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Using ²²⁴Ra/²²⁸Th disequilibrium to quantify benthic fluxes of dissolved inorganic carbon and nutrients into the Pearl River Estuary

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Abstract

The ²²⁴Ra/²²⁸Th disequilibrium that was recently observed in coastal sediments has been proven to be an excellent proxy for tracing the benthic processes that regulate solute transfer across the sediment–water interface. In order to better utilize this proxy, there is a need to understand the reaction kinetics of ²²⁴Ra in sediments. In this study, depth profiles of ²²⁴Ra and ²²⁸Th in bulk sediments were collected along a transect in the Pearl River Estuary (PRE). Together with bulk sediment measurements, dissolved ²²⁴Ra, dissolved inorganic carbon (DIC), and nutrients (NO₂⁻ + NO₃⁻, NH₄⁺) in pore water and in the overlying waters were also determined. A marked deficit of ²²⁴Ra with respect to ²²⁸Th with large spatial variations was observed in the PRE sediments. By use of a diagenetic model for the distributions of dissolved and adsorbed ²²⁴Ra in sediments, we infer that adsorption removes ²²⁴Ra from aqueous phase at a rate of 0.1 ± 1.1 –2000 ± 400 d⁻¹. In addition, adsorption of ²²⁴Ra exhibits a rate sequence of oxic freshwater > anoxic freshwater > anoxic brackish water, probably reflecting the effect of the redox conditions and ionic strength on the adsorption–desorption kinetics of ²²⁴Ra.

Benthic fluxes of ²²⁴Ra were estimated from the observed deficit of ²²⁴Ra in the sediments using a one-dimensional (1D) mass balance exchange model. We demonstrated that irrigation was the predominant process that controls solute transfer across the sediment–water interface, whereas molecular diffusion and sediment mixing together contributed <5% of the total ²²⁴Ra fluxes from bottom sediments. We then utilized the ²²⁴Ra/²²⁸Th disequilibrium approach to quantify the benthic fluxes of DIC and nutrients. We showed that sediment interstitial waters delivered approximately $42 \pm 6 \times 10^9$ mol of DIC and $\sim 16 \pm 1 \times 10^9$ mol of NH⁺₄ into the PRE in the dry season. In contrast, it removed about $13 \pm 1 \times 10^9$ mol of NO⁻₃ from the overlying water column. The benthic flux of DIC is equivalent to $\sim 18\%$ of the riverine input in this season. In terms of nutrients, our results suggest that bottom sediments are a major sink of water column NO⁻₃, and are a predominant source of NH⁺₄ in the PRE. Overall, this study indicates that irrigation is an important process and must be considered in the mass balance of DIC and nutrients in estuaries.

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1. INTRODUCTION

Exchange of sediment interstitial waters with the water column has long been considered a major source of many chemical species in estuaries (e.g., Knox et al., 1981).

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However, release rates of these species from estuarine sediments remain poorly constrained, due primarily to limitations inherent in the traditional approaches for quantifying the benthic fluxes across the sediment-water interface in dynamic coastal settings, i.e., the benthic chamber method and the modeling approach (e.g., Hensen et al., 2006). The newly developed 224 Ra/ 228 Th disequilibrium approach utilizes the deficit of ²²⁴Ra with respect to ²²⁸Th in near-surface sediments to determine the ²²⁴Ra flux across the sediment-water interface. By combining the concentration gradient of ²²⁴Ra in pore water with the concentration gradient of the dissolved species of interest at the interface. the ²²⁴Ra flux is converted into a benthic flux of that species (Cai et al., 2012, 2014). This approach does not impose any interference on the system and is particularly suitable for studies on the benthic exchange in estuaries, where a marked deficit of ²²⁴Ra with respect to ²²⁸Th is observed in the near-surface sediments (Cai et al., 2014).

In this study, we take advantage of 224 Ra/ 228 Th disequilibrium to quantify benthic fluxes of dissolved inorganic carbon (DIC) and nutrients $(NO_3^- + NO_2^-, NH_4^+)$ in the Pearl River Estuary (PRE), southern China. DIC and nutrients are intimately involved in the biogeochemical processes in estuaries. As a consequence of diagenetic reactions, DIC and NH₄⁺ concentrations are generally significantly elevated in sediment interstitial waters as compared to the overlying water column. In contrast, nitrate is generally depleted in anoxic interstitial waters due to a process termed denitrification. These biogeochemical reactions produce distinct chemical compositions in interstitial waters, making bottom sediments a potential source (or sink) of DIC and nutrients in estuarine waters. As such, there is a need to understand solute transport processes occurring at the sediment-water interface and to assess quantitatively, the role of interstitial water exchange in the water column budget of these constituents. As to the PRE, previous investigations have revealed that water column DIC and NH⁺₄ concentrations in the upper estuary can be up to ~ 3000 and $\sim 800 \,\mu\text{M}$, respectively (Dai et al., 2006, 2008; Guo et al., 2008). However, the source of such high levels of DIC and NH₄⁺ remains to be identified, and the role of bottom sediments in the water column budget of these constituents is yet to be evaluated.

2. SAMPLING AND ANALYSIS

2.1. Study area

The Pearl River is the 13th largest river in the world with a water discharge of $\sim 330 \times 10^9$ m³ y⁻¹ (Dai et al., 2014). It delivers $\sim 80 \times 10^6$ tons of sediments annually, most of which is believed to be trapped within the estuary (Liu et al., 2009). The river discharges to the South China Sea via three estuaries, Huangmaohai, Modaomen, and Lingdingyang, the last of which are traditionally referred to as the Pearl River Estuary (PRE). The PRE is a north–south bell-shaped estuary, with an axial length of ~ 70 km (Fig. 1). It has a narrow head of only several kilometers in the upper estuary and a relatively wide seaside entrance of ~ 50 km in the lower estuary. Water depth increases from



Fig. 1. Map of the Pearl River Estuary. Sampling stations are indicated by solid triangles.

 \sim 2.8 m in the upper estuary to \sim 20 m in the lower estuary. Tides are mainly semi-diurnal (M2) and diurnal (K1) around the PRE region. Tide amplitudes are typically 1– 1.5 m within the estuary. Tides form a counter-clockwise tidal residual circulation and may affect the estuarine circulation on both tidal and sub-tidal frequencies (Mao et al., 2004). With the seaward freshwater discharge from the river, they lead to a gravitational circulation within the estuary. Consequently, the PRE is classified as a salt wedge estuary in the wet season (Lu and Gan, 2015).

The seabed of the PRE is dominated by fine-grained sediments. In the river mouth area, the bottom sediments are relatively coarse, but the mid- and lower estuary region contains a mixture of silt and clay (He et al., 2010; Zhang et al., 2013). Mollusca and polychaetes are the main benthic fauna in the PRE. Mollusca were identified to be the dominant species in spring and autumn. Based on the surveys conducted during 1980–2000, the average biomass of the benthic fauna varied between 7.4 and 30.1 g/m². Spatial variations in the biomass of the benthic fauna, however, were found to be large in this region. High biomass was generally observed in the mid-estuary region, which can be >80 g/m² (Huang et al., 2002).

2.2. Sample collection

Sediment and seawater samples were collected during a cruise to the PRE from 16 to 27 November 2013 (dry season) onboard R/V "Tianlong". Sampling stations are indicated in Fig. 1. Detailed information about the sampling stations is provided in Appendix T1. A total of 6 stations were occupied along the salinity gradient in the PRE. Sediment cores were taken from a standard box corer $(20 \times 20 \times 60 \text{ cm})$ and were checked visually to ensure that the interface was undisturbed. Overlying seawater samples (41) were collected from the box corer and were filtered

using a 142-mm 0.7 µm (nominal pore size) GFF filter. The filtrate and the particulate samples were obtained to determine seawater 224 Ra activity and to characterize the disequilibrium between 224 Ra and 228 Th in suspended particles. Sediment sub-samples were collected by inserting PVC tubes with a diameter of 47 mm into the bulk sediment core, and were subsequently analyzed for porosity, grain size, ²²⁴Ra, ²²⁸Th, as well as ²³⁴Th in bulk sediments. Pore water was extracted from independent sediment sub-cores with a diameter of 65 mm using a Rhizon sampling device (Seeberg-Elverfeldt et al., 2005). Aliquots of \sim 15–20, \sim 5–10, and \sim 10 ml were extracted separately for the analyses of ²²⁴Ra, DIC, and nutrients. In addition to sediment samples, surface seawater samples for the analyses of DIC and nutrients were collected with a higher spatial resolution using Niskin bottles attached on a CTD rosette sampler.

2.3. ²²⁴Ra, ²²⁸Th, and ²³⁴Th analyses

²²⁴Ra and ²²⁸Th in bulk sediment/suspended particles were determined following the method described in Cai et al. (2012). In brief, the sediment sub-cores were sliced into 1-cm thick slabs immediately after sample collection. Milli-O water was added to sediment slabs to form a slurry. Subsequently, KMnO₄ and MnCl₂ solutions were added to form a suspension of MnO₂, which serves to absorb dissolved ²²⁴Ra in interstitial water. The MnO₂ suspension and the sediment with absorbed ²²⁴Ra and ²²⁸Th were filtered onto a 142-mm 0.7 µm (nominal pore size) GFF filter. The filter was placed onto a sample holder specified for sediment samples and counted for 4-6 h in a delayed coincidence counting system (the RaDeCC system). About 8-10 d later, the sample was re-counted using a same RaDeCC system. In order to verify the performance of the system, a third measurement was conducted \sim 25 d after sample collection. ²²⁴Ra and ²²⁸Th activities can be calculated either from the first and second measurements or from the first and third measurements. The RaDeCC system is calibrated with a ²³²U-²²⁸Th standard using the method of standard addition. It must be stressed that ²²⁴Ra and ²²⁸Th measurements based on this method represent the exchangeable ²²⁴Ra fraction and the production rate of exchangeable ²²⁴Ra in bulk sediment. For convenience, we will refer to these measurements as total ²²⁴Ra and ²²⁸Th activities as opposed to pore water ²²⁴Ra measurements described below.

Pore water was extracted from two parallel sediment sub-cores (area = 33.2 cm^2) by inserting an array of Rhizon samplers directly into the sediment. Approximately 15– 20 ml of pore water was retrieved at discrete depths in the sediment. The pore water sample was transferred into a 250 ml Teflon beaker and 150 ml of Milli-Q water was added. With addition of concentrated NH₃·H₂O, the pH was adjusted to 8.0–9.0. Subsequently, 5.0 ml of KMnO₄ solution (3.0 g l⁻¹) and 5.0 ml of MnCl₂ solution (8.0 g MnCl₂·4H₂O l⁻¹) were added to form a suspension of MnO₂. The MnO₂ suspension was then filtered onto a 142-mm 0.7 µm GFF filter and counted in the RaDeCC system for ~10 h, which would generally result in a minimum of 100 counts in the channel 220. About 25 d later, a second measurement was performed to determine the background count rate of the sample due to 228 Th. Overlying seawater 224 Ra activity was determined in a manner similar to the measurement of pore water 224 Ra. The counting efficiency of 224 Ra is calibrated with a 232 U $^{-228}$ Th standard that was prepared in the same manner as the sample.

Excess ²³⁴Th (²³⁴Th_{ex}) in the sediment was acquired from two parallel measurements. We have described our protocol most recently in Cai et al. (2014). It involves leaching of ²³⁴Th from an initial sediment sample with a hot solution of 6 N HCl + H₂O₂, and purification of ²³⁴Th based on the classical ion-exchange chemistry. ²³⁴Th was co-precipitated with MnO₂ and determined by counting its daughter ^{234m}Pa on a gas-flow proportional low-level beta counter (GM-25-5, RISØ National Laboratory, Denmark). About 5–6 months after sample collection, a parallel sediment sample was processed and counted in the same manner as the initial sample. The overall yields of the procedure were determined using the method of standard addition.

2.4. DIC, nutrients, and TSM analyses

DIC samples were preserved with saturated HgCl₂. Within 2 weeks after sample collection, DIC was determined by Apollo Dissolved Inorganic Carbon Analyzer. This method has a precision of 0.1–0.2% (Cai et al., 2004; Guo et al., 2008). Certified reference materials from A. G. Dickson of Scripps Institution of Oceanography were used for DIC calibration.

Nutrient samples were stored at -20 °C until analysis except for NH₄⁺, which was measured onboard with the indophenol blue spectrophotometric method (Dai et al., 2008). NO₃⁻ and NO₂⁻ measurements were based on the classical colorimetric methods and were conducted in our land-based laboratory at Xiamen University using a Four-channel Continuous Flow Technicon AA3 Auto-Analyzer (Bran-Lube GmbH). The detection limits for NO₃⁻, NO₂⁻, and NH₄⁺ were 0.07, 0.02, and 0.16 μ M, respectively.

For the determination of total suspended matter (TSM) concentrations, approximately 250–500 ml of seawater was filtered through a pre-weighed Nuclepore filter (0.45 μ m). The filter was rinsed and dried at 60 °C until a constant weight was reached. The concentration of TSM was calculated from the weight difference between the sample and the filter.

3. RESULTS

Total and pore water ²²⁴Ra, ²²⁸Th, ²³⁴Th_{ex} activities as well as pore water DIC and nutrient concentrations in the near-surface sediments of the PRE are listed in Appendix T1. Sediment porosity, bottom water temperature and salinity as well as TSM at each station are also presented. Sediment ²³⁴Th_{ex} activity was calculated from the two parallel measurements and reported with a propagated uncertainty that includes the two parallel countings of

²³⁴Th, the error of detector calibration, and the standard error of the overall yields. ²²⁴Ra and ²²⁸Th activities are reported with an error that was propagated from counting statistics, counter calibration, chance coincidence correction and in-growth/decay correction. We have compared the first and second measurements $(^{228}Th_{1,2})$ and the first and third measurements $({}^{228}Th_{1,3})$ (Fig. 2). The results show that the mean ratio of ${}^{228}Th_{1,3}/{}^{228}Th_{1,2}$ is 1.01 ± 0.02 (1SD, n = 66), not significantly different from unity. In addition, the standard deviation is consistent with the counting statistics of an individual measurement of ²²⁸Th $(\pm 2-3\%)$. This suggests that counting statistics is the predominant source of error for the ²²⁴Ra and ²²⁸Th measurements. It must be noted that the final ²²⁸Th activities are averages of 228 Th_{1,2} and 228 Th_{1,3}. As such, we are confident that our method is very good and the difference between 224 Ra and 228 Th activities as shown in Fig. 3 is a real reflection of the deviation of 224 Ra relative to 228 Th in estuarine sediments.

3.1. Distributions of ²²⁴Ra and ²²⁸Th

Depth profiles of pore water ²²⁴Ra, total ²²⁴Ra, and ²²⁸Th in the upper 0–15 cm sediment within the PRE are presented in Fig. 3. Pore water ²²⁴Ra activity varied between 0.0011 \pm 0.0014 and 0.056 \pm 0.005 dpm ml⁻¹, which is ~1–2 orders of magnitude higher than ²²⁴Ra activity in the overlying seawater. On average, however, it accounted for only 1.1% of the total desorbable ²²⁴Ra activity was low at the river mouth, where it constituted only ~0.2% of the total ²²⁴Ra activity. This manifests the dramatically different geochemical characteristics of ²²⁴Ra is bound strongly onto



Fig. 2. Comparison of ²²⁸Th activities based on the first and second measurements (²²⁸Th_{1,2}), and on the first and third measurements (²²⁸Th_{1,3}). The mean ratio of ²²⁸Th_{1,3}/²²⁸Th_{1,2} is 1.01 ± 0.02 (1SD, n = 66).

particle surfaces; however, as the ionic strength increases during mixing into seawater, desorption occurs and some ²²⁴Ra is released (Swarzenski et al., 2003). Away from the river mouth, vertical profiles of pore water ²²⁴Ra generally exhibited a maximum at the depth of 6-8 cm. Total ²²⁴Ra activity in sediments ranged from 0.75 ± 0.03 to 3.20 \pm 0.11 dpm g⁻¹. As ²²⁴Ra is produced by ²²⁸Th in sediments, vertical profiles of total ²²⁴Ra generally showed a depth pattern similar to ²²⁸Th. Sediment ²²⁸Th activity fell in the range of $0.97 \pm 0.03 - 3.34 \pm 0.08$ dpm g⁻¹, much higher than those observed in the Yangtze River Estuary (Cai et al., 2014). This could reflect the difference in sediment mineralogy and particle size between the two major estuaries in China. There was a general trend of decreasing ²²⁸Th activity in the surface sediment from the inner to the outer estuary. Depth profiles of ²²⁸Th collected from St. P03, A01, A04, and possible A09 showed a minimum at a certain depth (Fig. 3). Below this depth layer, ²²⁸Th activity was relatively constant with depth. This depth pattern could be a result of the combination of 228 Th supply from the overlying water column via sinking particles coupled with a loss of ²²⁸Ra from the upper sediment column. Depth profiles of sediment ²²⁸Th collected from A04 and A06 showed a minimum in the surface sediment. This feature has also been observed in the Yangtze River Estuary and was ascribed to lateral transport of bottom sediment across the estuary (Cai et al., 2014).

As shown in Fig. 3, total ²²⁴Ra was generally in deficit with respect to ²²⁸Th in the near-surface sediments. The most marked deficit of total ²²⁴Ra occurred in the midsalinity region, where total ²²⁴Ra/²²⁸Th activity ratios (ARs) can be as low as ~0.40. Away from the midestuary, the depletion of total ²²⁴Ra was less prominent. At the river mouth, ²²⁴Ra deficit was confined in the upper 0–4 cm sediment; below this layer total ²²⁴Ra approached secular equilibrium with ²²⁸Th. In the high-salinity region, the total ²²⁴Ra/²²⁸Th AR in the sediment was generally >0.90.

Total ²²⁴Ra and ²²⁸Th activities in suspended particles ranged from 0.42 ± 0.04 to 6.09 ± 0.23 dpm g⁻¹, and from 0.79 ± 0.04 to 5.54 ± 0.21 dpm g⁻¹, respectively. At the river mouth, ²²⁴Ra was in slight excess relative to ²²⁸Th (Fig. 4). In the mixing zone, however, the 224 Ra/ 228 Th AR in suspended particles was well below 1, reflecting a marked depletion of ²²⁴Ra. It fell in the range of 0.37-0.74, with an average of 0.53. This reveals that about 47%of the exchangeable 224 Ra in suspended particles was released during mixing into seawater. Seawater ²²⁴Ra activity ranged from 22.6 ± 1.6 to 98.9 ± 3.9 dpm $100 \, 1^{-1}$, and showed a maximum in the mid-salinity region (Fig. 4). This mode of seawater ²²⁴Ra vs. salinity is a common feature in estuaries. Finally, it is important to note that high seawater ²²⁴Ra activity in the mid-salinity region coincides with the large deficit of total ²²⁴Ra observed in bottom sediments.

3.2. Excess ²³⁴Th in the sediment

Excess ²³⁴Th activity (²³⁴Th_{ex}) was evident in the upper 0–3 cm sediment only at A01 and A09. ²³⁴Th_{ex} activity in the surface 0–0.5 cm sediment ranged from 1.27 ± 0.12 to



Fig. 3. Depth profiles of dissolved 224 Ra (filled triangle), total 224 Ra (open square), 228 Th (filled square) in the upper 0–15 cm sediment. Note that different coordinate scales are used in the upper panel and the lower panel to highlight the deficit of 224 Ra.



Fig. 4. Distributions of seawater 224 Ra activity (filled triangle) and 224 Ra/ 228 Th ratio on suspended particles (open square) along the salinity gradient in the Pearl River Estuary.



Fig. 5. Depth profiles of sediment 234 Th_{ex} collected at St. A01 (open square) and A09 (filled square).

 1.60 ± 0.16 dpm g⁻¹. The ²³⁴Th_{ex} profiles showed a decreasing excess with depth (Fig. 5). This must be ascribed to a balance of decay and penetration of ²³⁴Th_{ex} into deeper sediment subsequent to its deposition over the seafloor. With a steady-state assumption, sediment mixing rates can be inferred from the depth profiles of ²³⁴Th_{ex} (e.g., Aller and Cochran, 1976).

3.3. Distributions of DIC and nutrients

Depth profiles of pore water DIC, nitrate + nitrite (hereafter simply referred to as NO_3^-), and NH_4^+ in the upper 0-15 cm sediment are illustrated in Fig. 6. DIC and NH₄⁺ contents varied from 1208 to 19,500 µM, and from 33 to 5410 µM, respectively. There was a general trend of decreasing DIC and NH₄⁺ concentrations in pore water from the inner to the outer estuary. DIC and NH⁺₄ normally increased with depth in the sediment. In contrast, NO_3^- was depleted within the upper $\sim 3 \text{ cm}$ sediment. In the inner estuary, depth profiles of pore water DIC and NH_4^+ were characterized by sharp concentration gradients. In comparison, the concentration gradients of pore water DIC and NH_4^+ were much less prominent in the mid- and outer estuary. Notably, the river core had an unusual low C:N ratio of \sim 3.2 in pore water below 1 cm sediment. This value is remarkably close to the C:N ratio of 3.3 that is characteristic of a process termed methanogenesis. Indeed, previous studies have shown that methane concentration in the surface water of the upper PRE can be as high as >2000 nM (e.g., Chen et al., 2008). This suggests that methane fermentation may dominate the organic break down in sediments at this site. From Fig. 6, one can also see DIC and NH₄⁺ minima in pore water, which were probably a result of physical and/or bio-irrigation processes in the sediment.

 NO_3^- and NH_4^+ concentrations in the surface waters varied from 6.9 to 258 μ M, and from <0.25 to 414 μ M, respectively. High NO_3^- and NH_4^+ contents were observed at the river mouth. Surface NO_3^- concentration decreased steadily during mixing into the open ocean, whereas NH_4^+ was efficiently removed close to the river mouth (Fig. 7).

4. DISCUSSION

4.1. Adsorption-desorption kinetics of ²²⁴Ra

4.1.1. The diagenetic model for ²²⁴Ra in sediments

In order to better utilize 224 Ra/ 228 Th disequilibrium as a proxy of solute transport processes across the sediment–water interface, it is essential to understand the reaction kinetics of 224 Ra in sediments. As we know, some 224 Ra produced by decay of 228 Th may enter the surrounding interstitial water. This soluble 224 Ra is then free to diffuse and, as a consequence, it migrates upward and outward resulting in a deficiency of total 224 Ra with respect to 228 Th in the upper portions of the sediments. A diagenetic model for the distributions of dissolved and adsorbed 224 Ra in sediments of the PRE can be constructed:

$$\frac{\partial(\phi \mathbf{R} \mathbf{a}_{d})}{\partial t} = \frac{\partial(\phi D_{\mathbf{S}} \frac{\partial \mathbf{R} \mathbf{a}_{d}}{\partial z})}{\partial z} + \frac{\partial\left(D_{\mathbf{B}} \frac{\partial(\phi \mathbf{R} \mathbf{a}_{d})}{\partial z}\right)}{\partial z} - \mathbf{I}_{\mathbf{R} \mathbf{a}} - \omega \frac{\partial(\phi \mathbf{R} \mathbf{a}_{d})}{\partial z} + (1 - \phi)\rho_{s}\mathbf{S} - \phi\lambda_{\mathbf{R} \mathbf{a}}\mathbf{R} \mathbf{a}_{d} \qquad (1)$$

$$\frac{\partial [(1-\phi)\rho_{s}\mathbf{R}\mathbf{a}_{s}]}{\partial t} = \frac{\partial \left(D_{B}\frac{\partial [(1-\phi)\rho_{s}\mathbf{R}\mathbf{a}_{s}]}{\partial z}\right)}{\partial z} - \omega \frac{\partial [(1-\phi)\rho_{s}\mathbf{R}\mathbf{a}_{s}]}{\partial z} - (1-\phi)\rho_{s}\mathbf{S} + (1-\phi)\rho_{s}\lambda_{\mathrm{Ra}}(\mathrm{Th} - \mathrm{Ra}_{s})$$
(2)

where Ra and Th are ²²⁴Ra and ²²⁸Th activities in a unit of $dpm g^{-1} dry mass$, subscript d and s denote activities in pore water and on sediment particles; Note that Rad is defined as the concentration of Ra in dpm per volume of pore water; ϕ and ρ_s represent the sediment porosity and density; $D_{\rm S}$ is the diffusivity of ²²⁴Ra in the sediment, $D_{\rm B}$ denotes the sediment mixing coefficient, ω is the sedimentation rate, and λ_{Ra} is the decay constant of ²²⁴Ra; I_{Ra} represents the rate at which 224 Ra is transported out of the sediment by irrigation, and S is the net release rate of ²²⁴Ra from sediment particles into the surrounding interstitial water. For the ²²⁴Ra distribution in marine sediments, the advective burial of sediment is generally a negligible term (e.g., Hancock et al., 2000). With the assumption of constant ρ_s and D_B values with depth, the formulations for the steady-state distribution of ²²⁴Ra in the sediment are written as:

$$\frac{\partial \left(\phi D_{\mathbf{S}} \frac{\partial \mathbf{R} \mathbf{a}_{d}}{\partial z}\right)}{\partial z} + \frac{\partial \left(D_{\mathbf{B}} \frac{\partial \left(\phi \mathbf{R} \mathbf{a}_{d}\right)}{\partial z}\right)}{\partial z} - \mathbf{I}_{\mathbf{R} \mathbf{a}} + (1 - \phi) \rho_{s} \mathbf{S} - \phi \lambda_{\mathbf{R} \mathbf{a}} \mathbf{R} \mathbf{a}_{d} = 0$$
(3)

$$\frac{\partial \left(D_{\rm B} \frac{\partial \left[(1-\phi) Ra_{\rm s} \right]}{\partial z} \right)}{\partial z} - (1-\phi) \mathbf{S} + (1-\phi) \lambda_{\rm Ra} ({\rm Th} - {\rm Ra}_{\rm s}) = 0$$
(4)

In the above formulations, $D_{\rm S}$ is estimated from the molecular diffusion coefficient for radium in seawater (*D*) using the relation $D_{\rm S} = D/\theta^2$, where θ denotes the sediment tortuosity and is derived from the porosity using the expression $\theta^2 = 1-2\ln(\phi)$ (Boudreau, 1997). $D_{\rm B}$ is inferred from



Fig. 6. Depth profiles of pore water DIC, $NO_2^- + NO_3^-$, and NH_4^+ in the upper 0–15 cm sediment. Note that different coordinate scales are used in each plot to highlight the gradient of concentration.



Fig. 7. Distributions of $NO_2^- + NO_3^-$ (open square) and NH_4^+ (filled square) in the surface water along the salinity gradient in the Pearl River Estuary.

the distributions of excess ²³⁴Th in the upper sediment column. We derived a $D_{\rm B}$ value of 0.006 and 0.014 cm² d⁻¹ for station A01 and A09, respectively. For the other stations where excess ²³⁴Th was not detectable in the near-surface sediment, an average $D_{\rm B}$ value of 0.010 cm² d⁻¹ was used to assess the role of sediment mixing in the mass balance of ²²⁴Ra. The second derivatives $\frac{\partial \left(\phi D_{\rm S} \frac{\partial (\theta, q_{\rm S})}{\partial z}\right)}{\partial z}$, $\frac{\partial \left(D_{\rm B} \frac{\partial (\theta, q_{\rm S})}{\partial z}\right)}{\partial z}$, and $\frac{\partial \left(D_{\rm B} \frac{\partial (1-\phi)({\rm Ras})}{\partial z}\right)}{\partial z}$ are computed from the depth distributions of dissolved and adsorbed ²²⁴Ra in sediments using a numerical approach similar to that described in Ku et al. (1995). With this knowledge, the terms I_{Ra} and S can be calculated from Eqs. (3) and (4) for any specific depth in a sediment column.

4.1.2. Estimation of adsorption rate constants of ²²⁴Ra

Recoil ejection accompanying its production by alpha decay in the solid particles has been recognized as a dominant source of short-lived uranium- and thorium-series radionuclides (such as ²²²Rn and ²²⁴Ra) in groundwater (e.g., Kigoshi, 1971; Krishnaswami et al., 1982). In addition to recoil ejection, adsorption and desorption of a radionuclide generally take place over a time scale of minutes and thus are important processes that affect the partition of ²²⁴Ra between the aqueous phase and solid particles (e.g., Krishnaswami et al., 1982; Luo et al., 2000). If we assume that the kinetics of adsorption and desorption of ²²⁴Ra are both a first-order process, then the term S in Eqs. (3) and (4) can be expressed as:

$$(1 - \phi)\rho_{s}\mathbf{S} = (1 - \phi)\rho_{s}\mathbf{P} + (1 - \phi)\rho_{s}k_{-1}\mathbf{R}\mathbf{a}_{s}$$
$$-\phi k_{1}\mathbf{R}\mathbf{a}_{d}$$
(5)

where P denotes the ejection rate of 224 Ra to the aqueous phase by recoil; k_1 and k_{-1} represent the adsorption and desorption rate constant of 224 Ra. In order to determine k_1 and k_{-1} from Eq. (5), P and activities of two radium isotopes (e.g., 224 Ra $^{-226}$ Ra or 224 Ra $^{-228}$ Ra) must be known (Krishnaswami et al., 1982; Luo et al., 2000). Unfortunately, activities of a longer-lived radium isotope were not determined in this study. Nonetheless, we can use the measurements of ²²⁴Ra and ²²⁸Th on suspended particles to place a constraint on the supply terms of ²²⁴Ra in sediment particles. As mentioned in Section 3.1, in the mixing zone of the PRE the total ²²⁴Ra activities were considerably lower than the ²²⁸Th activities on suspended particles. We may assume that the deficiency of ²²⁴Ra on suspended particles was solely caused by the recoil ejection plus desorption of ²²⁴Ra into seawater. Adsorption of dissolved ²²⁴Ra onto suspended particles is supposed to be essentially zero, because of the low ²²⁴Ra concentration and the high ionic strength in the water column. Consequently, the entire ²²⁴Ra measurement on suspended particles represents the emanation rate (i.e., the ejection rate and the desorption rate). If we assume that the relative emanation rate of ²²⁴Ra on sediment particles is similar to that on suspended particles in the overlying seawater, then Eq. (5) can be applied to sediment particles and takes the form:

$$(1-\phi)\rho_{s}\mathbf{S} = (1-\phi)\rho_{s}\lambda_{Ra}(1-R_{s})\mathrm{Th} - \phi k_{1}\mathrm{Ra}_{d}$$
(6)

where R_s denotes the ²²⁴Ra/²²⁸Th AR for the sediment, as determined from suspended particles in the overlying seawater. With Eq. (6), knowledge of k_1 can be derived for any specific depth at a sediment column. For the station located at the river mouth (St. P03), the total ²²⁴Ra activity was found to be in slight excess with respect to ²²⁸Th on suspended particles (Fig. 4). This excess is considered to be a result of an increase in k_1 value in oxic freshwater. As such, we assume that the relative emanation rate of ²²⁴Ra on suspended particles and sediment particles at this location is also similar to the rate on suspended particles in the mixing zone. An average R_s AR of 0.53 is thus assigned so as to estimate the adsorption rate constant of ²²⁴Ra (k_1) at St. P03.

In the application of Eq. (6), however, it is important to note that suspended particles are probably finer-grained and the fraction of ²²⁴Ra emanated may represent an upper limit of sediment particles. In such a case, k_1 would be overestimated. On the contrary, if R_s is smaller for sediment particles due to larger grain size, then k_1 would be underestimated. Despite of this limitation, the application of Eq. (6) would provide a first approximation of k_1 values of ²²⁴Ra. The results show that for suspended particles in freshwater, k_1 is equal to $2000 \pm 400 \,\mathrm{d^{-1}}$, or 1.4 $\pm 0.3 \text{ min}^{-1}$. This value is in agreement with previous results in oxic groundwater, which indicated that sorption of radium takes place on a time scale of minutes or less (e.g., Krishnaswami et al., 1982; Luo et al., 2000). For the sediment particles at the river mouth, k1 varied between 25 ± 6 and $90 \pm 70 \text{ d}^{-1}$ (see Table 1), which is 1–2 orders of magnitude lower than the value derived for suspended particles in the overlying water. For the sediment samples collected in the mixing zone, k_1 was even lower, ranging from 0.1 ± 1.1 to $34 \pm 4 d^{-1}$. This reflects the decrease of the ²²⁴Ra adsorption rate constant in high ionic strength seawater. We do not know what causes the difference in k_1 between the suspended particles and sediment particles at the river mouth. One plausible explanation may be related to the different redox conditions in the sediment column and in the overlying water. In comparison to the oxic

the Fearl A	River Estuary.					
Station	$^{*}k_{1}$ (d ⁻¹)	${}^{*}R_{\mathrm{f}}$	$F_{\rm Ra}~({ m dpm}{ m cm}^{-2}~{ m d}^{-1})$	$F_{\rm M} ({\rm dpm} {\rm cm}^{-2} {\rm d}^{-1})$	$F_{ m B}~({ m dpm~cm^{-2}~d^{-1}})$	$F_{ m g}~({ m dpm}~{ m cm}^{-2}~{ m d}^{-1})$
P03	$25\pm 6-90\pm 70~(48\pm 22)$	$340\pm90{-}2400\pm600~(710\pm660)$	0.125 ± 0.070	0.0004 ± 0.0001	I	0.125 ± 0.070
A01	$3.4\pm0.9{ extrm{}34\pm4}~(12\pm10)$	$58 \pm 5 - 320 \pm 30 \; (130 \pm 80)$	0.562 ± 0.081	0.0048 ± 0.0005	-0.0001 ± 0.0004	0.557 ± 0.081
A03	0.1 ± 1.1 – $7.4\pm1.3~(4.0\pm2.7)$	$35\pm3{-}110\pm10~(80\pm26)$	0.652 ± 0.046	0.0070 ± 0.0006	0.0006 ± 0.0007	0.644 ± 0.046
A04	0.9 ± 0.5 – $5.7\pm2.0~(2.5\pm1.4)$	$26\pm2{-}190\pm20~(99\pm54)$	0.656 ± 0.063	0.0040 ± 0.0004	0.0130 ± 0.0011	0.639 ± 0.063
A06	$4.5 \pm 0.6 ext{-}9.8 \pm 1.2 \; (7.4 \pm 1.8)$	$40 \pm 3{-}97 \pm 12~(70 \pm 17)$	0.201 ± 0.054	0.0063 ± 0.0005	0.0077 ± 0.0006	0.187 ± 0.054
A09	$5.2 \pm 0.9 - 16.8 \pm 2.0 \ (8.0 \pm 3.5)$	67 ± 7 -140 $\pm 15 (97 \pm 20)$	0.260 ± 0.044	0.0093 ± 0.0006	0.0086 ± 0.0009	0.242 ± 0.044

Adsorption rate constant (k_1) , retardation factor (R_f) , observed flux of 224 Ra (F_{Ra}) , theoretical flux of 224 Ra induced by molecular diffusion (F_M) , by sediment mixing (F_B) , and by irrigation (F_g) in

Table 1

overlying water, estuarine sediments are generally anoxic so that manganese oxides (MnO₂), a major carrier of ²²⁴Ra, may be actively reduced to dissolved Mn²⁺, thereby lowering the k_1 value of ²²⁴Ra in sediments.

4.1.3. Calculation of partition coefficients and retardation factors of 224 Ra

Direct measurements of pore water 224 Ra and exchangeable 224 Ra on sediment particles also allow us to estimate the partition coefficient (*K*) and the retardation factor (*R*_f) for 224 Ra. These parameters are commonly expressed as:

$$K = \frac{\rho_{\rm s}(1-\phi)\mathbf{R}\mathbf{a}_{\rm s}}{\phi\mathbf{R}\mathbf{a}_{\rm d}}; \text{ and } R_{\rm f} = 1+K \tag{7}$$

Results of the calculation show that values of $R_{\rm f}$ for sediments at the river mouth varied between 340 ± 90 and 2400 ± 630 , with an average of 710 (see Table 1). In comparison, values of $R_{\rm f}$ for sediments in the mixing zone are much lower, ranging from 26 ± 2 to 320 ± 30 . The average of all sediment samples collected in the mixing zone gives a $R_{\rm f}$ value of 96. The lower $R_{\rm f}$ values indicate that ²²⁴Ra is more mobile in saline sediments than in fluvial sediments. Notably, the $R_{\rm f}$ values in this study are two orders of magnitude lower than previous results reported for oxic groundwaters (e.g., Krishnaswami et al., 1982; Luo et al., 2000). This may reflect the combined effect of redox conditions and the ionic strength on the mobility of ²²⁴Ra in estuarine sediments. Nonetheless, our results are in good agreement with an $R_{\rm f}$ estimate of 112 that is inferred from a study of ²²²Rn emanation rate (i.e., ²²⁶Ra activity) in sandy sediments in Catalina Harbor, California (Colbert et al., 2008). Also, they are not inconsistent with laboratory determinations of K_d for Ra, which is defined as the ratio of Ras to Rad. Serene and Relyea (1982) reported that minimum values of K_d for Ra in granites, basalts, and tuffs are 50, 50, and 200 ml/g, respectively. If the porosity of sediment is 0.70, then the above K_d values give values of R_f in the range of 60 and 200, similar to the range of $R_{\rm f}$ determined using in situ ²²⁴Ra measurements.

4.2. Benthic fluxes of ²²⁴Ra

Outward fluxes of 224 Ra from bottom sediments in the PRE can be estimated using a one-dimensional (1D) mass balance exchange model (Cai et al., 2012). This model assumes that in-situ production via alpha decay of 228 Th is the sole source of 224 Ra in the sediment. At steady state, a mass balance consideration of 224 Ra in the sediment gives:

$$F_{\rm Ra} = \int_0^\infty \lambda_{224} (A_{\rm Th} - A_{\rm Ra}) dz \tag{8}$$

where F_{Ra} (dpm cm⁻² d⁻¹) represents the flux of ²²⁴Ra ("+" upward) across the sediment–water interface. A_{Th} and A_{Ra} are ²²⁸Th and ²²⁴Ra activities in bulk sediment (unit: dpm cm⁻³). In practice, we assume that ²²⁴Ra and ²²⁸Th reach secular equilibrium below the sampling depth, i.e., 15 cm in this case. F_{Ra} was then calculated by trapezoidal integration of the ²²⁴Ra deficit from 0 to 15 cm depth in the sediment. Results of the calculation show that outward fluxes of ²²⁴Ra from bottom sediments varied between 0.125 ± 0.070 and 0.656 ± 0.063 dpm cm⁻² d⁻¹ in our study region (Table 1). In addition, ²²⁴Ra fluxes exhibited a plateau in the midsalinity region. This pattern coincides with the mode of seawater ²²⁴Ra vs. salinity in the water column, and suggests that pore water exchange could be a predominant source of the seawater ²²⁴Ra in the PRE.

Like any dissolved species in sediment interstitial waters, outward fluxes of ²²⁴Ra from bottom sediments are regulated by a variety of processes that operate at the sediment-water interface. Predominant processes include molecular diffusion, sediment mixing, and irrigation (e.g., Berner, 1980). The relative importance of these processes in regulating the ²²⁴Ra flux has been evaluated by modeling the sediment ²²⁴Ra profiles collected in the Yangtze River Estuary using the general diagenetic equation (Cai et al., 2014). A more direct approach, however, is to compare the theoretical fluxes induced by molecular diffusion and sediment mixing with the observed flux of ²²⁴Ra. Given that the concentration gradient at the sediment-water interface can be computed from the depth profiles of pore water ²²⁴Ra, we are thus able to calculate the fluxes of ²²⁴Ra induced by molecular diffusion and sediment mixing using Fick's first law:

$$F_{\rm M} = \phi D_{\rm S} \frac{\partial R a_{\rm d}}{\partial z} \Big|_{z=0}$$
, and $F_{\rm B} = D_{\rm B} \frac{\partial [\phi R a_{\rm d} + \rho_{\rm s}(1-\phi)R a_{\rm s}]}{\partial z} \Big|_{z=0}$
(9)

where $F_{\rm M}$ and $F_{\rm B}$ denote the theoretical fluxes of $^{224}{\rm Ra}$ induced by molecular diffusion and by sediment mixing, respectively. The concentration gradients $\frac{\partial Ra_d}{\partial z}$ and $\frac{\partial [\phi Ra_d + \rho_s(1-\phi)Ra_s]}{\partial z}$ at the sediment-water interface, i.e., z = 0, are computed from the measurements in the overlying water and at the layer of 0-1 cm sediment. In regard to the 224 Ra flux induced by irrigation (F_g), we assume that it is equivalent to the difference between the observed flux derived from ²²⁴Ra deficit and the theoretical fluxes caused by molecular diffusion plus sediment mixing. Results of the calculation show that values of $F_{\rm M}$ varied between 0.0004 \pm 0.0001 and 0.0093 ± 0.0006 dpm cm⁻² d⁻¹ (Table 1), which on average accounted for only $\sim 1.6\%$ of the observed fluxes of 224 Ra. In comparison, values of $F_{\rm B}$ ranged from -0.0001 ± 0.0004 to 0.0130 ± 0.0011 dpm cm⁻² d⁻¹, and were responsible for 0.0-3.8% of the measured fluxes of 224 Ra. Thus, molecular diffusion and sediment mixing can explain only <5% of the measured deficit of 224 Ra. This implies that the process of irrigation must be the dominant mechanism that controls outward fluxes of ²²⁴Ra from bottom sediments in the PRE.

The importance of irrigation in regulating solute transfer across the sediment-water interface has been recognized in previous studies (e.g., Vanderborght et al., 1977; Aller, 1980; Huettel and Webster, 2001). In a recent study, we demonstrated that molecular diffusion was generally responsible for ~10% of the measured deficit of ²²⁴Ra in the Yangtze River Estuary (Cai et al., 2014), which is an order of magnitude higher than the fraction identified in the present study. However, it must be noted that in the Yangtze River Estuary, only total ²²⁴Ra activity in bulk sediment was determined. Under such a circumstance, we had to use a reference value of 11.7 for K_d so as to infer the concentration gradient of pore water ²²⁴Ra at the sediment–water interface. Had the K_d value as measured in this study been used in the Yangtze River Estuary, the contribution of molecular diffusion to the total deficit of ²²⁴Ra would have decreased from ~10% to ~2%. This value is remarkably close to the fraction determined in the PRE.

4.3. Transfer of DIC and nutrients across the sediment-water interface

In a most recent study, the ²²⁴Ra/²²⁸Th disequilibrium has been proposed as a new approach to determine the transfer rate of a dissolved species across the sediment–water interface (Cai et al., 2014). This approach, which we termed the ²²⁴Ra/²²⁸Th disequilibrium approach, is built upon the observed deficit of ²²⁴Ra in the sediment and a concept of increased surface area for exchange by irrigation. The concept was proposed by early investigators and is now increasingly accepted in the field. In this concept, exchange of a dissolved species between the sediment and the overlying water is deemed to take place at a highly invaginated interface due to irrigation. In addition, molecular diffusion is assumed to be the predominant mechanism of transport at the interface. Consequently, the benthic flux of a dissolved species *i* (*F_i*) is expressed as (Cai et al., 2014):

$$F_{i} = F_{Ra} \left(\frac{D_{S}^{i}}{D_{S}^{Ra}} \right) \left(\frac{\frac{\partial c^{i}}{\partial z}}{\frac{\partial z}{\partial z}} \right)$$
(10)

where superscript Ra and *i* denote ²²⁴Ra and the dissolved species *i*, respectively; $\frac{\partial c}{\partial z}$ is the concentration gradient of the species of interest at the interface. Note that the ratio of molecular diffusion coefficient $D_{\rm S}^i/D_{\rm S}^{\rm Ra}$ in sediments is identical to the ratio in seawater, and is constant over the temperature range 0–25 °C that is generally encountered in the sediment (Boudreau, 1997).

The performance of the 224 Ra/ 228 Th disequilibrium approach relies on our ability to estimate the flux of ²²⁴Ra, and how accurately we can characterize the concentration gradients of dissolved ²²⁴Ra and the species of interest at the sediment-water interface. An important assumption associated with this approach is that the concentration gradients across the burrow wall are identical to those across the surficial sediment-water interface (Cai et al., 2014). In this regard, it must be pointed out that pore water ²²⁴Ra, DIC, NO₃⁻ and NH₄⁺ have rather different scale lengths. A consideration of the general diagenetic equation (e.g., see Eq. (10) in Cai et al., 2014) shows that relative to DIC and NH₄⁺, pore water ²²⁴Ra has a considerably small scale length such that its largest concentration gradient takes place over a few mm across the sedimentwater interface. This difference is mainly caused by the kinetic nature of 224 Ra which, as elucidated in Section 4.1., leads to rapid adsorption of 224 Ra onto sediment particles. As such, high-resolution profiles of pore water ²²⁴Ra are critically important for the application of the ²²⁴Ra/²²⁸Th disequilibrium approach. In this study, pore water ²²⁴Ra, DIC, NO_3^- , and NH_4^+ concentrations were determined at a depth interval of 1 cm in the upper sediment column. We use the measurements in the overlying water and at the layer of 0–1 cm sediment to estimate the term $\frac{\partial c^l}{\partial z}$ and $\frac{\partial c^{Ra}}{\partial z}$. The derived concentration gradients across the sediment– water interface are listed in Table 2. In particular, we find a strong correlation of DIC vs. dissolved ²²⁴Ra and of NH₄⁺ vs. dissolved ²²⁴Ra in the upper several centimeters of sediments at some stations (Fig. 8). This level of correlation tends to support our assumption that the concentration gradients across the burrow wall are identical to those across the surficial sediment–water interface. Under such a circumstance, the benthic fluxes are most reliably determined using the ²²⁴Ra/²²⁸Th disequilibrium approach.

The benthic fluxes of DIC and nutrients were highly variable in the PRE, and in general, they spanned over a range of two orders of magnitude (Table 2). The DIC fluxes varied between -38 ± 12 and $8500 \pm 16,000 \text{ mmol m}^{-2} \text{ d}^{-1}$ ("+" upward). In comparison, the NO_3^- and NH_4^+ fluxes ranged from -5 ± 1 to $-660 \pm 1200 \text{ mmol m}^{-2} \text{ d}^{-1}$, and from 10 ± 3 to $7500 \pm 14,000 \text{ mmol m}^{-2} \text{ d}^{-1}$, respectively. Notably, all the highest fluxes were observed in the upper estuary (St. P03). Unfortunately, these estimates are associated with a very large uncertainty, which is mainly caused by the error associated with the measurement of pore water 224 Ra activity at the 0–1 cm sediment (see Appendix T1). As such, our later discussion will not be focused on these estimates. Away from the upper estuary into the mid-estuary, the fluxes diminished dramatically. It should also be noted that a negative flux of DIC was observed at St. A06, indicating a net removal of DIC from the overlying water column into bottom sediments. We do not know what caused the DIC removal at this location. Nonetheless, we have observed a lot of shells in the upper sediment column at this site. In addition, previous studies also showed that the benthic fauna at this location is characterized by high biomass of a mollusca species, Potamocorbula laevis (e.g., Huang et al., 2002). As such, we speculate that the DIC removal at this location may be caused by the formation of calcium carbonate in the top several centimeters of sediments, which could lower the DIC concentration in pore water and hence result in a negative gradient of DIC concentration across the sediment-water interface as shown in Fig. 6.

The benthic fluxes of DIC and nutrients from bottom sediments are expected to have a large impact on the biogeochemistry of the overlying water column. For a wellmixed water column, the benthic fluxes would induce a change of -4.3 ± 1.3 to $61 \pm 16 \,\mu mol \, l^{-1} d^{-1}$ for DIC, of -0.2 ± 0.0 to $-14 \pm 3 \ \mu mol \ l^{-1} \ d^{-1}$ for NO₃⁻, and of 1.2 ± 0.2 to $14 \pm 4 \ \mu mol \ l^{-1} \ d^{-1}$ for NH₄⁺ (see Table 2; "+" values denote net addition and "-" values represent net removal from the overlying water column). In the overlying water column, sources of DIC also include aerobic respiration and denitrification. In an earlier study, Guo et al. (2008) suggested that in the PRE aerobic respiration and denitrification would add DIC to the water column at a rate of 14–18, and of $\sim 4 \,\mu mol \, l^{-1} \, d^{-1}$, respectively. In the meantime, CO2 evasion would remove DIC at a rate of $12-36 \,\mu\text{mol}\,l^{-1}\,d^{-1}$. In terms of NH₄⁺, Dai et al. (2008) demonstrated that in winter, nitrification would remove NH_4^+ at a rate of 0–5.4 umol $l^{-1} d^{-1}$. At any rate, our

Table 2										
Concenti	ation gradient of dissolv	ved ²²⁴ Ra, DIC,	NO_3^- , and NH_4^+ ;	at the sediment-v	water interface, and l	benthic fluxes ("+" u	ipward) of DIC, NC	3^{-}_{3} and NH ⁺ ₄ in the	Pearl River Estua	ry. Change rate
(ð) of wa	tter column DIC, NO_3^-	and NH ⁺ ₄ due to	o pore water exci	hange are also p.	resented. Estimates	with uncertainty >1	00% are highlighted	l in italic.		
Station	$\partial \mathbf{R} \mathbf{a} / \partial z \; (\mathrm{dpm} \; \mathrm{cm}^{-4})$	$\partial DIC/\partial z$ (µmol cm ⁻⁴)	$\partial NO_3^-/\partial z$ (µmol cm ⁻⁴)	$\partial \mathrm{NH}_4^+/\partial z$ (µmol cm ⁻⁴)	DIC flux $(mmol m^{-2} d^{-1})$	NO $_{3}^{-}$ flux (mmol m ⁻² d ⁻¹)	NH_4^+ flux (mmol m ⁻² d ⁻¹)	δDIC (µmol 1 ⁻¹ d ⁻¹)	δNO_3^- (µmol 1 ⁻¹ d ⁻¹)	$\frac{\delta NH_4^+}{(\mu mol \ l^{-1} \ d^{-1})}$
P03	0.0015 ± 0.0028	7.8	-0.38	4.1	8500 ± 16000	-660 ± 1200	7500 ± 14000	940 ± 1800	-73 ± 140	<i>830</i> ± <i>1600</i>
A01	0.016 ± 0.003	2.6	-0.22	0.36	1200 ± 300	-166 ± 42	278 ± 71	61 ± 16	-8.3 ± 2.1	14 ± 4
A03	0.023 ± 0.004	0.18	-0.14	0.12	70 ± 13	-87 ± 17	77 ± 15	10 ± 2	-13 ± 2	11 ± 2
A04	0.027 ± 0.004	0.31	-0.21	0.16	103 ± 19	-111 ± 20	91 ± 16	13 ± 2	-14 ± 3	11 ± 2
A06	0.025 ± 0.004	-0.36	-0.044	0.058	-38 ± 12	-8 ± 2	10 ± 3	-4.3 ± 1.3	-0.8 ± 0.3	1.2 ± 0.4
A09	0.038 ± 0.004	0.58	-0.033	0.20	54 ± 11	-5 ± 1	31 ± 6	2.1 ± 0.4	-0.2 ± 0.0	1.2 ± 0.2



Fig. 8. Correlation of pore water DIC vs. dissolved 224 Ra in the upper 0–10 cm sediment interval at St.A04 (left panel), and of NH₄⁺ vs. dissolved 224 Ra in the upper 0–6 cm sediment interval at St. A06 (right panel). Note that measurements in the overlying water are also included in the plots.

results suggest that pore water exchange is an important process and must be considered in the mass balance of DIC and nutrients in the Pearl River estuary.

Provided with a residence time of water mass, net addition or removal of water column DIC and nutrients due to pore water exchange can be assessed using the relation:

$$\Delta i = \frac{F_i \times \tau}{H} \tag{11}$$

where Δi refers to net addition or removal of a dissolved species from the overlying water column, τ is the water residence time in the estuary. Recently, Wang (2014) has utilized a tidal prism model (e.g., Sanford et al., 1992) to estimate water residence time in the PRE. This researcher derived a value of 2.2 ± 0.2 d in the wet season, and a value of 4.0 ± 1.4 d in the dry season. If we use an average of 3.0 d as the water residence time in November, a transition season in this region, then on average pore water exchange would lead to a change of $49 \,\mu\text{mol}\,l^{-1}$ for DIC, of $-22 \,\mu\text{mol}\,l^{-1}$ for NO₃, and of 23 $\mu\text{mol}\,l^{-1}$ for NH₄⁺ in the water column of PRE. It is striking that bottom sediment removes water column NO_3^- at a rate remarkably close to the rate at which it adds NH_{4}^{+} to the water column. If all the NH_4^+ added to the water column is nitrified, then the gain and loss of water column NO_3^- would be in balance. This may explain the quasi-conservative behavior of NO_3^- during estuarine mixing as shown in Fig. 7.

Total fluxes of DIC and nutrients due to pore water exchange can be estimated by integrating the site-specific fluxes over the whole PRE. Given the large uncertainty associated with the flux estimates at P03, we excluded this point and placed the other estimates onto a five-box grid with a total area of 1180 km^2 , i.e., the area of the PRE. The choice of box dimensions was based on the estuary geometry. Box boundaries are shorelines, and demarcations that intersect the axis of the estuary. Box dimensions were determined from navigation charts. Consequently, the estuary is divided into five boxes with an area of 105, 179, 211, 383, and 302 km², respectively. The benthic fluxes as calculated from the ²²⁴Ra/²²⁸Th disequilibrium approach are multiplied by the sediment area to compute the flux from the sediments. Fluxes of DIC and nutrients at each box are added up. Assuming that our flux estimates are typical for the dry season (November-March), we would derive an annual flux of $42 \pm 6 \times 10^9$ mol yr⁻¹ for DIC, of -13 $\pm 1 \times 10^9$ mol yr⁻¹ for NO₃, and of $16 \pm 1 \times 10^9$ mol yr⁻¹ for NH_4^+ . The benthic flux of DIC is ~18% of the riverine input of DIC into the PRE, which was estimated to be $\sim 237 \times 10^9$ mol yr⁻¹ in the dry season (Guo et al., 2008). During this cruise, NO_3^- and NH_4^+ concentrations at zero salinity spanned over a range of 104-319 and 1.3-414 μ mol l⁻¹, respectively. Taking the medians as the endmember values and a discharge rate of 3600 m³ s⁻¹ (Cai et al., 2004), we would obtain a riverine flux of 9.1×10^9 mol yr⁻¹ for NO₃⁻, and of 3.1×10^9 mol yr⁻¹ for NH_4^+ in the dry season. In this regard, our results indicate that bottom sediments are a major sink of water column NO_3^- , and are a predominant source of NH_4^+ in the PRE. It is interesting, however, to note that NO_3^- concentration in the surface water exhibited a quasi-conservative behavior, whereas NH_4^+ concentration in the surface water was almost depleted once it is away from the river mouth (Fig. 7). This pattern highlights the importance of water column processes, like nitrification and biological uptake, in regulating the concentration level of NO_3^- and NH_4^+ in the PRE (e.g., Dai et al., 2006,2008).

Total flux of DIC from bottom sediments into the PRE can also be deduced from a simple consideration of the mass balance of total organic carbon (TOC). As mentioned above, the PRE receives a delivery of $\sim 80 \times 10^6$ tons of sediments annually. Previous studies in the PRE suggest that TOC content in the surface sediments decreases from $\sim 2.0-4.0\%$ at the river mouth to $\sim 1.0\%$ in the lower estuary (e.g., Yang et al., 2011). On an annual basis, if we assume that 1.0–2.0% of TOC in the discharged sediments is combusted and converted to DIC, then we would derive a total flux of DIC:

$$F_{\text{DIC}} = 80 \times 10^{12} \text{ g yr}^{-1} \times (1.0-2.0\%)/12$$

= (67-134) × 10⁹ mmolC yr⁻¹.

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Total and pore water ²²⁴Ra (T. ²²⁴Ra and P. ²²⁴Ra), ²²⁸Th, ²³⁴Th_{ex} activities as well as pore water DIC and nutrient concentrations in the near-surface sediments of the PRE.

1		() <i>// / C</i> A	1					
Depth cm	Porosity	DIC (μ mol l ⁻¹)	$NO_3^- + NO_2^- \ (\mu mol \ l^{-1})$	$\mathrm{NH}_4^+ \ (\mu mol \ l^{-1})$	P. 224 Ra (dpm ml ⁻¹)	T. 224 Ra (dpm g ⁻¹)	228 Th (dpm g ⁻¹)	²²⁴ Ra/ ²²⁸ Th A.R.	234 Th (dpm g ⁻¹)
*P03; 23° 04.4	52'N, 113° 2	28.218'E; 9.0 m; T	$r = 20.9 \circ C, S = 0.2; TSM =$	$= 48.2 mg l^{-1}$					
Bottom water		1303	190.3	250	0.00033 ± 0.00002	6.09 ± 0.23	5.54 ± 0.21	1.10 ± 0.06	
0-1	0.705	5210	17.9	2300	0.0011 ± 0.0014	2.43 ± 0.10	2.99 ± 0.11	0.81 ± 0.05	ND
1–2	0.665	13,218	6.7	3419	0.0032 ± 0.0024	2.12 ± 0.08	2.06 ± 0.08	1.03 ± 0.05	ND
2–3	0.678	15,857	4.3	3861	0.0070 ± 0.0025	2.20 ± 0.08	2.33 ± 0.08	0.94 ± 0.05	ND
3–4	0.672	15,487	3.2	5739	ND	2.14 ± 0.08	2.22 ± 0.08	0.96 ± 0.05	ND
4–5	0.681	14,752	4.7	4523	0.0069 ± 0.0025	1.90 ± 0.07	1.77 ± 0.07	1.07 ± 0.06	ND
5–6	0.663	13,527	17.0	5145	0.0043 ± 0.0023	1.62 ± 0.07	1.72 ± 0.07	0.94 ± 0.05	ND
7–8	0.646	14,011	8.4	5352	0.0046 ± 0.0017	2.03 ± 0.08	2.03 ± 0.07	1.01 ± 0.05	ND
9–10	0.627	15,986	2.2	5394	0.0067 ± 0.0015	1.60 ± 0.06	1.61 ± 0.06	0.99 ± 0.05	ND
11-12	0.591	17,418	1.0	5407	0.0056 ± 0.0017	1.83 ± 0.07	1.71 ± 0.06	1.07 ± 0.06	ND
14–15	0.581	19,457	0.6	5352	0.0094 ± 0.0022	1.72 ± 0.07	1.84 ± 0.07	0.93 ± 0.05	ND
A01; 22° 44.22	2'N, 113° 3	9.257'E; 20.0 m; T	$T = 21.5 \circ C, S = 19.5; TSM$	$l = 279 mg l^{-1}$					
Bottom water		1755	113.0	117	0.00099 ± 0.00004	1.66 ± 0.07	3.42 ± 0.07	0.48 ± 0.02	
0-1	0.729	3070	12.0	296	0.0091 ± 0.0017	1.27 ± 0.07	3.34 ± 0.08	0.38 ± 0.02	1.60 ± 0.16
1–2	0.663	4253	1.0	474	0.031 ± 0.003	1.63 ± 0.06	2.01 ± 0.05	0.81 ± 0.04	0.52 ± 0.11
2-3	0.673	4795	2.3	557	0.033 ± 0.003	1.50 ± 0.06	1.76 ± 0.04	0.85 ± 0.04	0.15 ± 0.10
3-4	0.707	6886	1.1	825	0.027 ± 0.003	2.01 ± 0.07	2.07 ± 0.05	0.97 ± 0.04	0.01 ± 0.10
4–5	0.693	6573	5.4	1026	0.034 ± 0.003	3.20 ± 0.11	2.90 ± 0.07	1.10 ± 0.05	ND
5-6	0.689	9598	0.5	1145	0.034 ± 0.003	2.62 ± 0.12	3.30 ± 0.09	0.80 ± 0.04	ND
7-8	0.671	12,034	0.8	1482	0.034 ± 0.003	2.93 ± 0.12	2.69 ± 0.07	1.09 ± 0.05	ND
9–10	0.674	14.263	0.4	1778	0.032 ± 0.003	2.68 ± 0.11	2.96 ± 0.07	0.91 ± 0.04	ND
11-12	0.556	16,568	0.4	1932	0.018 ± 0.002	2.73 ± 0.11	2.60 ± 0.07	1.05 ± 0.05	ND
14–15	0.646	19,458	0.8	2054	0.016 ± 0.002	2.45 ± 0.10	2.40 ± 0.06	1.02 ± 0.05	ND
A03; 22° 36.41	'1'N, 113° 4	2.581'E; 7.0 m; T	$= 21.2 \circ C, S = 21.6; TSM$	$= 51.7 mg l^{-1}$					
Bottom water		1828	90.4	5.9	0.00064 ± 0.00003	1.63 ± 0.07	2.84 ± 0.07	0.58 ± 0.03	
0–1	0.748	ND	18.6	66	0.012 ± 0.002	1.39 ± 0.06	2.56 ± 0.07	0.54 ± 0.03	ND
1-2	0.716	2103	4.7	302	0.018 ± 0.003	1.85 ± 0.07	2.22 ± 0.05	0.83 ± 0.04	ND
2-3	0.701	2216	2.2	340	0.023 ± 0.003	1.47 ± 0.06	2.03 ± 0.05	0.72 ± 0.03	ND
3-4	0.707	3087	2.9	232	0.021 ± 0.002	1.83 ± 0.07	2.01 ± 0.05	0.91 ± 0.04	ND
4-5	0.722	3372	1.7	300	0.030 ± 0.003	2.64 ± 0.09	2.56 ± 0.05	1.03 ± 0.04	ND
5-6	0.763	3849	2.8	350	0.037 ± 0.003	1.61 ± 0.07	1.88 ± 0.05	0.86 ± 0.04	ND
7-8	0 743	4469	0.9	514	0.040 ± 0.003	2.19 ± 0.08	2.36 ± 0.06	0.93 ± 0.04	ND
9–10	0.669	5167	12	562	0.035 ± 0.003	1.48 ± 0.06	1.88 ± 0.05	0.79 ± 0.04	ND
11-12	0.710	5787	19	569	0.018 ± 0.002	1.10 ± 0.00 1.71 ± 0.07	1.80 ± 0.02 1.84 ± 0.05	0.93 ± 0.04	ND
14–15	0.657	7511	0.5	656	0.025 ± 0.003	1.90 ± 0.07	1.97 ± 0.05	0.97 ± 0.04	ND
404· 22° 31 19	05'N 113° 4	4 481' F · 8 0 m · T	$= 22.3 \circ C$ $S = 13.9 \cdot TSM$	$= 165 mg l^{-1}$					
Rottom water	5 11, 115 7	1698	138.5	79	0.00074 ± 0.00003	0.59 ± 0.06	0.79 ± 0.04	0.74 ± 0.08	
	0.518	1854	33.7	90	0.014 ± 0.0003	1.11 ± 0.05	1.45 ± 0.04	0.77 ± 0.00	ND
1_2	0.512	2366	23.7	120	0.019 ± 0.002	2.14 ± 0.08	236 ± 0.04	0.91 ± 0.04	ND
2_3	0.528	2704	27.1	157	0.029 ± 0.003	1.61 ± 0.06	1.89 ± 0.05	0.81 ± 0.04 0.85 ± 0.04	ND
	5.520				0.001 ± 0.000	+ 0.00		0.00 ± 0.01	

Appendix	T1	(continued)
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Depth cm	Porosity	$DIC \; (\mu mol \; l^{-1})$	$NO_3^- + NO_2^- \ (\mu mol \ l^{-1})$	$\mathrm{NH_4^+}~(\mathrm{\mu mol}~l^{-1})$	P. 224 Ra (dpm ml ⁻¹)	T. 224 Ra (dpm g ⁻¹)	228 Th (dpm g ⁻¹)	²²⁴ Ra/ ²²⁸ Th A.R.	234 Th (dpm g ⁻¹)
3–4	0.535	3347	10.8	216	0.040 ± 0.004	1.73 ± 0.07	1.98 ± 0.05	0.87 ± 0.04	ND
4–5	0.510	3249	3.8	176	0.051 ± 0.004	1.55 ± 0.06	1.72 ± 0.04	0.90 ± 0.04	ND
5–6	0.583	3344	1.8	186	0.055 ± 0.005	0.75 ± 0.03	0.97 ± 0.03	0.78 ± 0.04	ND
7–8	0.554	3427	0.3	204	0.052 ± 0.004	1.02 ± 0.04	1.13 ± 0.03	0.90 ± 0.05	ND
9–10	0.533	3732	0.5	222	0.051 ± 0.005	1.60 ± 0.07	1.70 ± 0.04	0.94 ± 0.05	ND
11-12	0.533	3226	0.7	232	0.037 ± 0.003	1.68 ± 0.07	1.86 ± 0.05	0.91 ± 0.04	ND
14–15	0.557	3698	0.3	260	0.041 ± 0.004	1.62 ± 0.07	1.80 ± 0.05	0.90 ± 0.04	ND
A06; 22° 23.48	6'N, 113° 4	6.497'E; 9.0 m; T	= 23.1 °C, S = 26.6; TSM	$= 17.5 mg l^{-1}$					
Bottom water		1886	55.0	4.0	0.00048 ± 0.00003	0.42 ± 0.004	1.12 ± 0.04	0.37 ± 0.04	
0-1	0.664	1707	33.0	33	0.013 ± 0.002	0.96 ± 0.04	1.07 ± 0.03	0.89 ± 0.04	ND
1–2	0.594	1208	15.2	69	0.029 ± 0.003	1.39 ± 0.05	1.40 ± 0.04	0.99 ± 0.05	ND
2–3	0.577	1554	2.1	96	0.042 ± 0.005	1.24 ± 0.05	1.36 ± 0.04	0.92 ± 0.04	ND
3–4	0.608	1468	1.2	90	0.037 ± 0.004	1.39 ± 0.05	1.48 ± 0.04	0.94 ± 0.04	ND
4–5	0.612	1231	2.0	129	0.047 ± 0.004	1.54 ± 0.06	1.54 ± 0.04	1.00 ± 0.04	ND
5–6	0.605	1567	0.5	140	0.056 ± 0.005	1.34 ± 0.06	1.35 ± 0.04	1.00 ± 0.05	ND
7–8	0.581	1956	0.6	165	0.043 ± 0.004	1.74 ± 0.07	1.87 ± 0.05	0.93 ± 0.04	ND
9–10	0.586	2318	0.7	172	0.042 ± 0.003	1.49 ± 0.06	1.57 ± 0.04	0.95 ± 0.05	ND
11-12	0.613	2780	0.8	255	0.032 ± 0.003	1.52 ± 0.06	1.61 ± 0.04	0.95 ± 0.05	ND
14–15	0.613	2814	0.6	242	0.033 ± 0.004	1.72 ± 0.07	1.74 ± 0.05	0.99 ± 0.05	ND
A09; 22° 12.04	6'N, 113° 4	8.795'E; 26.0 m; T	$T = 22.7 \circ C, S = 31.9; TSM$	$l = 14.9 mg l^{-1}$					
Bottom water		1484	22.2	2.4	0.00023 ± 0.00002	0.49 ± 0.05	1.02 ± 0.04	0.47 ± 0.05	
0-1	0.666	1774	5.6	99	0.019 ± 0.002	1.32 ± 0.05	1.48 ± 0.04	0.89 ± 0.04	1.27 ± 0.12
1–2	0.604	2075	2.5	96	0.014 ± 0.002	0.91 ± 0.04	1.16 ± 0.03	0.79 ± 0.04	0.89 ± 0.11
2–3	0.611	2319	2.1	113	0.019 ± 0.003	1.21 ± 0.04	1.27 ± 0.03	0.95 ± 0.04	0.04 ± 0.09
3–4	0.628	2174	2.4	62	0.018 ± 0.003	1.09 ± 0.04	1.24 ± 0.03	0.88 ± 0.04	0.07 ± 0.08
4–5	0.609	2097	1.7	145	0.021 ± 0.003	1.35 ± 0.05	1.39 ± 0.03	0.97 ± 0.04	ND
5–6	0.614	2127	1.1	141	0.019 ± 0.002	1.62 ± 0.07	1.44 ± 0.04	1.13 ± 0.06	ND
7–8	0.611	2376	0.7	188	0.024 ± 0.002	0.96 ± 0.04	1.05 ± 0.03	0.91 ± 0.05	ND
9–10	0.603	2428	0.4	215	0.027 ± 0.003	1.32 ± 0.05	1.31 ± 0.04	1.01 ± 0.05	ND
11-12	0.614	2910	0.5	239	0.021 ± 0.003	1.17 ± 0.05	1.41 ± 0.04	0.83 ± 0.04	ND
14–15	0.628	3125	1.2	221	0.021 ± 0.003	1.12 ± 0.06	1.23 ± 0.03	0.90 ± 0.05	ND

ND: Not detectable, i.e. when the second measurement of sediment ²³⁴Th is larger or indistinguishable from the first measurement.

* T, S, and TSM represent temperature, salinity and total suspended matter concentration in the bottom water.

Guo et al. (2008) suggested that over an annual cycle (wet season + dry season), the Pearl River delivers $\sim 478 \times 10^9$ mmol yr⁻¹ of DIC into the sea. In comparison, the flux of DIC from bottom sediments is $\sim 14-28\%$ of the riverine input of DIC. Our results based on the ²²⁴Ra/²²⁸Th disequilibrium approach in the dry season are in general agreement with the above estimate (18% vs. 14–28%).

5. CONCLUSIONS

The importance of pore water exchange in the overall budgets of water column DIC and nutrients in an estuary has long been recognized. Quantifying the benthic fluxes of these species, however, remains difficult due to limitations inherent in the traditional approaches, i.e., the benthic chamber method and the modeling approach. In this study, we have elaborated a newly developed approach - the ²²⁴Ra/²²⁸Th disequilibrium approach, to quantify the benthic fluxes of DIC and nutrients in a major estuary in China - the Pearl River Estuary (PRE). We demonstrate that irrigation was the predominant process that controls solute transfer across the sediment-water interface, whereas molecular diffusion and sediment mixing together accounted for <5% of the solute flux from bottom sediments. We show that sediment interstitial waters delivered approximately $42 \pm 6 \times 10^9$ mol of DIC and ~16 $\pm 1 \times 10^9$ mol of NH₄⁺ into the PRE in the dry season. In contrast, it removed about $13 \pm 1 \times 10^9$ mol of NO₃⁻ from the overlying water column. Overall, our study highlights irrigation as an important process that controls the concentration levels of water column DIC and nutrients in estuaries.

We believe that the $^{224}Ra/^{228}Th$ disequilibrium approach is a very powerful tool for quantifying the rate of solute transfer across the sediment-water interface. In this study, we found that dissolved ²²⁴Ra activities were generally well correlated with DIC and NH₄⁺ concentrations in the upper several centimeters of sediments. Under such a circumstance, the 224 Ra/ 228 Th disequilibrium approach would provide very reliable flux estimates of DIC and NH₄⁺. Future applications of the ²²⁴Ra/²²⁸Th disequilibrium approach may be extended to discern pore water exchange from submarine groundwater discharge (SGD) in estuarine and coastal environments. In terms of the methodology, inter-comparison between the ²²⁴Ra/²²⁸Th disequilibrium approach and the traditional approaches, such as a benthic chamber approach, should also be encouraged.

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APPENDIX

See Appendix T1.

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