

Cloning and expression of prostaglandin E₂ receptor subtype 1 (*ep₁*) in *Bostrichthys sinensis*

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Received: 7 May 2013 / Accepted: 17 February 2014 / Published online: 25 February 2014
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Abstract Our previous studies suggested that prostaglandin E₂ (PGE₂) is a putative sex pheromone in Chinese black sleeper *Bostrichthys sinensis*, a fish species that inhabits intertidal zones and mates and spawns inside a muddy burrow. We found immunoreactivities of PGE₂ receptor subtypes (Ep_{1–3}) expressed in the olfactory sac, but only Ep₁ presented higher density of immunoreactivity in mature fish than that in immature fish in both sexes. To gain a better understanding of the underlying molecular mechanism for the detection of PGE₂ in the olfactory system, we cloned an *ep₁* cDNA from the adult olfactory sac. The open-reading frame of the *ep₁* consisted of 1,134-bp nucleotides that encoded a 378-amino acid-long protein with a seven-transmembrane domain, typical

for the G protein-coupled receptors superfamily. Expression of *ep₁* mRNA was observed in all tissues examined, with higher levels obtained in the olfactory sacs and testes. The expression of *ep₁* mRNA in the olfactory sacs and gonads was significantly higher in both sexes of mature fish than in those of immature ones. Taken together, our results suggested that Ep₁, which is highly expressed in the olfactory sacs and gonads of mature fish, is important for the control of reproduction and may be involved in PGE₂-initiated spawning behavior in *B. sinensis*.

Keywords *Bostrichthys sinensis* · PGE₂ · Ep₁ · Sex pheromone · Olfactory system

Electronic supplementary material The online version of this article (doi:10.1007/s10695-014-9923-x) contains supplementary material, which is available to authorized users.

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Introduction

Animals demonstrate their reproductive readiness in various ways in order to find mating partners. Many animal species, including fishes, employ sex pheromones during their reproductive seasons in order to attract mating partners (Wyatt 2003). It has become increasingly evident that many fish species release steroids (e.g., *Carassius auratus*), prostaglandins (e.g., *Salmo salar*) or other metabolites (e.g., *Petromyzon marinus*, *Oncorhynchus masou*) during gamete maturation as exogenous signals that trigger mating behavior (Dulka et al. 1987; Sorensen et al. 1988; Moore and Waring 1996; Li et al. 2002; Yambe et al. 2006). Furthermore, electro-olfactogram (EOG)

recording studies in more than 100 fish species (from the Cypriniformes, Siluriformes, Salmoniformes, Scorpaeniformes and Perciformes) (Sveinsson and Hara 2000) demonstrate that the olfactory system of teleosts is an important seat for processing reproduction-related or pheromone-triggered signals (Biju et al. 2003). However, the molecular mechanisms underlying the detection of sex pheromones in the teleost olfactory system are not well understood.

Signal transduction in the olfactory system begins with the binding of an odorant ligand to a receptor on the olfactory neuron cell surface, initiating a cascade of enzymatic reactions that results in the production of a second messenger and the eventual depolarization of the cell membrane (Breer 1994). The odorant receptors are believed to be members of the superfamily of G protein-coupled receptors (GPCRs) that recognize diverse ligands (Zhao et al. 1998).

The Chinese black sleeper (*Bostrichthys sinensis* Lacepede) belongs to the family Eleotridae and the order Perciformes. This species inhabits intertidal zones. In the mudflats, the fish build 'Y'-shaped muddy burrows of 40–65 cm depth, with one entrance and one exit. During the spawning season, a pair of fish mates and spawns inside a burrow (Hong et al. 2004). This spawning behavior suggests that mature males and females may release sex pheromones to attract each other and to elicit courtship and spawning behavior. In response to prostaglandin E_2 (PGE₂), EOG values are higher in mature fish than in immature ones (Ma et al. 2003). Moreover, PGE₂ concentrations in water samples obtained from spawning pairs are higher than those from non-spawners. Further analyses show that artificial nests with a PGE₂-releasing tube inside attract more males and females than the control and result in the highest percentage of spawning (Hong et al. 2006). All this evidence indicated that PGE₂ might act as a sex pheromone in this species.

To understand the molecular mechanisms of PGE₂ in inducing sexual behavior, we focused on the receptor for PGE₂. PGE₂ exerts its potential by acting on a group of GPCRs. There are four GPCRs that respond to PGE₂, designated as subtypes Ep₁, Ep₂, Ep₃, and Ep₄ (Narumiya et al. 1999). In a previous study, using immunocytochemistry, we found PGE₂ receptors in the complete olfactory system of *B. sinensis* (Lai and Hong 2010). In the olfactory sacs, among Ep_{1–3}, only Ep₁ presents a significantly higher density of immunoreactivity in mature fish than that in

immature fish in both sexes. Moreover, the density of Ep₁ immunoreactivity in olfactory epithelium is higher (above 7- to 20-fold) than that of Ep_{2,3}. These results suggest that Ep₁ may play reproduction-related functions in the olfactory epithelium. To gain a better understanding of the underlying molecular mechanism for the detection of PGE₂ in the olfactory sac, molecular technologies were used in the present study. We first cloned the full-length *ep1* cDNA from the *B. sinensis* olfactory sac and further analyzed its expression pattern in adult fish tissues. Finally, the changes in *ep1* mRNA levels in the olfactory sac were analyzed in different reproductive statuses.

Materials and methods

Animal and tissue sampling

Adult Chinese black sleepers (*B. sinensis*) were collected from the Jiulong River Estuary, Fujian, China, during the spawning (May and June, 2010) and non-spawning seasons (December 2009 and January 2010). Body length and body weight ranges were 162–166 mm and 58.5–75.6 g for females, and 160–177 mm and 51.5–67.5 g for males. The sexual maturity of males was confirmed by applying pressure on the abdomen to check for milt flow. For sexually mature females, eggs could be squeezed out by gentle pressure on the abdomen.

For tissue sampling, the fish were anesthetized with 0.2 % 3-aminobenzoic acid ethyl ester (MS-222, Sigma, St. Louis, MO, US) before being killed. The gonadosomatic index (GSI) was calculated as $GSI (\%) = \text{gonad weight (g)} \times 100 / \text{total body weight (g)}$.

Cloning and sequence analysis of *ep1* and β -actin cDNA

Total RNA was extracted from the adult olfactory sac using Trizol reagent following the manufacturer's instructions (NRC, USA) and reverse-transcribed to cDNA using a RevertAid First Strand cDNA synthesis Kit (Fermentas, Canada). To obtain a partial *ep1* and β -actin cDNA sequence, olfactory cDNAs were used as a template in PCR with a primer set (Table 1) corresponding to highly conserved amino acid sequences found in known Ep₁ and β -actin. The following thermal cycling parameters were used:

Table 1 Primer sequences used in this study

Primers	Primer set (5' → 3')
First round of RT-PCR	
<i>β-actin</i> F	AGACCTTCAACACCCCHGCCAT
<i>β-actin</i> R	ACTCCTGCTTGCTRATCCACAT
<i>ep₁</i> F	GCCVGSACKTGGTGCTTCAT
<i>ep₁</i> R	TAMACCCANGGRTCSARDATCTG
<i>ep₁</i> 5', 3'-RACE	
3'GSP1	CTGCCGTTTGGTGATGCGATA
3'GSP2	AGTGCGGACATCAACCCAAAGA
Outer primer	CATGGCTACATGCTGACAGCCTA
Inner primer	CGCGGATCCACAGCCTACTGATGATCAGTCGATG
5'GSP1	CTGCCGTTTGGTGATGCGATA
5'GSP2	AGTGCGGACATCAACCCAAAGA
Real-time PCR	
<i>β-actin</i> F-q	GACAGGTCATCATCATTGGC
<i>β-actin</i> R-q	CAGACAGCACAGTGGTGGCATA
<i>ep₁</i> F-q	CAAGACTAAAGAAGACGACCTGC
<i>ep₁</i> R-q	AGTGCGGACATCAACCCAAAG

94 °C for 3 min (1 cycle); 94 °C for 30 s, 56 °C for *β-actin* and 58 °C for *ep₁* for 1 min and 72 °C for 1 min (35 cycles) followed by a final DNA extension at 72 °C for 5 min. All PCR products were then purified from agarose gel and sub-cloned into vector PTZR/T (InsTAclone PCR Cloning Kit, Fermentas, Canada), and then, the plasmid DNA of several positive clones was prepared for DNA sequence analysis.

To obtain full-length *ep₁*, olfactory sac total RNA was reverse-transcribed to 5'- and 3'-RACE ready cDNA using either a 5'-Full RACE Kit or a 3'-Full RACE Core Set (Takara, Japan), in accordance with the manufacturer's instructions. We isolated the 5'- and 3'-ends of the *ep₁* cDNA, using gene-specific primers (Table 1). Based on the partial cDNA sequence obtained, gene-specific primers were designed for further extension by 5'- and 3'-RACE. These initial 5'- and 3'-RACE products were then used for nested PCR amplification with gene-specific nested primers (GSP). The PCR products were sub-cloned and sequenced. All RACE reactions were carried out following the manufacturer's instructions.

Phylogenetic analysis

After obtaining the *B. sinensis ep₁* cDNA sequence, a BLAST homology search was performed using the

deduced amino acid sequences. The alignment of multiple *Ep₁* protein sequences from different vertebrate species was performed using the Megalign program of the Lasergene software package (DNASTAR Inc., Madison, WI, USA) with the Clustal V (PAM 250) algorithm. A phylogenetic tree was constructed using the neighbor-joining method (Saitou and Nei 1987).

Quantification of *ep₁* expression

To assess *ep₁* mRNA expression in different tissues of *B. sinensis*, olfactory sac, olfactory bulb, telencephalon, gill, muscle, liver, spleen, intestine, stomach, ovary, seminal vesicle and testis samples were collected from three females or males. In addition, olfactory sac, stomach, ovary and testis tissues were also collected during the spawning and non-spawning seasons, placed into liquid nitrogen and then stored in a −80 °C refrigerator. Total RNA was isolated, treated with DNase I (Fermentas, Canada) and reverse-transcribed to cDNAs as described above. Samples of the cDNAs were diluted 1:10 prior to use as templates for quantitative real-time PCR (qPCR) using a SYBR GREEN 2 × Premix Kit (Takara, Japan) following the manufacturer's instructions. The specificity and efficiency of these primer sets (Table 1) were examined using serial dilutions of

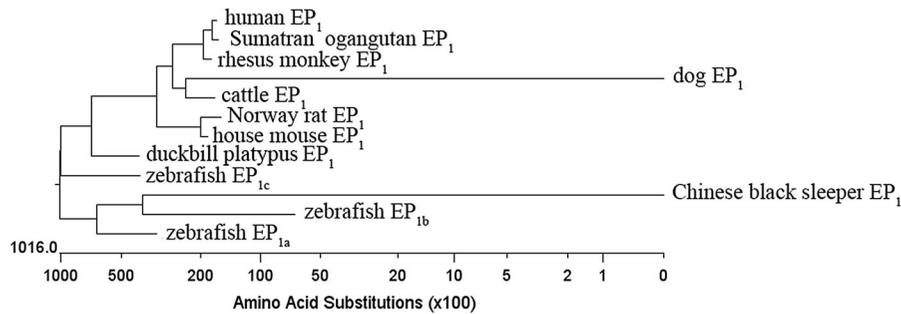
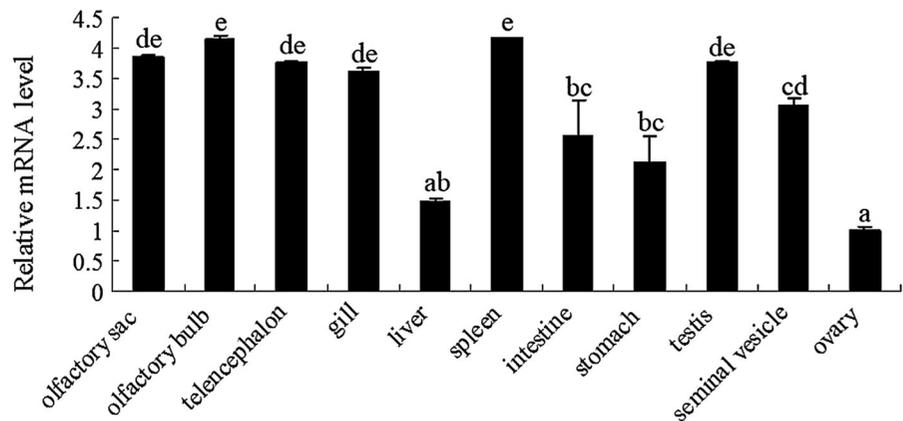


Fig. 2 Phylogenetic analysis of EP₁. Cattle EP₁ accession number: XP_584035.3; Chinese black sleeper EP₁ accession number: JF946753; dog EP₁ accession number: XP_853418.1; duckbill platypus EP₁ accession number: XP_001516507.1; house mouse EP₁ accession number: BAB29498.1; human EP₁ accession number: AAP32302.1; Norway rat EP₁ accession number: NP_037232.1; rhesus monkey EP₁ accession number:

NP_001028205.1; Sumatran orangutan EP₁ accession number: XP_002828836.1; zebrafish EP_{1a} accession number: NP_001159805.1; zebrafish EP_{1b} accession number: NP_001159802.1; and zebrafish EP_{1c} accession number: ACX47465.1. The horizontal distances to the branching points are proportional to the number of amino acid substitutions

Fig. 3 Relative expression levels of *ep1* mRNA in various tissues of *B. sinensis*. The *ep1* mRNA expression levels were normalized by the expression of β -actin mRNA in each tissue. Values represent mean \pm SEM ($n = 3$) relative to *ep1* mRNA levels measured in the tissue with the lowest value. Bars marked with different letters are significantly different from each other ($P < 0.05$)



expression levels of *ep1* mRNA were found in the olfactory sac, olfactory bulb, spleen and testis, while significantly ($P < 0.05$) low expression levels were observed in the liver and ovary.

High expression of PGE₂ receptor subtype *ep1* mRNA in mature fish

The GSI values of sexually mature males ($0.14 \pm 0.042\%$) and females ($7.38 \pm 3.0\%$) were significantly ($P < 0.05$) higher than those of immature males ($0.030 \pm 0.0084\%$) and females ($0.48 \pm 0.072\%$).

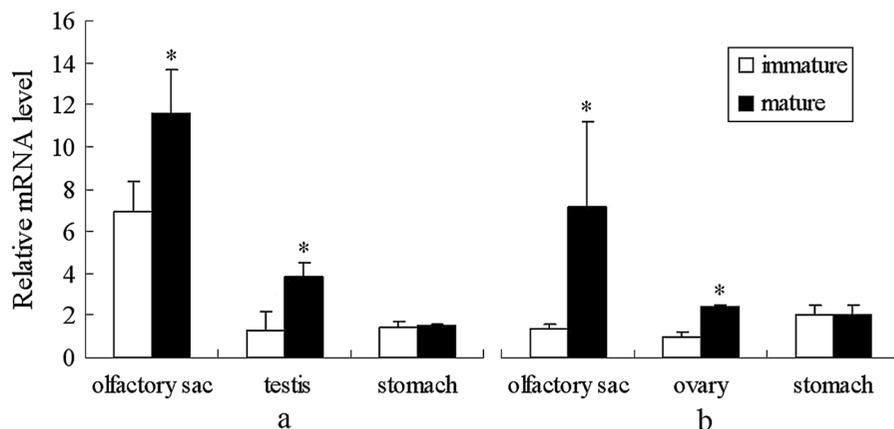
Significantly ($P < 0.05$) higher expression of *ep1* was observed in the olfactory sac in sexually mature fish than in immature ones in both sexes (Fig. 4). Similarly, significantly ($P < 0.05$) higher expression of *ep1* mRNA was also observed in the gonads in

mature fish than in immature ones in both sexes. In contrast, no significant differences in *ep1* mRNA expression were found in the stomach tissues of sexually mature fish compared with the immature fish in both sexes (Fig. 4).

Discussion

For many genes, ray-finned fish have two paralogous copies, whereas only one ortholog is present in tetrapods (Wittbrodt et al. 1998). This is related to the teleost-specific genome duplication which occurred after the split of the Acipenseriformes and the Semionotiformes from the lineage leading to teleost fish, but before the divergence of the Osteoglossiformes (Hoegg et al. 2004). Three zebrafish

Fig. 4 Relative expression levels of *ep₁* mRNA in olfactory sac, ovaries and stomach tissues of male (a) and female (b) *B. sinensis*. Values represent mean \pm SEM ($n = 3$) relative to *ep₁* mRNA levels measured in the tissue with the lowest value. Bars marked with an * indicate significant difference between mature and immature *B. sinensis* ($P < 0.05$)



prostaglandin E receptor subtype 1 (*zep₁*) isoforms—*zep_{1a}* (GQ911587), *zep_{1b}* (GQ911588) and *zep_{1c}* (GQ911589)—are identified from the adult ovary (Kwok et al. 2012). However, experimental trials to isolate additional *ep₁* cDNAs or in silico approaches to identify related sequences did not provide evidence for the existence of any additional *ep₁*-like genes or mRNA isoforms from one gene in *B. sinensis*. Thus, it is likely that the *B. sinensis* genome had lost the additional *ep₁* gene.

In the mouse (*Mus musculus*), the distribution of *ep₁* mRNA is restricted to several organs, such as the kidney, lung and stomach (Watabe et al. 1993). In the brain, *ep₁* mRNA is also found in the mouse dorsal root ganglion (Oida et al. 1995) and the paraventricular nucleus of the piglet hypothalamus (Coleman et al. 1994). In the present study, *ep₁* mRNA was expressed in all the tissues sampled from adult *B. sinensis*, with a relatively higher expression in the brain tissues. This expression pattern suggests that Ep₁ mediates a vast range of PGE₂ actions in teleost fish, including gastric motor activity and emptying, inhibiting sodium salt absorption in collecting tubules, sensitization of neurons to nociception, inhibition of sleep induction and stress-induced impulsive behavior (Breyer et al. 1998; Matsuoka et al. 2005; Mizuguchi et al. 2010; Moriyama et al. 2005; Yoshida et al. 2000). It is interesting that in the present study, *ep₁* mRNA was highly expressed in the spleen. Studies in mammalian models indicated Ep₁ possibly functions as a potent immunomodulator in spleen B lymphocytes (Phipps et al. 1991). Further study in mouse indicated that EPs are sufficient for mediating PGE-induced growth inhibition of susceptible B lineage cells (Fedyk et al. 1996). Besides, the high

expression levels of *ep₁* mRNA in spleen may suggest spleen plays an important role in control of PGE₂ synthesis in *B. sinensis*. However, our previous study indicated that sex organs serve as major sources of sex pheromones/PGE₂ in *B. sinensis* (Hong et al. 2006). Therefore, the major function of PGE₂ synthesis in spleen may be related to local homeostasis (Smith and Langenbach 2001; North et al. 2007), but not released into water as sex pheromones. The Ep₁-mediated PGE₂ function on spleen of *B. sinensis* needs to be further investigated.

A recent study in mice also demonstrates that the four PGE₂ receptor subtypes (EP₁, EP₂, EP₃ and EP₄) are expressed in the olfactory epithelium (Fukuiri et al. 2013). EP₄ is the most abundantly expressed, the expression of EP₂ is moderate, that of EP₃ is mild and EP₁ is hardly detectable. However, so far, we have not been successful in cloning *ep₂₋₄* in the olfactory sac of mature *B. sinensis*. Also, a transcriptome analysis did not detect *ep₂₋₄* in the olfactory sac in immature *B. sinensis* (data unpublished). It is worth noting that our previous study indicates that the density of Ep₁ immunoreactivity in olfactory epithelium is significantly higher (above 7–20 fold) than that of Ep_{2,3} in both sexes regardless of the reproductive stages (Lai and Hong 2010). Further study is necessary to clone *ep₂₋₄* using genomic DNA as template in the spawning season or using cDNA template from an *eps* highly expressed organ, i.e., the kidney.

Our present study revealed that the PGE₂ receptor subtype *ep₁* mRNA was present in the olfactory sac and gonad of *B. sinensis* and that its expression was significantly higher in mature *B. sinensis* than in immature *B. sinensis*. These results coincide with our

previous immunocytochemistry and electrophysiology studies (Lai and Hong 2010; Ma et al. 2003), which demonstrate that the number of immunoreactive olfactory receptor neurons and the EOG of the olfactory organ are related to reproductive status in *B. sinensis*. Variations with reproductive status in response to putative sex pheromones are found widely in teleosts (Irvine and Sorensen 1993; Moore and Waring 1995; Belanger et al. 2004). The levels of olfactory sensitivities may be modulated by changes in the number of receptors and/or the sensitivities of the receptors (Creese and Sibley 1981; Habibi et al. 1989). Our results suggest that olfactory sensitivities in response to sex pheromones are related to the number of receptors in *B. sinensis*, which might further explain the season-related variation of olfactory sensitivities in response to sex pheromones in teleosts (Moore and Waring 1996).

Conclusions

In the present study, we cloned a full-length cDNA coding an *ep₁* from the olfactory sac of *B. sinensis*. The higher expression of *ep₁* mRNA in the olfactory sac in mature males and females suggested that *Ep₁* might be a mediator for PGE₂ to act as a sex pheromone. Moreover, our results suggest that the increased olfactory sensitivity to PGE₂ might be related to the number of the *Ep₁* in olfactory sac of *B. sinensis*. Further studies are needed to investigate the *Ep₁*-mediated signal transduction pathway in the olfactory sac of *B. sinensis*.

Acknowledgments This work was supported by the National Natural Science Foundation of China (No. 41276129, 40776080), the fund for Doctor Station of the Ministry of China (No. 20120121110029) and the Major Project of Agricultural Science and Technology of Fujian Province (2012N5011). Professors J. Hodgkiss and Y. Zhu are thanked for their help with English language of the manuscript.

References

- Belanger AJ, Arbuckle WJ, Corkum LD, Gammon DB, Li W, Scott AP, Zielinski BS (2004) Behavioral and electrophysiological responses by reproductive female *Neogobius melanostomus* to odours released by conspecific males. *J Fish Biol* 65:933–946. doi:10.1111/j.0022-1112.2004.00494.x
- Biju KC, Singru PS, Schreibman MP, Subhedar N (2003) Reproduction phase-related expression of GnRH-like immunoreactivity in the olfactory receptor neurons, their projections to the olfactory bulb and in the nervous terminalis in the female Indian major carp *Cirrhinus mrigala* (Ham.). *Gen Comp Endocrinol* 133:358–367. doi:10.1016/S0016-6480(03)00190-4
- Breer H (1994) Odor recognition and second messenger signaling in olfactory receptor neurons. *Semin Cell Biol* 5:25–32. doi:10.1006/scel.1994.1004
- Breyer MD, Zhang Y, Guan YF, Hao CM, Hebert RL (1998) Regulation of renal function by prostaglandin E receptors. *Kidney Int* 54:S88–S94. doi:10.1046/j.1523-1755.1998.06718.x
- Coleman RA, Grix SP, Head SA, Louttit JB, Mallett A, Sheldrick RL (1994) A novel inhibitory prostanoid receptor in piglet saphenous vein. *Prostaglandins* 47:151–168. doi:10.1016/0090-6980(94)90084-1
- Creese I, Sibley DR (1981) Receptor adaptations to centrally acting drugs. *Annu Rev Pharmacol Toxicol* 21:357–391. doi:10.1146/annurev.pa.21.040181.002041
- Dulka JG, Stacey NE, Sorensen PW, Van Der Kraak GJ (1987) A sex steroid pheromone synchronizes male-female spawning readiness in the goldfish. *Nature* 325:251–253. doi:10.1038/325251a0
- Fedyk BR, Ripper JM, Brown DM, Phipps RP (1996) A molecular analysis of PGE receptor (EP) expression on normal and transformed B lymphocytes: coexpression of EP₁, EP₂, EP_{3β} and EP₄. *Mol Immunol* 33:33–45. doi:10.1016/0161-5890(95)00130-1
- Fukui T, Takumida M, Nakashimo Y, Hirakawa K (2013) Expression of prostanoid receptors (EP₁, 2, 3, and 4) in normal and methimazole-treated mouse olfactory epithelium. *Acta Otolaryngol* 133:70–76. doi:10.3109/00016489.2012.712214
- Habibi HR, de Leeuw R, Nahorniak CS, Th Goos HJ, Peter RE (1989) Pituitary gonadotropin-releasing hormone (GnRH) receptor activity in goldfish and catfish: seasonal and gonadal effects. *Fish Physiol Biochem* 7:109–118. doi:10.1007/BF00004696
- Hoegg S, Brinkmann H, Taylor JS, Meyer A (2004) Phylogenetic timing of the fish-specific genome duplication correlates with the diversification of teleost fish. *J Mol Evol* 59:190–203. doi:10.1007/s00239-004-2613-z
- Hong WS, Zhao WH, Ma XL, Guo XF, Zhang QY, Chai MJ, Zheng WY (2004) A preliminary study on the induction of spawning by sex pheromones in *Bostrichthys sinensis*. *J Fisheries China* 28:225–230. doi:10.3321/j.issn:1000-0615.2004.03.001 (in Chinese with English abstract)
- Hong WS, Chen SX, Zhang QY, Zheng WY (2006) Sex organ extracts and artificial hormonal compounds as sex pheromones to attract broodfish and to induce spawning of Chinese black sleeper (*Bostrichthys sinensis* Lacepede). *Aquacul Res* 37:529–534. doi:10.1111/j.1365-2109.2006.01462.x
- Irvine IAS, Sorensen PW (1993) A cute olfactory sensitivity of wild common carp, *Cyprinus Carpio*, to goldfish hormonal sex pheromones is influenced by gonadal maturity. *Can J Zool* 71:2199–2210. doi:10.1139/z93-309
- Kwok A, Wang Y, Leung F (2012) Molecular characterization of prostaglandin F receptor (FP) and E receptor subtype 1

- (EP₁) in zebrafish. *Gen Comp Endocrinol* 178:216–226. doi:[10.1016/j.ygcen.2012.05.002](https://doi.org/10.1016/j.ygcen.2012.05.002)
- Lai XJ, Hong WS (2010) Variation with reproductive status of PGE₂ receptor immunoreactivities in the *Bostrichthys sinensis* olfactory system. *J Fish Biol* 77:1542–1551. doi:[10.1111/j.1095-8649.2010.02791.x](https://doi.org/10.1111/j.1095-8649.2010.02791.x)
- Li W, Scott AP, Siefkes MJ, Yan H, Liu Q, Yun S, Gage D (2002) Bile acid secreted by male sea lamprey that acts as a sex pheromone. *Science* 296:138–141. doi:[10.1126/science.1067797](https://doi.org/10.1126/science.1067797)
- Ma XL, Hong WS, Chai MJ, Pan LA, Huang HY, Zhang QY (2003) Comparisons of EOG to sex pheromones in *Bostrichthys sinensis* Lacepede. *J Xiamen Univ (Natural Science)* 42:781–786. doi:[10.3321/j.issn:0438-0479.2003.06.022](https://doi.org/10.3321/j.issn:0438-0479.2003.06.022) (in Chinese with English abstract)
- Matsuoka Y, Furuyashiki T, Yamada K, Nagai T, Bito H, Tanaka Y, Kitaoka S, Ushikubi F, Nabeshima T, Narumiya S (2005) Prostaglandin E receptor EP₁ controls impulsive behavior under stress. *Proc Natl Acad Sci USA* 102:16066–16071. doi:[10.1073/pnas.0504908102](https://doi.org/10.1073/pnas.0504908102)
- Mizuguchi S, Ohno T, Hattori Y, Ae T, Minamino T, Satoh T, Arai K, Saeki T, Hayashi I, Sugimoto Y, Narumiya S, Saigenji K, Majima M (2010) Roles of prostaglandin E₂-EP₁ receptor signaling in regulation of gastric motor activity and emptying. *Am J Physiol Gastrointest Liver Physiol* 299:G1078–G1086. doi:[10.1152/ajpgi.00524.2009](https://doi.org/10.1152/ajpgi.00524.2009)
- Moore A, Waring CP (1995) Seasonal changes in olfactory sensitivity of mature male Atlantic salmon (*Salmo salar* L.) parr to prostaglandins. In: Goetz FW, Thomas P (eds) *Proceedings of the 5th international symposium on reproductive physiology of fish*. Fish Symp, Austin, 1995, pp 273
- Moore A, Waring CP (1996) Electrophysiological and endocrinological evidence that F-series prostaglandins function as priming pheromones in mature male Atlantic salmon (*Salmo salar* parr). *J Exp Biol* 199:2307–2316. doi:[10.1016/S0008-6223\(98\)00300-5](https://doi.org/10.1016/S0008-6223(98)00300-5)
- Moriyama T, Higashi T, Togashi K, Iida T, Segi E, Sugimoto Y, Tominaga T, Narumiya S, Tominaga M (2005) Sensitization of TRPV1 by EP₁ and IP reveals peripheral nociceptive mechanism of prostaglandins. *Mol Pain* 1: 3. doi:[10.1186/1744-8069-1-3](https://doi.org/10.1186/1744-8069-1-3)
- Morrison TB, Weis JJ, Wittwer CT (1998) Quantification of low-copy transcripts by continuous SYBR Green I monitoring during amplification. *Biotechniques* 24:954–962
- Narumiya S, Sugimoto Y, Ushikubi F (1999) Prostanoid receptors structures, properties, and functions. *Physiol Rev* 79:1193–1226
- North TE, Goessling W, Walkley CR, Lengerke C, Kopani KR, Lord AM, Weber GJ, Bowman TV, Jang IH, Grosser T, Fitzgerald GA, Daley GQ, Orkin SH, Zon LI (2007) Prostaglandin E₂ regulates vertebrate haematopoietic stem cell homeostasis. *Nature* 447(7147):1007–1011. doi:[10.1038/nature05883](https://doi.org/10.1038/nature05883)
- Oida H, Namba T, Sugimoto Y, Ushikubi F, Ohishi H, Ichikawa A, Narumiya S (1995) In situ hybridization studies of prostacyclin receptor mRNA expression in various mouse organs. *Br J Pharmacol* 116:2828–2837. doi:[10.1111/j.1476-5381.1995.tb15933.x](https://doi.org/10.1111/j.1476-5381.1995.tb15933.x)
- Phipps RP, Stein SH, Roper RL (1991) A new view of prostaglandin E regulation of the immune response. *Immunol Today* 12:349–352. doi:[10.1016/0167-5699\(91\)90064-Z](https://doi.org/10.1016/0167-5699(91)90064-Z)
- Saitou N, Nei M (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4:406–425
- Smith WL, Langenbach R (2001) Why there are two cyclooxygenase isozymes. *J Clin Invest* 107(12):1491–1495. doi:[10.1172/JCI13271](https://doi.org/10.1172/JCI13271)
- Sorensen PW, Hara TJ, Stacey NE, Goetz FW (1988) F prostaglandins function as potent olfactory stimulants that comprise the postovulatory female sex pheromone in goldfish. *Biol Reprod* 39:1039–1050. doi:[10.1095/biolreprod39.5.1039](https://doi.org/10.1095/biolreprod39.5.1039)
- Sveinsson T, Hara TJ (2000) Olfactory sensitivity and specificity of Arctic char, *Salvelinus alpinus*, to a putative male pheromone, prostaglandin F_{2α}. *Physiol Behav* 69:301–307. doi:[10.1016/S0031-9384\(99\)00253-X](https://doi.org/10.1016/S0031-9384(99)00253-X)
- Watabe A, Sugimoto Y, Honda A, Irie A, Namba T, Negishi M, Ito S, Narumiya S, Ichikawa A (1993) Cloning and expression of cDNA for a mouse EP₁ subtype of prostaglandin E receptor. *J Biol Chem* 268:20175–20178
- Wittbrodt J, Meyer A, Scharlt M (1998) More genes in fish? *BioEssays* 20:511–515
- Wyatt TD (2003) *Pheromones and animal behaviour: communication by smell and taste*. Cambridge University Press, Cambridge
- Yambe H, Kitamura S, Kamio M, Yamada M, Matsunaga S, Fusetani N, Yamazaki F (2006) L-Kynurenine, an amino acid identified as a sex pheromone in the urine of ovulated female masu salmon. *Proc Natl Acad Sci USA* 103:15370–15374. doi:[10.1073/pnas.0604340103](https://doi.org/10.1073/pnas.0604340103)
- Yoshida Y, Matsumura H, Nakajima T, Mandai M, Urakami T, Kuroda K, Yoneda H (2000) Prostaglandin E (EP) receptor subtypes and sleep: promotion by EP₄ and inhibition by EP₁/EP₂. *NeuroReport* 11:2127–2131
- Zhao H, Ivic L, Otaki JM, Hashimoto M, Mikoshiba K, Firestein S (1998) Functional expression of a mammalian odorant receptor. *Science* 279:237–242. doi:[10.1126/science.279.5348.237](https://doi.org/10.1126/science.279.5348.237)