore excluded from this model. The equations are based on Sen et al. (2012):

$$\text{MESP (or MFSP)} = \frac{\text{OC} + \text{ROI} + \text{IT}}{\text{P} \times \text{Y} \times \text{A} \times \text{C}} \quad (1)$$

$$\text{ROI} = \frac{\text{DR} \times (1 + \text{DR})^{\text{ELS}}}{(1 + \text{DR})^{\text{ELS}} - 1} \times \text{TPI} \quad (2)$$

$$\text{IT} = \text{TR} \times (\text{BR} - \text{OC}) \quad (3)$$

where OC is operating costs, ROI is return on investment, IT is income tax, P is biomass productivity (t ha⁻¹ year⁻¹), Y is lipid content (microalgae) or biomethane yield (macroalgae) (m³ t⁻¹), A is cultivation area, C is conversion coefficient (80% for algal lipid to biodiesel; 40% for CH₄ to electricity), DR is discount rate (10%), ELS is equipment life span (20 years), TPI is total project investment, TR is tax rate (35%), and BR is biofuel revenue.

To make MESP equivalent to the current electricity price of \$0.1043 kWh⁻¹, the biomass productivity for macroalgae should be 73.5 t ha⁻¹ year⁻¹, with a methane yield of 285 m³ t⁻¹ dry mass (Figure 6). Based on the previous studies (Figures 2 and 4), it seems possible for some species of *Ulva* to achieve this when cultivated in tanks. It is worth noting that this model is based on the cost of nearshore

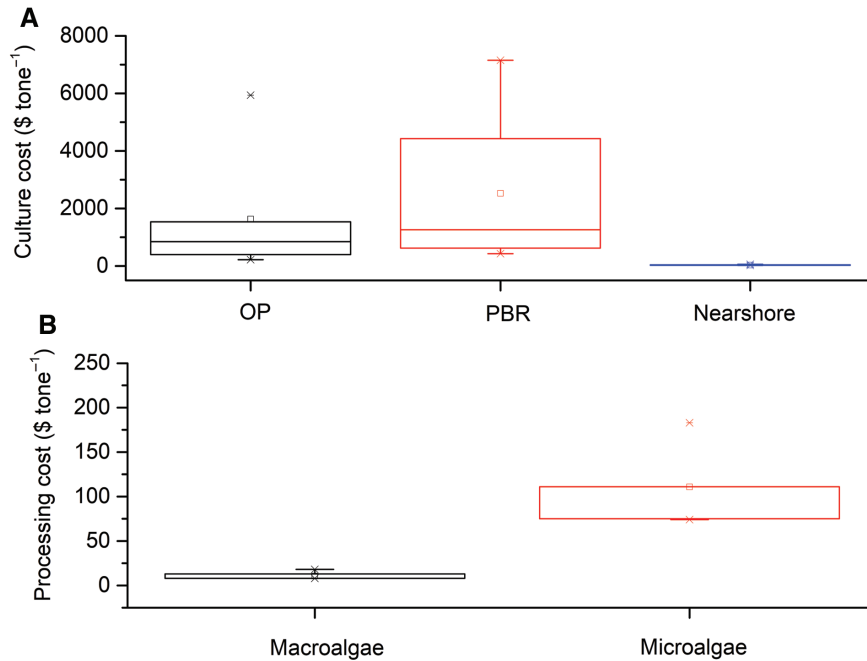


Figure 5: Box charts of culture (A) and processing costs (B) for biofuel production from macroalgae and microalgae in different culture systems.

OP, Open pond; PBR, photobioreactor. Data were based on Roesijadi et al. (2010b), Norsker et al. (2011), Richardson et al. (2012), Nagarajan et al. (2013), Davis et al. (2014), Dave et al. (2013), Hoffman et al. (2017), Soleymani and Rosentrater (2017). Please see Supplementary Table S4 for details.

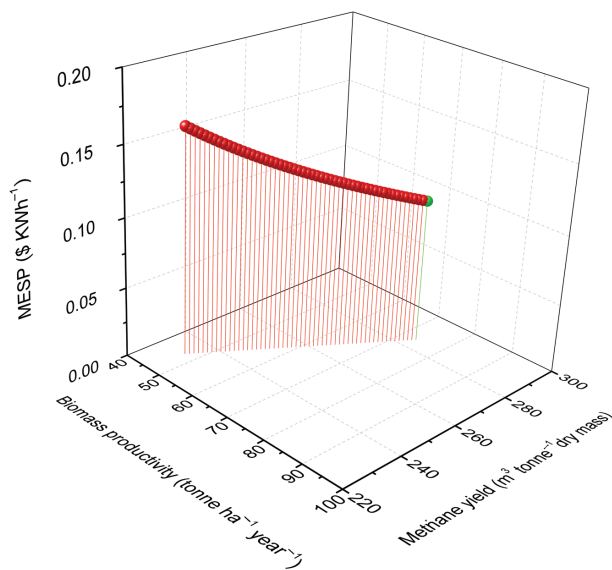


Figure 6: Effects of biomass productivity and methane yield of macroalgae on minimum electricity selling price (MESP). Green circle indicates current electricity price. MESP calculation was based on data from Dave et al. (2013) using equation from Sen et al. (2012) with assumptions that discount rate is 10%, equipment life span is 20 years and tax rate is 35%.

systems. Until now, the highest biomass productivity in nearshore systems is 62 t ha⁻¹ year⁻¹ for *Macrocystis pyrifera* (Linnaeus) C. Agardh (Supplementary Table S2).

Therefore, biomass productivity in nearshore systems needs to be improved to make MESP equal to the current electricity price.

The MFSP for microalgae cultured in photobioreactors (\$8.36 l⁻¹) is much higher than in open ponds (\$3.88 l⁻¹). To compete with diesel from fossil fuel (currently \$0.82 l⁻¹), the biomass productivity and lipid content for microalgae cultured in open ponds must be more than 61.8 g DM m⁻² day⁻¹ and 71% (Figure 7A). From the previous studies (Figures 1 and 2), this seems very difficult for microalgae cultured in open ponds. For microalgae cultured in photobioreactors to compete with diesel from fossil fuel, even higher biomass productivity (232 g DM m⁻² d⁻¹) and lipid content (80.2%) are required (Figure 7B). Crucially, this is well above the theoretical maximum microalgal productivity of 196 g m⁻² day⁻¹ (Weyer et al. 2010). The target lipid content of 80.2% also seems currently impossible.

To optimize culture conditions for microalgae so that both high biomass yield and high lipid yield are achieved is always a challenge because the optimal culture conditions for cell growth and lipid production are not the same. Accordingly, a two-stage culture method is proposed to resolve this problem, in which growth and lipid production are split into separate phases. This culture system has been proven successful in *Nannochloropsis oculata* (Droop) D. J. Hibberd (Aléman-Nava et al. 2017), *Chlorella* sp. (Nayak

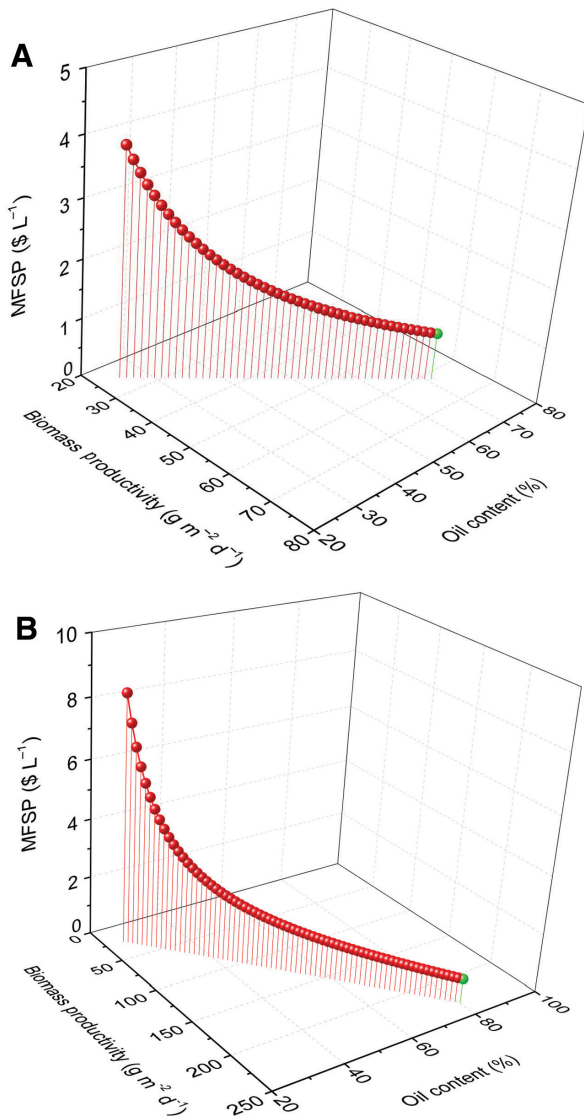


Figure 7: Effects of biomass productivity and lipid content of microalgae cultivated in open pond (A) and photobioreactor (B) on minimum fuel selling price (MFSP).

Green circle indicates current diesel price from fossil fuel. MFSP calculation based on data from Davis et al. (2011) using equation from Sen et al. (2012) with assumptions that discount rate is 10%, equipment life span is 20 years and tax rate is 35%.

et al. 2019) and *Skeletonema costatum* (Greville) Cleve (Gao et al. 2019). Currently, the culture cost is the main obstacle for biodiesel from microalgae to achieve a positive energy balance. Because of the difficulty of near shore or off shore cultivation, it would be difficult to reduce the farming costs for microalgae. Although genetic engineering approaches have been used to improve lipid content in microalgae without significantly affecting biomass yield, commercially producing biodiesel from microalgae remains difficult without subsidies (Sheridan 2013, Ajjawi

et al. 2017, Posewitz 2017, Remmers et al. 2018). In addition, de Boer et al. (2012) proposed several pathways to reduce the cost of biofuel from microalgae, while more research is still required to increase the efficiency of these pathways and to apply them at commercial scale. It seems that commercially viable biofuel production from microalgae can only be possible if the other (more valuable) constituents of the algal biomass are exploited as co-products (Borowitzka 2013, Foteinis et al. 2018).

Conclusion

Compared to microalgae, the biomass productivity of macroalgae is lower. However, the lower cultivation and processing costs make the production of biomethane from macroalgae very close to profitability. The main reason for the lower annual biomass productivity of most macroalgae is their periodic reproduction (Wei et al. 2013, Gao et al. 2017c). This is particularly obvious in summer when most macroalgae have to survive via the form of microscopic propagules. Therefore, to obtain a stable and high biomass yield, it would be beneficial to obtain some species without reproduction to improve the biomass productivity (Gao et al. 2017c) and some strains which are tolerant of high temperatures. Compared to *Porphyra* and *Laminaria*, *Ulva* has higher tolerance to high environmental pressures, making it feasible for year-round cultivation (Carl et al. 2016, Gao et al. 2016a,b). In addition, the biomass of *Ulva* can be directly collected from green tides that commonly occur in eutrophic waters worldwide (Ye et al. 2011, Paumier et al. 2018), which can further reduce the cost of the biomass. For instance, around 20 million tons (fresh weight) of *Ulva* biomass were produced by the green tide occurring in the Yellow Sea of China in 2008 (Gao et al. 2010, Ye et al. 2011). In addition to green tides, another type of macroalgal bloom termed golden tides is also on the rise (Smetacek and Zingone 2013, Milledge and Harvey 2016, Xu et al. 2017). The coverage area of the golden tide in 2017 even exceeded that of most of the green tides in the Yellow Sea during past 10 years (Qi et al. 2017). Although there are some studies on extracting biofuels from *Sargassum* (Li et al. 2012, Borines et al. 2013, Soto et al. 2015), biofuel production from *Sargassum* is still in its infancy. Using bloom-forming macroalgae as biofuel should be given priority because of their high growth rate and direct availability from the field. In addition, to develop engineered microbial platforms is an effective approach to improve biofuel yield from macroalgae (Wargacki et al. 2012, Enquist-Newman et al. 2014, Camus et al. 2016).

The lower growth rate of macroalgae compared to microalgae means that larger areas are required to produce the same biomass. For instance, to meet the global natural gas demand (3848.9 bcm) in 2018 (Dudley 2019), an area of 315 million ha is needed to culture *Ulva lactuca* (Bruhn et al. 2011), which is equivalent to the whole ocean area of China. In addition, there is a possible future shortfall in phosphate supplies globally which would adversely affect the ability to fertilize mass cultures (Raven 2017, Gao et al. 2018b). Meanwhile, macroalgae are ideal materials for wastewater bioremediation because of their strong capacity to absorb nutrients and heavy metals (Gao et al. 2018c, Nardelli et al. 2019). To use bioremediating macroalgae for biofuel is thus a way to reduce the cultivation area and add ecological services. Therefore, integrated farming of macroalgae and other commercial marine animals needs further development as it can enhance the productivity of both macroalgae and animals (Pedra et al. 2017, Gao et al. 2018c, Laramore et al. 2018) as well as reducing eutrophication.

Acknowledgements: This study was supported by the National key R&D program of China (2016YFA0601400), the Lianyungang Innovative and Entrepreneurial Doctor Program (201702), the Jiangsu Planned Projects for Postdoctoral Research Funds (1701003A), the Postgraduate Research & Practice Innovation Program of Jiangsu Province (SJCX19_0951), and the Priority Academic Program Development of Jiangsu Higher Education Institutions of China.

References

- Abomohra, A., W. Jin, R. Tu and S. Han. 2016. Outdoor cultivation of the biodiesel promising microalga *Scenedesmus obliquus* in municipal wastewater: a case study. *Renew. Energ. Res.* 1: 17–23.
- Abomohra, A.E.-F., A.H. El-Naggar and A.A. Baeshen. 2018. Potential of macroalgae for biodiesel production: Screening and evaluation studies. *J. Biosci. Bioeng.* 125: 231–237.
- Adeniyi, O.M., U. Azimov and A. Burluka. 2018. Algae biofuel: current status and future applications. *Renew. Sust. Energy Rev.* 90: 316–335.
- Aitken, D., C. Bulboa, A. Godoyfaundez, J.L. Turrion Gomez and B. Antizarladislao. 2014. Life cycle assessment of macroalgae cultivation and processing for biofuel production. *J. Clean. Prod.* 75: 45–56.
- Ajjawi, I., J. Verruto, M. Aquí, L.B. Soriaga, J. Coppersmith, K. Kwok, L. Peach, E. Orchard, R. Kalb and W. Xu. 2017. Lipid production in *Nannochloropsis gaditana* is doubled by decreasing expression of a single transcriptional regulator. *Nat. Biotechnol.* 35: 647–655.
- Aléman-Nava, G.S., K. Muylaert, S.P.C. Bermudez, O. Depraetere, B. Rittmann, R. Parra-Saldívar and D. Vandamme. 2017. Two-stage cultivation of *Nannochloropsis oculata* for lipid production using reversible alkaline flocculation. *Bioresour. Technol.* 226: 18–23.
- Al-Hafedh, Y.S., A. Alam and A.H. Buschmann. 2015. Bioremediation potential, growth and biomass yield of the green seaweed, *Ulva lactuca* in an integrated marine aquaculture system at the Red Sea coast of Saudi Arabia at different stocking densities and effluent flow rates. *Rev. Aquacult.* 7: 161–171.
- Anastasakis, K and A.B. Ross. 2011. Hydrothermal liquefaction of the brown macro-alga *Laminaria Saccharina*: Effect of reaction conditions on product distribution and composition. *Bioresour. Technol.* 102: 4876–4883.
- Anastasakis, K and A.B. Ross. 2015. Hydrothermal liquefaction of four brown macro-algae commonly found on the UK coasts: an energetic analysis of the process and comparison with biochemical conversion methods. *Fuel* 139: 139546–139553.
- Astals, S., R.S. Musenze, X. Bai, S. Tannock, S. Tait, S. Pratt and P.D. Jensen. 2015. Anaerobic co-digestion of pig manure and algae: impact of intracellular algal products recovery on co-digestion performance. *Bioresour. Technol.* 181: 97–104.
- Balat, M. 2008. Mechanisms of thermochemical biomass conversion processes. Part 3: reactions of liquefaction. *Energy Sources Part A* 30: 649–659.
- Batan, L.Y., G.D. Graff and T.H. Bradley. 2016. Techno-economic and Monte Carlo probabilistic analysis of microalgae biofuel production system. *Bioresour. Technol.* 219: 45–52.
- Benavides, A.M.S., K. Ranglová, J.R. Malapascua, J. Masojádek and G. Torzillo. 2017. Diurnal changes of photosynthesis and growth of *Arthrospira platensis* cultured in a thin-layer cascade and an open pond. *Algal Res.* 28: 48–56.
- Bidwell, R.G.S., J. McLachlan and N.D.H. Lloyd. 1985. Tank cultivation of Irish moss, *Chondrus crispus* Stackh. *Bot. Mar.* 28: 87–98.
- Borines, M.G., R.L. de Leon, and J.L. Cuello. 2013. Bioethanol production from the macroalgae *Sargassum* spp. *Bioresour. Technol.* 138: 22–29.
- Borowitzka, M.A. 2013. High-value products from microalgae – their development and commercialisation. *J. Appl. Phycol.* 25: 743–756.
- Borowitzka, M.A. and A. Vonshak. 2017. Scaling up microalgal cultures to commercial scale. *Eur. J. Phycol.* 52: 407–418.
- Börjesson, P. and B. Mattiasson. 2008. Biogas as a resource-efficient vehicle fuel. *Trends Biotechnol.* 26: 7–13.
- Bruhn, A., J. Dahl, H.B. Nielsen, L. Nikolaisen, M.B. Rasmussen, S. Markager, B. Olesen, C. Arias and P.D. Jensen. 2011. Bioenergy potential of *Ulva lactuca*: Biomass yield, methane production and combustion. *Bioresour. Technol.* 102: 2595–2604.
- Buschmann, A.H., O.A. Mora, P. Gómez, M. Böttger, S. Buitano, C. Retamales, P.A. Vergara and A. Gutierrez. 1994. *Gracilaria chilensis* outdoor tank cultivation in Chile: use of land-based salmon culture effluents. *Aquacult. Eng.* 13: 283–300.
- Buysman, E. 2009. *Anaerobic digestion for developing countries with cold climates*. Master thesis, Faculty of Environmental Sciences, University of Wageningen, Netherlands.
- Swapwa, J.K., A. Anandraj and C. Trois. 2017. Possibilities for conversion of microalgae oil into aviation fuel: a review. *Renew. Sust. Energy. Rev.* 80: 1345–1354.
- Camus, C., P. Ballerino, R. Delgado, Á. Olivera-Nappa, C. Leyton and A.H. Buschmann. 2016. Scaling up bioethanol production from

- the farmed brown macroalga *Macrocystis pyrifera* in Chile. *Biofuel. Bioprod. Bior.* 10: 673–685.
- Camus, C., J. Infante, and A.H. Buschmann. 2018. Overview of 3 year precommercial seafarming of *Macrocystis pyrifera* along the Chilean coast. *Rev. Aquacult.* 10: 543–559.
- Carl, C., R.J. Lawton, N.A. Paul and R. de Nys. 2016. Reproductive output and productivity of filamentous tropical *Ulva* over time. *J. Appl. Phycol.* 28: 429–438.
- Chan, P.T. and P. Matanjun. 2017. Chemical composition and physicochemical properties of tropical red seaweed, *Gracilaria changii*. *Food Chem.* 221: 302–310.
- Chaumont, D., C. Thepenier, C. Gudín and C. Junjas. 1988. Scaling up a tubular photoreactor for continuous culture of *Porphyridium cruentum* from laboratory to pilot plant (1981–1987). In: (T. Stadler, J. Mollion M.-C. Verdus, Y. Karamanos, H. Morvan and D. Christiaen, eds.) *Algal biotechnology*. Elsevier, New York. pp. 199–208.
- Chen, Y., J. Wang, W. Zhang, L. Chen, L. Gao and T. Liu. 2013. Forced light/dark circulation operation of open pond for microalgae cultivation. *Biomass Bioenergy* 56: 464–470.
- Chen, W.T., Y. Zhang, J. Zhang, G. Yu, L.C. Schideman, P. Zhang and M. Minarick. 2014. Hydrothermal liquefaction of mixed-culture algal biomass from wastewater treatment system into bio-crude oil. *Bioresour. Technol.* 152: 130–139.
- Chen, H., D. Zhou, G. Luo, S. Zhang and J. Chen. 2015. Macroalgae for biofuels production: Progress and perspectives. *Renew. Sustain. Energy Rev.* 47: 427–437.
- Cheng, F., Z. Cui, L. Chen, J. Jarvis, N. Paz, T. Schaub, N. Nirmalakhandan and C.E. Brewer. 2017. Hydrothermal liquefaction of high- and low-lipid algae: bio-crude oil chemistry. *Appl. Energy* 206: 278–292.
- China Fishery Statistical Yearbook (CFSY). 2017. China Agriculture Press, Beijing.
- Chisti, Y. 2007. Biodiesel from microalgae. *Biotechnol. Adv.* 25: 294–306.
- Correa, T., A. Gutiérrez, R. Flores, A.H. Buschmann, P. Cornejo and C. Bucarey. 2016. Production and economic assessment of giant kelp *Macrocystis pyrifera* cultivation for abalone feed in the south of Chile. *Aquac. Res.* 47: 698–707.
- Chung, I.K., J. Beardall, S. Mehta, D. Sahoo and S. Stojkovic. 2011. Using marine macroalgae for carbon sequestration: a critical appraisal. *J. Appl. Phycol.* 23: 877–886.
- Chynoweth, D.P. 2002. Review of biomethane from marine biomass. History, results and conclusions of the “US Marine Biomass Energy Program” (1968–1990) Gainesville: Department of Agricultural and Biological Engineering, University of Florida. pp. 194.
- Duarte, C.M., I.J. Losada, I.E. Hendriks, I. Mazarrosa and N. Marbà. 2013. The role of coastal plant communities for climate change mitigation and adaptation. *Nat. Clim. Change* 3: 961–968.
- Duarte, C.M., J. Wu, X. Cáo, A. Bruin and D. Krause-Jensen. 2017. Can seaweed farming play a role in climate mitigation and adaptation? *Front. Mar. Sci.* 4: 100.
- Dudley, B. 2018. *BP statistical review of world energy*. BP Statistical Review London, UK.
- Dudley, B. 2019. *BP statistical review of world energy*. BP Statistical Review London, UK.
- Dave, A., Y. Huang, S. Rezvani, D. McIlveen-Wright, M. Novaes and N. Hewitt. 2013. Techno-economic assessment of biofuel development by anaerobic digestion of European arine cold-water seaweeds. *Bioresour. Technol.* 135: 120–127.
- Dave, N., R. Selvaraj, T. Varadavenkatesan and R. Vinayagam. 2019. A critical review on production of bioethanol from macroalgal biomass. *Algal Res.* 42: 101606.
- Davis, R., A. Aden and P.T. Pienkos. 2011. Techno-economic analysis of autotrophic microalgae for fuel production. *Appl. Energy* 88: 3524–3531.
- Davis, R., C. Kinchin, J. Markham, E. Tan, L. Laurens, D. Sexton, D. Knorr, P. Schoen and J. Lukas. 2014. Process design and economics for the conversion of algal biomass to biofuels: algal biomass fractionation to lipid- and carbohydrate-derived fuel products. Golden, CO, USA
- Demirbas, A. 2010. Use of algae as biofuel sources. *Energy Convers. Manage.* 51: 2738–2749.
- de Boer, K., N.R. Moheimani, M.A. Borowitzka and P.A. Bahri. 2012. Extraction and conversion pathways for microalgae to biodiesel: a review focused on energy consumption. *J. Appl. Phycol.* 24: 1681–1698.
- de Mooij, T., G. de Vries, C. Latsos, R.H. Wijffels and M. Janssen. 2016. Impact of light color on photobioreactor productivity. *Algal Res.* 15: 32–42.
- Dickinson, S., M. Mientus, D. Frey, A. Amini-Hajbashi, S. Ozturk, F. Shaikh, D. Sengupta and M.M. El-Halwagi. 2017. A review of biodiesel production from microalgae. *Clean Technol. Environ.* 19: 637–668.
- Diprat, A.B., T. Menegol, J.F. Boelter, A. Zmozinski, M.G. Rodrigues Vale, E. Rodrigues and R. Rech. 2017. Chemical composition of microalgae *Heterochlorella luteoviridis* and *Dunaliella tertiolecta* with emphasis on carotenoids. *J. Sci. Food Agric.* 97: 3463–3468.
- Doucha, J. and K. Lívanský. 2006. Productivity, CO₂/O₂ exchange and hydraulics in outdoor open high density microalgal (*Chlorella* sp.) photobioreactors operated in a Middle and Southern European climate. *J. Appl. Phycol.* 18: 811–826.
- Doucha, J., F. Straka and K. Lívanský. 2005. Utilization of flue gas for cultivation of microalgae *Chlorella* sp.) in an outdoor open thin-layer photobioreactor. *J. Appl. Phycol.* 17: 403–412.
- El-Mashad, H.M. 2015. Biomethane and ethanol production potential of *Spirulina platensis* algae and enzymatically saccharified switchgrass. *Biochem. Eng. J.* 93: 119–127.
- Energy Statistics Database – United Nations Statistics Division, 2019.
- Enquist-Newman, M., A.M.E. Faust, D.D. Bravo, C.N.S. Santos, R.M. Rainsner, A. Hanel, P. Sarvabhowman, C. Le, D.D. Regitsky, S.R. Cooper and L. Peereboom. 2014. Efficient ethanol production from brown macroalgae sugars by a synthetic yeast platform. *Nature* 505: 239–246.
- Fasaee, F., J.H. Bitter, P.M. Slegers and A.J.B.V. Boxtel. 2018. Techno-economic evaluation of microalgae harvesting and dewatering systems. *Algal Res.* 31: 347–362.
- Foteinis, S., A. Antoniadis-Gavriil and T. Tsoutsos. 2018. Life cycle assessment of algae-to-biodiesel shallow pond production systems in the Mediterranean: influence of species, pond type, by (co)-product valorisation and electricity mix. *Biofuel Bioprod Bior.* 12: 542–558.
- Gao, K. and K.R. McKinley. 1994. Use of macroalgae for marine biomass production and CO₂ remediation: a review. *J. Appl. Phycol.* 6: 45–60.

- Gao, G., Y. Liu, X. Li, Z. Feng, and J. Xu. 2016a. An ocean acidification acclimatised green tide alga is robust to changes of seawater carbon chemistry but vulnerable to light stress. *PLoS One* 11: 0169040.
- Gao, G., Z. Zhong, X. Zhou and J. Xu. 2016b. Changes in morphological plasticity of *Ulva prolifera* under different environmental conditions: a laboratory experiment. *Harmful Algae* 59: 51–58.
- Gao, G., A.S. Clare, C. Rose and G.S. Caldwell. 2017a. Eutrophication and warming-driven green tides (*Ulva rigida*) are predicted to increase under future climate change scenarios. *Mar. Pollut. Bull.* 114: 439–447.
- Gao, G., A.S. Clare, C. Rose and G.S. Caldwell. 2017b. Intrinsic and extrinsic control of reproduction in the green tide-forming alga, *Ulva rigida*. *Environ. Exp. Bot.* 139: 14–22.
- Gao, G., A.S. Clare, C. Rose and G.S. Caldwell. 2017c. Reproductive sterility increases the capacity to exploit the green seaweed *Ulva rigida* for commercial applications. *Algal Res.* 24: 64–71.
- Gao, G., A.S. Clare, E. Chatzidimitriou, C. Rose and G. Caldwell. 2018a. Effects of ocean warming and acidification, combined with nutrient enrichment, on chemical composition and functional properties of *Ulva rigida*. *Food Chem.* 258: 71–78.
- Gao, G., J. Beardall, M. Bao, C. Wang, W. Ren and J. Xu. 2018b. Ocean acidification and nutrient limitation synergistically reduce growth and photosynthetic performances of a green tide alga *Ulva linza*. *Biogeosciences* 15: 3409–3420.
- Gao, G., A.S. Clare, C. Rose and G.S. Caldwell. 2018c. *Ulva rigida* in the future ocean: potential for carbon capture, bioremediation, and biomethane production. *G. C. B. Bioenergy* 10: 39–51.
- Gao, G., M. Wu, Q. Fu, X. Li and J. Xu. 2019. A two-stage model with nitrogen and silicon limitation enhances lipid productivity and biodiesel features of the marine bloom-forming diatom *Skeletonema costatum*. *Bioresour. Technol.* 289: 121717.
- Gao, S., X. Chen, Q. Yi, G. Wang, G. Pan, A. Lin and G. Peng. 2010. A strategy for the proliferation of *Ulva prolifera*, main causative species of green tides, with formation of sporangia by fragmentation. *PLoS One* 5: e8571.
- Ghadiryfar, M., K.A. Rosentrater, A. Keyhani and M. Omid. 2016. A review of macroalgae production, with potential applications in biofuels and bioenergy. *Renew. Sustain. Energy Rev.* 54: 473–54481.
- Gressel, J. 2008. Transgenics are imperative for biofuel crops. *Plant Sci.* 174: 246–263.
- Han, T., Y.S. Han, J.M. Kain and D.P. Häder. 2003. Thallus differentiation of photosynthesis, growth, reproduction, and UV-B sensitivity in the green alga *Ulva pertusa* (Chlorophyceae). *J. Phycol.* 39: 712–721.
- Hansson, G. 1983. Methane production from marine, green macroalgae. *Resour. Conserv.* 8: 185–194.
- Harun, R., M. Davidson, M. Doyle, R. Gopiraj, M. Danquah and G. Forde. 2011. Technoeconomic analysis of an integrated microalgae photobioreactor, biodiesel and biogas production facility. *Biomass Bioenerg.* 35: 741–747.
- Hein, M., M. Pedersen and K. Sand-Jensen. 2014. Size-dependent nitrogen uptake in micro- and macroalgae. *Mar. Ecol. Prog. Ser.* 118: 247–253.
- Herrmann, R., C. Jumbe, M. Bruentrup and E. Osabuohien. 2018. Competition between biofuel feedstock and food production: Empirical evidence from sugarcane outgrower settings in Malawi. *Biomass Bioenerg.* 114: 100–111.
- Hoffman, J., R.C. Pate, T. Drennen and J.C. Quinn. 2017. Technoeconomic assessment of open microalgae production systems. *Algal Res.* 23: 51–57.
- Hong, I.K., H. Jeon and S.B. Lee. 2014. Comparison of red, brown and green seaweeds on enzymatic saccharification process. *J. Ind. Eng. Chem.* 20: 2687–2691.
- Horn, S.J. 2000. *Bioenergy from brown seaweeds*. PhD thesis. Department of Biotechnology, Norwegian University of Science and Technology.
- Hu, Q., H. Guterman and A. Richmond. 1996. A flat inclined modular photobioreactor for outdoor mass cultivation of photoautotrophs. *Biotechnol. Bioeng.* 51: 51–60.
- Ishika, T., P.A. Bahri, D.W. Laird and N.R. Moheimani. 2018. The effect of gradual increase in salinity on the biomass productivity and biochemical composition of several marine, halotolerant, and halophilic microalgae. *J. Appl. Phycol.* 30: 1453–1464.
- Jang, S.-S., Y. Shirai., M. Uchida and M. Wakisaka. 2012. Production of mono sugar from acid hydrolysis of seaweed. *Afr. J. Biotechnol.* 11: 1953–1963.
- Jiang, X., Q. Han, X. Gao and G. Gao. 2016. Conditions optimising on the yield of biomass, total lipid, and valuable fatty acids in two strains of *Skeletonema menzeli*. *Food Chem.* 194: 723–732.
- Jiménez, C., B.R. Cossío, D. Labela and F. Xavier Niell. 2003. The Feasibility of industrial production of *Spirulina (Arthrospira)* in Southern Spain. *Aquaculture* 217: 179–190.
- Jung, K.A., S.R. Lim, Y. Kim and J.M. Park. 2013. Potentials of macroalgae as feedstocks for biorefinery. *Bioresour. Technol.* 135: 182–190.
- Jung, K.A., S.R. Lim, Y. Kim and J.M. Park. 2016. Opportunity and challenge of seaweed bioethanol based on life cycle CO₂ assessment. *Environ. Prog. Sustain. Energy* 36: 200–207.
- Kalita, N., G. Baruah, R. Chandra, D. Goswami, J. Talukdar and M.C. Kalita. 2017. *Ankistrodesmus falcatus*: a promising candidate for lipid production, its biochemical analysis and strategies to enhance lipid productivity. *J. Microbiol. Biotechnol. Res.* 1: 148–157.
- Khanam, M.R.M., Y. Shimasaki, M.Z. Hosain, K. Mukai, M. Tsuyama, X. Qiu, R. Tasmin, H. Goto and Y. Oshima. 2017. Diuron causes sinking retardation and physiochemical alteration in marine diatoms *Thalassiosira pseudonana* and *Skeletonema marinoi-dohrnii* complex. *Chemosphere* 175: 200–209.
- Khatoun, H., N.A. Rahman, S. Banerjee, N. Harun, S.S. Suleiman, N.H. Zakaria, F. Lananan, S.H.A. Hamid and A. Endut. 2014. Effects of different salinities and pH on the growth and proximate composition of *Nannochloropsis* sp. and *Tetraselmis* sp. isolated from South China Sea cultured under control and natural condition. *Int. Biodeter. Biodegr.* 95: 11–18.
- Khuong, L.S., H.H. Masjuki, N.W.M. Zulkifli, E. Niza Mohamad, M.A. Kaiam, A. Alabdulkarem, A. Arslan, M.H. Mosarof, A.Z. Syahir and M. Jamshaid. 2017. Effect of gasoline–bioethanol blends on the properties and lubrication characteristics of commercial engine oil. *RSC Adv.* 7: 15005–15019.
- Kim, N.-J., H. Li, K. Jung, H.N. Chang and P.C. Lee. 2011. Ethanol production from marine algal hydrolysates using *Escherichia coli* KO11. *Bioresour. Technol.* 102: 7466–7469.
- Kiyasudeen, K., M.H. Ibrahim, S. Quaik, and S.A. Ismail. 2016. An introduction to anaerobic digestion of organic wastes. In: (K. Kiyasudeen, M.H. Ibrahim, S. Quaik and S.A. Ismail, eds.) *Prospects of organic waste management and the significance of earthworm*. Springer, Cham. pp. 23–44.

- Kraan, S. 2010. Mass-cultivation of carbohydrate rich macroalgae, a possible solution for sustainable biofuel production. *Mitig. Adapt. Strat. Gl.* 18: 27–46.
- Kruse, A. and E. Dinjus. 2007. Hot compressed water as reaction medium and reactant: properties and synthesis reactions. *J. Supercrit. Flu.* 39: 362–380.
- Landis, D.A., C. Gratton, R.D. Jackson, K.L. Gross, D.S. Duncan, C. Liang, T.D. Meehan, B.A. Robertson, T.M. Schmidt and K.A. Stahlheber. 2017. Biomass and biofuel crop effects on biodiversity and ecosystem services in the North Central US. *Biomass Bioenergy* 114: 18–29.
- Laramore, S., R. Baptiste and D.H. Wills P S. 2018. Utilization of IMTA-produced *Ulva lactuca* to supplement or partially replace pelleted diets in shrimp (*Litopenaeus vannamei*) reared in a clear water production system. *J. Appl. Phycol.* 30: 3603–3610.
- Laws, E.A., S. Taguchi, J. Hirata and L. Pang. 1986. High algal production rates achieved in a shallow outdoor flume. *Biotechnol. Bioeng.* 28: 191–197.
- Laws, E.A., S. Taguchi, J. Hirata and L. Pang. 1988. Optimization of microalgal production in a shallow outdoor flume. *Biotechnol. Bioeng.* 32: 140–147.
- Lee, Y.K. and C.S. Low. 1991. Effect of photobioreactor inclination on the biomass productivity of an outdoor algal culture. *Biotechnol. Bioeng.* 38: 995–1000.
- Lee, Y.K., S.Y. Ding, C.S. Low, Y.C. Chang, W.L. Forday and P.C. Chew. 1995. Design and performance of an α -type tubular photobioreactor for mass cultivation of microalgae. *J. Appl. Phycol.* 7: 47–51.
- Lee, W.K., Y.Y. Lim, A.T. Leow, P. Namasivayam, A.J. Ong and C.L. Ho. 2017. Biosynthesis of agar in red seaweeds: a review. *Carbohydr. Polym.* 164: 23–30.
- Li, D., L. Chen, D. Xu, X. Zhang, N. Ye, F. Chen, and S. Chen. 2012. Preparation and characteristics of bio-oil from the marine brown alga *Sargassum patens* C. Agardh. *Bioresour. Technol.* 104: 737–742.
- Liu, J., Y. Zhuang, Y. Li, L. Chen, J. Guo, D. Li and N. Ye. 2013. Optimizing the conditions for the microwave-assisted direct liquefaction of *Ulva prolifera* for bio-oil production using response surface methodology. *Energy* 60: 69–76.
- Madhu, N.V., G.D. Martin, C.K. Haridevi, M. Nair, K.K. Balachandran and N. Ullas. 2017. Differential environmental responses of tropical phytoplankton community in the southwest coast of India. *Reg. Stud. Mar. Sci.* 16: 21–35.
- Magnusson, M., C.R. Glasson, M.J. Vucko, A. Angell, T.L. Neoh and R. de Nys. 2019. Enrichment processes for the production of high-protein feed from the green seaweed *Ulva ohnoi*. *Algal Res* 41: 101555.
- Mata, T.M., A.A. Martins and N.S. Caetano. 2010. Microalgae for biodiesel production and other applications: a review. *Renew. Sust. Energy Rev.* 14: 217–232.
- Mata, L., M. Magnusson, N.A. Paul and R. de Nys. 2016. The intensive land-based production of the green seaweeds *Derbesia tenuissima* and *Ulva ohnoi*: biomass and bioproducts. *J. Appl. Phycol.* 28: 365–375.
- Mathimani, T., A. Baldinelli, K. Rajendran, D. Prabakar, M. Matheswaran, R.P. van Leeuwen and A. Pugazhendhi. 2019. Review on cultivation and thermochemical conversion of microalgae to fuels and chemicals: process evaluation and knowledge gaps. *J. Clean. Prod.* 208: 1053–1064.
- Matsumoto, M., D. Nojima, T. Nonoyama, K. Ikeda, Y. Maeda, T. Yoshino and T. Tanaka. 2017. Outdoor cultivation of marine diatoms for year-round production of biofuels. *Mar. Drugs* 15: 1–12.
- Meinita, M.D.N., B. Marhaeni, T. Winanto, G.T. Jeong, M.N.A. Khan and Y.K. Hong. 2013. Comparison of agarophytes (*Gelidium*, *Gracilaria*, and *Gracilariopsis*) as potential resources for bioethanol production. *J. Appl. Phycol.* 25: 1957–1961.
- Melis, A. 2009. Solar energy conversion efficiencies in photosynthesis: minimizing the chlorophyll antennae to maximize efficiency. *Plant Sci.* 177: 272–280.
- Milledge, J. and P. Harvey. 2016. Golden tides: problem or golden opportunity? The valorisation of *Sargassum* from beach inundations. *J. Mar. Sci. Engineer.* 4: 1–19.
- Milledge, J., B. Smith, P. Dyer and P. Harvey. 2014. Macroalgae-derived biofuel: a review of methods of energy extraction from seaweed biomass. *Energies* 7: 7194–7222.
- Montingelli, M.E., K.Y. Benyounis, J. Stokes and A.G. Olabi. 2016. Pretreatment of macroalgal biomass for biogas production. *Energy Convers. Manage.* 108: 202–209.
- Mooy, B.A.S.V., H.F. Fredricks, B.E. Pedler, S.T. Dyhrman, D.M. Karl, M. Koblížek, M.W. Lomas, T.J. Mincer, L.R. Moore and T. Moutin. 2009. Phytoplankton in the ocean use non-phosphorus lipids in response to phosphorus scarcity. *Nature* 458: 69–72.
- Moreno, J., M. Vargas, H. Rodríguez, J. Rivas and M.G. Guerrero. 2003. Outdoor cultivation of a nitrogen-fixing marine cyanobacterium, *Anabaena* sp. ATCC 33047. *Biomol. Eng.* 20: 191–197.
- Nagarajan, S., S.K. Chou, S. Cao, C. Wu and Z. Zhou. 2013. An updated comprehensive techno-economic analysis of algae biodiesel. *Bioresour. Technol.* 145: 150–156.
- Narala, R.R., S. Garg, K.K. Sharma, S.R. Thomas-Hall, M. Deme, Y. Li and P.M. Schenk. 2016. Comparison of microalgae cultivation in photobioreactor, open raceway pond, and a two-stage hybrid system. *Front. Energy Res.* 4: 1–10.
- Nardelli, A.E., V.G. Chiozzini, E.S. Braga and F. Chow. 2019. Integrated multi-trophic farming system between the green seaweed *Ulva lactuca*, mussel, and fish: a production and bioremediation solution. *J. Appl. Phycol.* 31: 847–856.
- Nascimento, I.A., S.S.I. Marques, I.T.D. Cabanelas, G.C.D. Carvalho, M.A. Nascimento, C.O.D. Souza, J.I. Druzian, J. Hussain and W. Liao. 2014. Microalgae versus land crops as feedstock for biodiesel: productivity, quality, and standard compliance. *Bioenergy Res.* 7: 1002–1013.
- Nasir, I.M., T.I.M. Ghazi and R. Omar. 2012. Production of biogas from solid organic wastes through anaerobic digestion: a review. *Appl. Microbiol. Biotechnol.* 95: 321–329.
- Nayak, M., W.I. Suh, Y.K. Chang and B. Lee. 2019. Exploration of two-stage cultivation strategies using nitrogen starvation to maximize the lipid productivity in *Chlorella* sp. HS2. *Bioresour. Technol.* 276: 110–118.
- North W.J. 1987. Oceanic farming of *Macrocystis*, the problems and non-problems. Seaweed cultivation for renewable resources. In: (K.T. Bird and P.H. Benson, eds.) *Seaweed cultivation for renewable resources*. Elsevier, Amsterdam. pp. 39–68.
- Norsker, N.H., M.J. Barbosa, M.H. Vermuë and R.H. Wijffels. 2011. Microalgal production – a close look at the economics. *Biotechnol. Adv.* 29: 24–27.
- Pádua, M.D., P.S.G. Fontoura and A.L. Mathias. 2004. Chemical composition of *Ulvaria oxysperma* (Kützting) bliding, *Ulva lactuca*

- (Linnaeus) and *Ulva fasciata* (Delile). *Braz. Arch. Biol. Technol.* 47: 49–55.
- Pančić, M. and T. Kiørboe. 2018. Phytoplankton defence mechanisms: traits and trade-offs. *Biol. Rev.* 93: 1269–1303.
- Patterson, T., R. Dinsdale and S. Esteves. 2008. Review of energy balances and emissions associated with biomass-based transport fuels relevant to the United Kingdom context. *Energy Fuels* 22: 3506–3512.
- Paumier, A., T. Tatlian, E. Réveillac, E. Le Luherne, S. Ballu, M. Lepage and O. Le Pape. 2018. Impacts of green tides on estuarine fish assemblages. *Estuar. Coast Shelf Sci.* 213: 176–184.
- Pedra, A.G.L.M., F. Ramlov, M. Maraschin and L. Hayashi. 2017. Cultivation of the red seaweed *Kappaphycus alvarezii* with effluents from shrimp cultivation and brown seaweed extract: Effects on growth and secondary metabolism. *Aquaculture* 479: 297–303.
- Posewitz, M.C. 2017. Algal oil productivity gets a fat bonus. *Nat. Biotechnol.* 35: 636–638.
- Qi, L., C. Hu, M. Wang, S. Shang and C. Wilson. 2017. Floating algae blooms in the East China Sea. *Geophys. Res. Lett.* 44: 11–501.
- Raikova, S., M.J. Allen and C.J. Chuck. 2019. Hydrothermal liquefaction of macroalgae for the production of renewable biofuels. *Biofuels Bioprod. Bior.* 13: 1483–1504.
- Ramachandra, T.V. and D. Hebbale. 2020. Bioethanol from macroalgae: Prospects and challenges. *Renew. Sust. Energy Rev.* 117: 109479.
- Raven, J.A. 2017. The possible roles of algae in restricting the increase in atmospheric CO₂ and global temperature. *Eur. J. Phycol.* 52: 506–522.
- Ravi, V., A.H. Gao, N.B. Martinkus, M.P. Wolcott and B.K. Lamb. 2018. Air quality and health impacts of an aviation biofuel supply chain using forest residue in the Northwestern United States. *Environ. Sci. Technol.* 52: 4154–4162.
- Remmers, I.M., R.H. Wijffels, M.J. Barbosa and P.P. Lamers. 2018. Can we approach theoretical lipid yields in microalgae? *Trends Biotechnol.* 36: 265–276.
- Renaud, S.M., D.L. Parry and L.-V. Thinh. 1994. Microalgae for use in tropical aquaculture I: gross chemical and fatty acid composition of twelve species of microalgae from the Northern Territory, Australia. *J. Appl. Phycol.* 6: 337–345.
- Rhein-Knudsen, N., M.T. Ale, F. Ajallouéian, L. Yu and A.S. Meyer. 2017. Rheological properties of agar and carrageenan from Ghanaian red seaweeds. *Food Hydrocolloid* 63: 50–58.
- Richardson, J.W., M.D. Johnson and J.L. Outlaw. 2012. Economic comparison of open pond raceways to photo bio-reactors for profitable production of algae for transportation fuels in the Southwest. *Algal Res.* 1: 93–100.
- Richmond, A., E. Lichtenberg, B. Stahl and A. Vonshak. 1990. Quantitative assessment of the major limitations on productivity of *Spirulina platensis* in open raceways. *J. Appl. Phycol.* 2: 195–206.
- Richmond, A., S. Boussiba, A. Vonshak and R. Kopel. 1993. A new tubular reactor for mass production of microalgae outdoors. *J. Appl. Phycol.* 5: 327–332.
- Robertson, G.P., S.K. Hamilton, B.L. Barham, B.E. Dale, R.C. Izaurralde, R.D. Jackson, D.A. Landis, S.M. Swinton, K.D. Thelen and J.M. Tiedje. 2017. Cellulosic biofuel contributions to a sustainable energy future: choices and outcomes. *Science* 356: eaal2324.
- Roesijadi, G., A.E. Copping, M.H. Huesemann, J. Foster and J.R. Benemann. 2010a. *Techno-economic feasibility analysis of off-shore seaweed farming for bioenergy and biobased products*. PNNL-19944; U.S. Department of Energy, Washington, DC, USA.
- Roesijadi, G., S.B. Jones, L.J. Snowdenswan and Y. Zhu. 2010b. *Macroalgae as a biomass feedstock: a preliminary analysis*. Office of Scientific & Technical Information Technical Reports. Richland, WA, USA.
- Romero-Villegas, G.I., M. Fiamengo, F.A. Fernández and E.M. Grima. 2018. Utilization of centrate for the outdoor production of marine microalgae at pilot-scale in flat-panel photobioreactors. *J. Biotechnol.* 284: 102–114.
- Saratale, R.G., G. Kumar, R. Banu, A. Xia, S. Periyasamy and G.D. Saratale. 2018. A critical review on anaerobic digestion of microalgae and macroalgae and co-digestion of biomass for enhanced methane generation. *Bioresour. Technol.* 262: 319–322.
- Satyanarayana, K.G., A.B. Mariano and J.V.C. Vargas. 2011. A review on microalgae, a versatile source for sustainable energy and materials. *Int. J. Energy Res.* 35: 291–311.
- Sen, S.M., E.I. Gürbüz, S.G. Wettstein, D.M. Alonso, J.A. Dumesic and C.T. Maravelias. 2012. Production of butene oligomers as transportation fuels using butene for esterification of levulinic acid from lignocellulosic biomass: process synthesis and techno-economic evaluation. *Green Chem.* 14: 3289–3294.
- Setchell, W.A. 1908. Nereocystis and Pelagophycus. *Botanical Gazette* 45: 125–134.
- Shakya, R., S. Adhikari, R. Mahadevan, S.R. Shanmugam, H. Nam and T.A. Dempster. 2017. Influence of biochemical composition during hydrothermal liquefaction of algae on product yields and fuel properties. *Bioresour. Technol.* 243: 1112–1120.
- Sheridan, C. 2013. Big oil turns on biofuels. *Nat. Biotechnol.* 31: 870–873.
- Shuba, E.S. and D. Kifle. 2018. Microalgae to biofuels: 'Promising' alternative and renewable energy, review. *Renew. Sust. Energy Rev.* 81: 743–755.
- Singh, A., P.S. Nigam and J.D. Murphy. 2011. Mechanism and challenges in commercialisation of algal biofuels. *Bioresour. Technol.* 102: 26–34.
- Smetacek, V. and A. Zingone. 2013. Green and golden seaweed tides on the rise. *Nature* 504: 84–88.
- Soleymani, M. and K Rosentrater. 2017 Techno-economic analysis of biofuel production from macroalgae (Seaweed). *Bioengineering* 4: 1–10.
- Sondak, C.F., P.O. Ang, J. Beardall, A. Bellgrove, S.M. Boo, G.S. Gerung, C.D. Hepburn, D.D. Hong, Z. Hu, H. Kawai and D. Largo. 2017. Carbon dioxide mitigation potential of seaweed aquaculture beds (SABs). *J. Appl. Phycol.* 29: 2363–2373.
- Soto, M., M.A. Vazquez, A. de Vega, J.M. Vilarino, G. Fernandez and M.E.S. de Vicente. 2015. Methane potential and anaerobic treatment feasibility of *Sargassum muticum*. *Bioresour. Technol.* 189: 53–61.
- Spolaore, P., C. Joannis-Cassan, E. Duran and A. Isambert. 2006. Commercial applications of microalgae. *J. Biosci. Bioeng.* 101: 87–96.
- Stephenson, A., J. Dennis and S. Scott. 2008. Improving the sustainability of the production of biodiesel from oilseed rape in the UK. *Process Saf. Environ. Prot.* 86: 427–440.
- Stephenson, P.G., C.M. Moore, M.J. Terry, M.V. Zubkov and T.S. Bibby. 2011. Improving photosynthesis for algal biofuels: toward a green revolution. *Trends Biotechnol.* 29: 615–623.

- Tabassum, M.R., A. Xia and J.D. Murphy. 2017. Potential of seaweed as a feedstock for renewable gaseous fuel production in Ireland. *Renew. Sustain. Energy Rev.* 68: 136–146.
- Tedesco, S., T.M. Barroso and A.G. Olabi. 2014. Optimization of mechanical pre-treatment of Laminariaceae Spp. Biomass-derived biogas. *Renew. Energy* 62: 527–534.
- Thompson, T.M., B.R. Young and S. Baroutian. 2019. Advances in the pretreatment of brown macroalgae for biogas production. *Fuel Process. Technol.* 195: 106151.
- Tibbetts, S.M., J.E. Milley and S.P. Lall. 2015. Chemical composition and nutritional properties of freshwater and marine microalgal biomass cultured in photobioreactors. *J. Appl. Phycol.* 27: 1109–1119.
- Torzillo, G., B. Pushparaj, F. Bocci, W. Balloni, R. Materassi and G. Florenzano. 1986. Production of *Spirulina* biomass in closed photobioreactors. *Biomass* 11: 61–74.
- Uribe, E., A. Vega-Gálvez, V. Heredia, A. Pastén and K. Di Scala. 2018. An edible red seaweed (*Pyropia orbicularis*): influence of vacuum drying on physicochemical composition, bioactive compounds, antioxidant capacity, and pigments. *J. Appl. Phycol.* 30: 673–683.
- Vardon, D.R., B.K. Sharma, G.V. Blazina, K. Rajagopalan and T.J. Strathmann. 2012. Thermochemical conversion of raw and defatted algal biomass via hydrothermal liquefaction and slow pyrolysis. *Bioresour. Technol.* 109: 178–187.
- Wang, X., L. Sheng and X. Yang. 2017. Pyrolysis characteristics and pathways of protein, lipid and carbohydrate isolated from microalgae *Nannochloropsis* sp. *Bioresour. Technol.* 229: 119–125.
- Wargacki, A.J., E. Leonard, M.N. Win, D.D. Regitsky, C.N.S. Santos, P.B. Kim, S.R. Cooper, R.M. Raisner, A. Herman, A.B. Sivitz and A. Lakshmanaswamy. 2012. An engineered microbial platform for direct biofuel production from brown macroalgae. *Science* 335: 308–313.
- Wassink, E.C. 1959. Efficiency of light energy conversion in plant growth. *Plant Physiol.* 34: 356–361.
- Wei, N., J. Quarterman and Y.S. Jin. 2013. Marine macroalgae: an untapped resource for producing fuels and chemicals. *Trends Biotechnol.* 31: 70–77.
- Weyer, K.M., D.R. Bush, A. Darzins and B.D. Willson. 2010. Theoretical maximum algal oil production. *Bioenergy Res.* 3: 204–213.
- Wu, Y.N., M. Mattsson, M.W. Ding, M.T. Wu, J. Mei and Y.L. Shen. 2019. Effects of different pretreatments on improving biogas production of macroalgae *Fucus vesiculosus* and *Fucus serratus* in Baltic Sea. *Energy Fuels* 33: 2278–2284.
- Xu, Z., G. Gao, J. Xu and H. Wu. 2017. Physiological response of a golden tide alga (*Sargassum muticum*) to the interaction of ocean acidification and phosphorus enrichment. *Biogeosciences* 14: 671–681.
- Ye, N.H., X.W. Zhang, Y.Z. Mao, C.W. Liang, D. Xu, J. Zou, Z.M. Zhuang and Q.Y. Wang. 2011. 'Green tides' are overwhelming the coastline of our blue planet: taking the world's largest example. *Ecol. Res.* 26: 477–485.
- Yoon, J.J., Y.J. Kim, S.H. Kim, H.J. Ryu, J.Y. Choi, G.S. Kim and M.K. Shin. 2010. Production of polysaccharides and corresponding sugars from red seaweed. *Adv. Mater. Res.* 93: 463–466.
- Yu, J., P. Wang, Y. Wang, J. Chang, S. Deng and W. Wei. 2018. Thermal constraints on growth, stoichiometry and lipid content of different groups of microalgae with bioenergy potential. *J. Appl. Phycol.* 30: 1503–1512.
- Zheng, S., M. He, J. Jiang, S. Zou, W. Yang, Y. Zhang, J. Deng and C. Wang. 2016. Effect of kelp waste extracts on the growth and lipid accumulation of microalgae. *Bioresour. Technol.* 201: 80–88.
- Zhou, D., L. Zhang, S. Zhang, H. Fu and J. Chen. 2010. Hydrothermal liquefaction of macroalgae *Enteromorpha prolifera* to bio-oil. *Energy Fuels* 24: 4054–4061.
- Zhuang, Y., J. Guo, L. Chen, D. Li, J. Liu and N. Ye. 2012. Microwave-assisted direct liquefaction of *Ulva prolifera* for bio-oil production by acid catalysis. *Bioresour. Technol.* 116: 133–139.

Supplementary Material: The online version of this article offers supplementary material (<https://doi.org/10.1515/bot-2019-0065>).

Bionotes



Guang Gao

State Key Laboratory of Marine Environmental Science, Xiamen University, Xiamen 361005, China; and Jiangsu Key Laboratory for Marine Bioresources and Environment, Jiangsu Ocean University, Lianyungang 222005, China
guang.gao@xmu.edu.cn

Guang Gao received his PhD at Newcastle University, UK. He is an associate professor at Xiamen University. He is working on the interaction between marine primary producers and environmental changes, referring to algal use in food, biofuel and bioremediation for CO₂ rise and eutrophication.



James Grant Burgess

School of Natural and Environmental Sciences, Newcastle University, Newcastle upon Tyne NE1 7RU, UK

James Grant Burgess received his PhD at Imperial College, University of London. He is a professor of marine biotechnology at Newcastle University, UK. He is interested in novel bioactive compounds from marine bacteria, chemical defense in marine microbes, antifouling compounds from marine bacteria, microbiology of deep sea sediments, and sponge microbiology.

Min Wu

Jiangsu Key Laboratory for Marine Bioresources and Environment, Jiangsu Ocean University, Lianyungang 222005, China

Min Wu is a postgraduate student at Jiangsu Ocean University. She is working on biofuel and bioactive compounds from microalgae.

**Shunjun Wang**

Jiangsu Key Laboratory for Marine
Bioresources and Environment, Jiangsu
Ocean University, Lianyungang 222005,
China

Shunjun Wang received her PhD at Nanjing Agricultural University, China. She is a professor at Jiangsu Ocean University. Her research fields are marine microbiology and bioactive compounds from marine organisms.

**Kunshan Gao**

State Key Laboratory of Marine
Environmental Science, Xiamen University,
Xiamen 361005, China
ksgao@xmu.edu.cn

Kunshan Gao obtained his PhD at Kyoto University, Japan. He is currently a chair professor at Xiamen University, China. His research focuses on physiological ecology and photobiology of phytoplankton, with special reference to the interactive effects of CO₂ rise (and associated ocean acidification) and solar UV radiation on aquatic primary producers.

Graphical abstract

Guang Gao, James Grant Burgess, Min Wu,
Shujun Wang and Kunshan Gao
**Using macroalgae as biofuel: current
opportunities and challenges**

<https://doi.org/10.1515/bot-2019-0065>
Botanica Marina 2020; x(x): xxx–xxx

Review: Compared to microalgae, the biomass productivity of macroalgae is lower. However, the lower cultivation and processing costs move the production of biomethane from macroalgae closer to profitability.

Keywords: biofuel; biogas; bioremediation; cultivation; macroalgae; photosynthesis.

