A review of nitrogen isotopic alteration in marine sediments

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[1] Nitrogen isotopes are an important tool for evaluating past biogeochemical cycling from the paleoceanographic record. However, bulk sedimentary nitrogen isotope ratios, which can be determined routinely and at minimal cost, may be altered during burial and early sedimentary diagenesis, particularly outside of continental margin settings. The causes and detailed mechanisms of isotopic alteration are still under investigation. Case studies of the Mediterranean and South China Seas underscore the complexities of investigating isotopic alteration. In an effort to evaluate the evidence for alteration of the sedimentary N isotopic signal and try to quantify the net effect, we have compiled and compared data demonstrating alteration from the published literature. A >100 point comparison of sediment trap and surface sedimentary nitrogen isotope values demonstrates that, at sites located off of the continental margins, an increase in sediment ¹⁵N/¹⁴N occurs during early burial, likely at the seafloor. The extent of isotopic alteration appears to be a function of water depth. Depth-related differences in oxygen exposure time at the seafloor are likely the dominant control on the extent of N isotopic alteration. Moreover, the compiled data suggest that the degree of alteration is likely to be uniform through time at most sites so that bulk sedimentary isotope records likely provide a good means for evaluating relative changes in the global N cycle.

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1. Importance of the Marine N Cycle

[2] Nitrogen (N) is a major component of biomass and is required for photosynthesis. The total oceanic fixed N inventory and its availability in the surface ocean are thus important controls on primary production. Variations in these two parameters over geologic time have significant implications for the global biogeochemical cycling, not only of N, but also of carbon, and oxygen throughout Earth history. Sedimentary N isotope measurements have emerged as a powerful tool for monitoring the marine N cycle in the past and for testing important hypotheses explaining the ice core CO₂ records [Galbraith et al., 2008b, and references therein]. However, initial tests of the proxy demonstrated alteration of the primary isotopic signal during sinking and sedimentation and, more significantly, during burial in some settings [Altabet and Francois, 1994; Galbraith et al., 2008b]. Because local-to-regional scale spatial patterns of the surface and subsurface water column N isotopic composition of nitrate are largely mirrored in the sediments, despite evidence for alteration, paleoceanographic work proceeded [Altabet and Francois, 1994]. However, the focus of this work has been on the continental margins, where alteration is thought to be minimal [Altabet et al., 1999; Kienast et al., 2002; Thunell et al., 2004], or based on isotopic measurements of specific organic N pools (i.e., microfossil-bound and chlorophyll degradation products) [Higgins et al., 2009; Ren et al., 2009; Robinson et al., 2004]. Here, we evaluate evidence for and against alteration of bulk sedimentary N isotope ratios. A comparison of >100 published, co-located sediment trap and surface sediment N isotope values indicates that the degree of alteration increases with water depth in the ocean. The lack of evidence for an increase in ¹⁵N/¹⁴N with increasing sinking depth, tracked by sediment traps at the same geographic location and different depths, implies that alteration occurs largely at/in the seafloor. Despite the evidence for alteration, there exists the potential for a more comprehensive reconstruction of the paleo-marine N cycle that includes the widespread use of bulk measurements in combination with the measurement of specific organic N pools and global circulation models [Somes et al., 2010; E. Galbraith et al., Global nitrogen isotopic constraints on the acceleration of oceanic denitrification during the last deglacial warming, submitted to Nature Geoscience, 2012].

2. Processes Reflected by Sedimentary N Isotope Ratios

[3] Nitrogen in the ocean is present in many redox states, and biological processes are largely responsible for the various transformations of N from one form to another. These transformations are usually associated with fractionation of the N isotopes ($^{15}N/^{14}N$) [Altabet and Francois, 1994; Codispoti, 1989; Montoya, 1994; Wellman et al., 1968]. Kinetic isotope effects stem from differences in reaction rates between the heavy and light isotopes. The processes that are known to affect the isotopic composition of dissolved inorganic N (DIN) in the modern ocean include N₂ fixation by diazotrophic bacteria [Karl et al., 1997], nitrification, the extent of surface NO₃⁻ utilization [Altabet and Francois, 1994], denitrification [Liu and Kaplan, 1989],



Figure 1. Key steps in the marine N cycle typically recorded in the nitrogen isotopic composition of marine sediments. Estimates of isotopic enrichment factors for the individual transformations are noted. The green shaded area represents suboxic water columns where denitrification occurs (modified from *Sigman et al.* [2009]).

and the anammox reaction (S. Contreras et al., manuscript in preparation, 2012). The external processes, such as N_2 fixation and denitrification/anammox, control the inventory of nutrient N in the ocean while nitrification and uptake reflect internal cycling processes. In the modern ocean, the N isotopic compositions (as $\delta^{15}N$, where $\delta^{15}N =$ $15 \text{N}/14 \text{N}_{\text{sample}}/15 \text{N}/14 \text{N}_{\text{standard}} - 1*1000\%$, where the standard is atmospheric N₂) of different N pools are used to evaluate the relative roles of these various processes within the N cycle [Brandes and Devol, 2002; Deutsch et al., 2004; Lehmann et al., 2004; Sigman et al., 2009]. When organisms assimilate N to produce biomass, the ¹⁵N content of their N source is imprinted in the organic matter eventually deposited in sediments. The source is influenced by the preformed δ^{15} N of the water mass as well as any process that adds or removes N, such as remineralization or water column denitrification, along its flow path in the subsurface. The subeuphotic "source" signature may be overprinted by alteration of the isotopic signal in the surface ocean, through additions of newly fixed N or the partial consumption of NO_3^- . In sum, sedimentary δ^{15} N can reflect changes in ocean circulation, the biological pump, and large scale N cycling (Figure 1) [Brandes and Devol, 2002; Deutsch et al., 2004; Galbraith et al., 2008b; Robinson and Sigman, 2008; Sigman et al., 2010]. N isotopes also may be viewed as a paleoredox proxy because denitrification, the respiration of organic matter using NO_3^- , occurs under suboxic conditions [Galbraith et al., 2004; Jaccard and Galbraith, 2012; Kashiyama et al., 2008] (Figure 1).

[4] The δ^{15} N values of bulk sedimentary organic nitrogen reflect the δ^{15} N of the sinking flux of organic matter, plus any secondary isotopic alteration that occurs during sinking and burial due to either the removal or addition of N. To use the N isotopic composition of bulk sediments as a paleoceanographic tracer for surface waters, the extent of any alteration-related fractionation must be well constrained. Studies of sinking particles in the North Atlantic, Norwegian Sea, Sargasso Sea, Southern Ocean, and Southern California Bight have shown alteration of the δ^{15} N in sinking particles with depth during periods of low flux, or in low-productivity



Figure 2. Southern Ocean suspended and surface sediment δ^{15} N versus latitude [*Altabet and Francois*, 1994]. The large offset between suspended and surface sediment δ^{15} N values collapses in the region of higher accumulation rates under the Antarctic Polar Front (~44°S).

regions, perhaps due to the addition of nitrogen from other sources [*Altabet and Francois*, 2001; *Altabet et al.*, 1991; *Lehmann et al.*, 2002; *Lourey et al.*, 2003] or the loss of specific N_{org} fractions [*Macko and Estep*, 1984]. The observed changes are not unidirectional, but include both increases and decreases of δ^{15} N [*Lehmann et al.*, 2002]. However, during blooms, the δ^{15} N of sinking PN does not change with depth [*Altabet et al.*, 1991; *Altabet and Francois*, 1994]. Since high flux events contribute the majority of organic matter accumulating in sediments, organic N (N_{org}) that reaches the sediment-water interface generally should preserve the δ^{15} N signal of N_{org} in the surface.

3. Isotopic Alteration of the δ^{15} N Signal in Sediments

[5] Significant isotopic alteration of the organic N signal appears to occur at the sediment-water interface during early burial [Altabet and Francois, 1994]. Comparisons of sinking, suspended, and surface sedimentary δ^{15} N, led to the assertion that in low organic N flux regions such as the Southern Ocean, there is the potential for a large (3-6%)increase in the δ^{15} N values recorded in bulk sediments at the seafloor (Figure 2) [Altabet and Francois, 1994]. However, spatial variation in sedimentary δ^{15} N values fit expectations based on both surface nitrate concentration changes and δ^{15} N of nitrate values [Altabet and Francois, 1994; Farrell et al., 1995; Sigman et al., 1999]. In high sediment accumulation regions, such as on continental margins, the $\delta^{15}N$ values of surface sediments are essentially equal to the $\delta^{15}N$ of the sinking flux and/or subeuphotic zone nitrate, suggesting that no significant alteration occurs in these regions [Altabet et al., 1999; Kienast et al., 2002; Thunell et al., 2004] (Figure 3a).

[6] Several attempts to understand isotopic alteration of sedimentary N in the natural environment were made [*Altabet et al.*, 1999; *Freudenthal et al.*, 2001a; *Prokopenko et al.*, 2006a; *Velinsky et al.*, 1991]. Sequential extractions from margin sediments led to the operational definition

of four sedimentary nitrogen pools: (1) organic nitrogen, (2) acid released/mineral bound, (3) weakly bound ammonium (NH_4^+), and (4) tightly bound NH_4^+ [Freudenthal et al., 2001a]. For example, the $\delta^{15}N$ of the organic N pool sampled off the Moroccan coast increased within the top 10 cm and then stabilized, suggesting that even in margin sediments, some alteration may occur [Freudenthal et al., 2001a]. In a second study, measurements of sedimentary $\delta^{15}N$ and pore water-NH₄⁺ δ^{15} N from the same horizons suggest that there is little or no offset between the two N pools in organic-rich, marine dominated margin sediments (<1%), despite oxidation of a significant proportion of the organic nitrogen delivered to the seafloor and the large buildup of pore water NH₄⁺ [*Prokopenko et al.*, 2006a, 2006b]. This is consistent with the observation that in rapidly accumulating sediments there is no offset between sinking flux $\delta^{15}N$ and bulk $\delta^{15}N$ of surface sediment (Figure 3a) [Altabet et al., 1999; Kienast et al., 2002; Thunell et al., 2004]. One explanation for the lack of offset, despite clear evidence for loss of organic N, is that at high organic matter concentrations there is no preferential removal of specific organic fractions. That is, the fraction that is lost has a N isotopic composition that is equivalent to that of the residual organic matter. Alternatively, and probably less likely, the diversity of the sedimentary microbial consortium may lead to the cancellation of the various fractionating processes.

[7] In lower flux environments, where organic matter content is low and the δ^{15} N of the sinking flux is lower than that of the surface sediments, the processes must be different. The increase in bulk $\delta^{15}N$ appears to occur at/in the seafloor and is generally attributed to the preferential loss of components of the bulk organic matter with lower $\delta^{15}N$. While there is some field and experimental evidence for a decrease in $\delta^{15}N$ of the bulk organic pool during early sedimentary diagenesis, at least under anaerobic preservation conditions [Altabet et al., 1991, Libes and Deuser, 1988, Lehmann et al., 2002], the majority of existing studies to date suggests that the $\delta^{15}N$ of sedimentary organic matter increases during alteration. Evidence for the increase in δ^{15} N of organic nitrogen comes from both downcore profiles of δ^{15} N in the uppermost, oxic zone of the sediment column [Freudenthal et al., 2001a] as well as incubation experiments [Holmes et al., 1998]. Experimental investigation of the N isotope effect associated with deamination suggests that generally low δ^{15} N NH₄⁺ is produced [Macko and Estep, 1984]. Leakage of low δ^{15} N NH₄⁺ into pore waters could explain the increase in the bulk sedimentary δ^{15} N, provided this NH₄⁺ is not reincorporated into bacterial biomass or readily adsorbed onto the sediment matrix. This is plausible in oxic sedimentary environments where ammonium pools are small or negligible due to the rapid and essentially complete oxidization to NO_3^- (e.g., near sediment/water interface or beneath the oligotrophic gyres) and/or where clay/organic matter content is minimal (i.e., biogenic sediments dominate).

[8] Bulk δ^{15} N values can also be influenced by interference from terrestrial materials, NH⁴ absorption into clay minerals, and winnowing/size fractionation [*Mollenhauer et al.*, 2005; *Kienast et al.*, 2005; *Schubert and Calvert*, 2001]. The impact of these processes is more significant in organic-poor sediments. NH⁴₄ is associated with the solid





phase of the sediment in fixed and exchangeable forms. Exchangeable NH₄⁺ is adsorbed by ion exchange reactions on the surfaces of organics and minerals and makes up only a small proportion of the bulk sedimentary N (typically <1%) in pelagic sediments [de Lange, 1992; Freudenthal et al., 2001a; *Müller and Suess*, 1979]. It can account for a significantly larger fraction in anoxic, organic-rich sediments, as the amount of exchangeable NH_4^+ increases with the $[NH_4^+]$ of the pore waters [*Rosenfeld*, 1979]. Fixed NH⁺₄, on the other hand, is incorporated into clay mineral structures and can comprise a significant to dominant proportion of the bulk N [Carman et al., 1996; Freudenthal et al., 2001a; Kienast et al., 2005; Schubert and Calvert, 2001]. In organic-poor sediments, inorganic N can make up 94% of the total N [Schubert and Calvert, 2001]. The fixed fraction is made up of an inherited component, that is NH_4^+ delivered with the lithogenic sediments, as well as what is adsorbed in situ. As with the exchangeable NH_4^+ , the fixed fraction tends to increase with increasing pore water $[NH_4^+]$, but to a lesser degree [Rosenfeld, 1979]. The adsorptive potential of any given sediment depends upon its clay content and mineralogy, NH₄⁺ availability, and potassium content of the sediments and interstitial waters [Carman et al., 1996; Mackin and Aller, 1984; Rosenfeld, 1979]. Because fixed NH₄⁺ substitutes for a cation within the lattice structure of clay minerals, it occurs at a slower rate than the ion-exchange involved in adsorption of exchangeable NH₄⁺. Although this slow rate makes substitution difficult to evaluate quantitatively in the lab, it is likely relevant on geologic timescales. The consequences for bulk δ^{15} N depend upon the size of the NH₄⁺ pool relative to the ON pool, the $\delta^{15}N$ of the NH_4^+ pool (s) (both what was inherited with clay minerals and produced in situ), the proportion of the NH_4^+ that is fixed relative to what may be oxidized to NO_3^- , and any fractionation associated with the adsorption process [Karamanos and Rennie, 1978]. In sum, alteration of the bulk N isotopic value from that of the sinking flux may occur in organic poor sediments where NH_4^+ may make up a significant portion of total sedimentary N because it may have several fates (oxidation, adsorption, utilization) with fractionation effects associated with each process.

4. Case Studies

[9] In this section we present two case studies highlighting the challenges and complexities of investigations into the cause of "diagenetic" alterations of the δ^{15} N signal, and review some of the approaches taken/progress made over the last 20 years. The first, from the South China Sea (SCS), illustrates how alteration, due to diagenesis and variable

contributions of inorganic N from land, can result in misleading bulk δ^{15} N results [Kienast et al., 2005]. Based on a minor offset between δ^{15} N values in the fluff layer and the uppermost sediment sample of only 0.3-1.2‰, and the absence of a correlation between increasing $\delta^{15}N$ and decreasing percent TN in the downcore records, Kienast [2000] argued modification of the δ^{15} N signal in the water column or during sediment burial in the SCS to be insignificant. This seemed to be in line with expectations for minimal alteration in marginal settings with high Corg burial rates, compared to oligotrophic sites with low sedimentation rates [Altabet, 1996]. The downcore record that accompanied this assertion in Kienast [2000] is widely cited, as it remains one of the few sedimentary $\delta^{15}N$ records from an oligotrophic site. Three lines of evidence, however, suggest that the conclusion of no alteration, and thus the significance of this record, need to be reevaluated. First, since the study in 2000, trap time series from the SCS have become available [Gave et al., 2009], which suggest that " δ^{15} N in sediments underlying trap locations were 1.5–3‰ higher than the annual average of sinking particles," comparable to other sites studied (Figure 3a). Second, analysis of fluff samples from many more of the stations within the SCS (n = 32)reveals that the average offset between fluff samples and the topmost sediment sample of 0.6% at four of the core sites presented in Kienast [2000] is a minimum estimate (Figure 4). Indeed, there is a significant correlation between increased sedimentary δ^{15} N and a higher offset between fluff and sediment surface (Figure 4). To a first approximation, this trend implies that surface sediment values above 5-5.5%in the SCS are caused by an isotopic enrichment during early sedimentary diagenesis of >1% and up to 3% during the transition from fluff to surface sediment. Third, *Kienast et al.* [2005] showed that 35–65% of TN in the SCS is (operationally defined) inorganic N, with an isotopic composition of 3.1–4.8^w. These authors argued that "the significant percentage of N_{inorg} and the distinct isotopic signature will... dampen any variability associated with N_{org} in the bulk $\delta^{15}N$ signal" [Kienast et al., 2005, p. 5] Closer inspection of two of the records from the southern SCS (sites 17961 and 17964) reveals how a significant post-depositional alteration of the organic nitrogen could have been effectively masked in the sedimentary record: A substantial loss of organic nitrogen (ca. 20–25%) in the top 1-3 m, equivalent to the last 10 ka, of these two cores is paralleled by an increase in δ^{15} N of organic N by up to 2‰, which results in a near constant $\delta^{15}N$ of total N, similar to Freudenthal et al.'s [2001a] results. Finally, physical processing of sediment may also have contributed to spatial differences in downcore records in the South China Sea. Size-related differences in

Figure 3. Site locations of the global sediment trap compilation with color coding to highlight the geographic distribution of $\Delta \delta^{15}$ N values (sediment-sinking δ^{15} N) (map panel). Surface versus sinking sedimentary δ^{15} N values with color coding to denote water depth differences for each location (a) The *Gaye et al.* [2009] South China Sea sediment trap data are outlined in black. Surface sedimentary versus nitrate δ^{15} N values (black diamonds) and surface versus sinking values from several margin locations fall along the 1-to-1 line (solid black line). Reduced major axis linear regression for all sites (dotted line) as well as a subset of sites made up of those with depths >1000 m (dashed line), suggest a line with a slope of 1.2 and an intercept of ~1‰ (1.1–1.4‰). (c) $\Delta \delta^{15}$ N (sediment-sinking δ^{15} N) shows a strong relationship to water depth (r² = 0.46, Japan Trench samples highlighted in gray oval marked JT are excluded from this correlation) but (b) not to sinking depth (r² = 0.11), suggesting (d) that the alteration takes place at the seafloor. See Table 1 for full details on data, including location, trap depths, and references.



Figure 4. Crossplot of $\Delta \delta^{15}$ N (surface δ^{15} N-fluff δ^{15} N) and surface sediment δ^{15} N values from the South China Sea showing the variability in preservation across the basin. The first order increase in sedimentary δ^{15} N values appears to be due to isotopic alteration (M. Kienast, unpublished data, 2006).

the δ^{15} N of particles exist and sediments that witness either winnowing or focusing will record an altered bulk δ^{15} N record. This is most likely to occur where there are heterogeneous mixtures of organic matter, such as on the continental margins and in marginal seas [*Kienast et al.*, 2005]. The end result is that the sediments in the South China Sea experience variable degrees of alteration and the *Kienast* [2000] records are not likely faithfully recording water column variations.

[10] The next case study involves the well-studied sediments of the Mediterranean Sea. The Mediterranean sediments present striking evidence for variability in their preservation of sedimentary organic matter (e.g., alternating marl and sapropel layers). Accompanying this variability are large systematic changes in the bulk N isotopic composition of the sediments with higher δ^{15} N values in the low TOC marls and lower δ^{15} N values in the organic rich sapropels. Some interpretations of the isotope records have inferred significant overprinting of the primary δ^{15} N values by alteration [*Möbius et al.*, 2010; *Sachs and Repeta*, 1999], while others have interpreted the records to reflect primary signatures [*Calvert et al.*, 1992; *Higgins et al.*, 2010].

[11] The evidence for isotopic alteration comes from observed increases in bulk δ^{15} N associated with post depositional oxidation of sapropelic organic carbon [Moodley et al., 2005] and from the comparison of bulk δ^{15} N values to the amino acid (AA) Degradation Index (DI) [Dauwe et al., 1999; Möbius et al., 2010]. DI is the result of statistical evaluation of systematic compositional changes of the 14 most common protein AAs during remineralization [Dauwe et al., 1999]. DI scores sediments with a value of 1 for fresh organic matter toward low values (≤ -0.5) with intensive organic matter degradation. Sediments from the Mediterranean, both surface sediments from a spatially distributed survey and downcore profiles, show a linear relationship between DI and bulk δ^{15} N values [*Möbius et al.*, 2010], suggesting that changes in δ^{15} N may be linked to organic matter quality. *Möbius et al.* [2010] suggested that the low δ^{15} N values that occur in the sapropel intervals are the result of enhanced preservation, and that diagenetic isotopic alteration raised the δ^{15} N values of the marl sedimentary organic matter. However, the correlation between DI and δ^{15} N does not, on its own, indicate causality.

[12] This interpretation is at odds with results comparing the δ^{15} N of chlorins, chlorophyll *a* degradation products, and bulk δ^{15} N from Mediterranean sedimentary sequences containing both sapropel and marl horizons. Throughout the profiles, δ^{15} N values from chlorins are consistently offset from bulk values by 4–5‰, implying that the bulk $\delta^{15}N$ values recorded in the organic poor marl sequences are not heavily overprinted by alteration, and that the low $\delta^{15}N$ values during sapropel formation represent significantly different N biogeochemical conditions in the overlying surface waters [Higgins et al., 2010]. These data call into question the basic assumption that δ^{15} N data from organic-poor sediments should be discounted categorically and suggest that, at least in this case, the degradation index reflects good preservation at times when the water column above produced low δ^{15} N organic matter. This would be consistent with abundant N₂ fixation and rapid accumulation of organic matter and/or O₂-poor bottom waters during sapropel formation, for example.

5. Sediment Trap and Surface Sediment Compilation

[13] In an effort to evaluate the degree of isotopic alteration from a global data set and to put some of the individual sediment trap studies discussed into a larger context, we compiled published, co-located, sediment trap-sinking particle and surface sediment $\delta^{15}{\rm N}$ values (Figure 3 and Table 1). The term co-located describes trap and surface sediment sampling stations that are nominally within the same location. In most cases, both data types were presented together, but in the case that we assigned surface sediment values from separate studies to available trap data, we defined co-location as being within 1° of latitude/longitude of one another and most are within 0.5°. The sites, not surprisingly, are largely located on the margins and in the Northern Hemisphere (see map in Figure 3). The compilation includes sites at a range of water depths (180–9200 m), primary production (based on Chl a $0.09-5 \text{ mg/m}^3$) and sedimentation rates (2->200 cm/kyr), as well as distances from land. Overall, the compilation reveals a fairly consistent relationship between the sinking flux and the sedimentary δ^{15} N values, despite the fairly large range of $\Delta \delta^{15}$ N values of -2 to 6.5% ($\Delta\delta^{15}N$ = surface sediment - sinking δ^{15} N). The mean is 2.3 \pm 1.8‰ (1 σ) (Figure 3a). Only a handful of surface sediment samples fall along the 1:1 line in the crossplot of sediment versus sinking $\delta^{15}N$ (Figure 3a) and they are from highly productive continental margin locations. Surface sediment δ^{15} N versus subeuphotic zone $\delta^{15}N_{\text{nitrate}}$ from several margin locations also fall along the

Source ^a	M. Altabet, unpublished data, 1991; Fry et al. [1991] (sediments)	M. Altabet, unpublished data, 1991; Fry et al. [1991] (sediments)	Möbius et al., 2010; J. Möbius (personal communication, 2012) (SR)	Möbius et al., 2010; J. Möbius (personal communication, 2012) (SR)	Möbius et al., 2010; J. Möbius (personal communication, 2012) (SR)	Möbius et al., 2010; J. Möbius (personal communication, 2012) (SR)	Freudenthal et al. [2001b]	Thunell et al. [2004]	Thunell et al. [2004]	Velinsky and Fogel [1999]; Næs et al. [1988]	Velinsky and Fogel [1999]; Næs et al. [1988]	Velinsky and Fogel [1999]; Næs et al. [1988]	Velinsky and Fogel [1999]; Næs et al. [1988]	Velinsky and Fogel [1999]; Næs et al. [1988]	Velinsky and Fogel [1999]; Næs et al. [1988]	Struck et al. [2004]; Kotilainen [2006] (SR)	Lavik [2001]; Sarnthein [2003a, 2003b] (SR)	Lavik [2001]; Bickert and Wefer [1996] (SR)	Lavik [2001]; Bickert and Wefer [1996] (SR)	Lavik [2001]; Bickert and Wefer [1996] (SR)	Lavik [2001]; Jahns et al. [1998] (SR)	Lavik [2001]; Meinecke [1992] (SR)	Lavik [2001]; Meinecke [1992] (SR)	Lavik [2001]; Sarnthein [2003a, 2003b] (SR)	Holmes et al. [2003]; Frank and Mackensen [2002] (SR)	Holmes et al. [2003]; Frank and Mackensen [2002] (SR)	Holmes et al. [2002]; Bickert and Mackensen [2003] (SR)	Holmes et al. [2002]; Bickert and Mackensen [2003] (SR)	Gaye-Haake et al. [2005]	Gaye-Haake et al. [2005]; Sirocko et al. [2000]	Gaye-Haake et al. [2005]; Sirocko et al. [2000]	Gaye-Haake et al. [2005]	Gaye-Haake et al. [2005]	Gaye-Haake et al. [2005]; Sirocko et al. [2000]	Gaye-Haake et al. [2005]; Sirocko et al. [2000]	Gaye-Haake et al. [2005]	Gaye-Haake et al. [2005]	Gaye-Haake et al. [2005]	Garrier Diarke et al. [2000] $Garrier af al [7006]$	Uaye-Haake et al. [2002], Dirocko et al. [2000]							
BW [02] (mg/L)	0.00	0.00	4.15	4.00	4.00	4.00	3.37	5.48	5.48	5.47	5.47	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		5.71	5.74	5.74	5.72	5.80	5.53	5.53	5.62	4.97	4.97	5.10	5.10	2.46	3.46	3.92 2.02	3.96	3.20	3.20	3.36	3.36	3.46	3.46	0.28	3.14	3.14	2.77	00.0
Sed Rates (cm/kyr)	300.00	300.00	6.00	6.00	6.00	6.00	9.1	29.13	29.13	1.5	1.5	30.00	30.00	21.58	21.58	21.58	21.58	21.58	21.58	108.89	3.71	3.01	3.01	3.01	2.20	5.17	3.54	5.49	10.83	10.83	3.1	3.1					7.75	7.75			4.13	4.13				25 C	00.7
Chl a (mg/L)	0.80	0.68	0.11	0.11	0.11	0.11	0.29	0.13	0.13	0.11	0.11	0.63	0.63								0.34	0.19	0.19	0.19	0.17	0.19	0.26	0.30	0.22	0.22	0.62	0.62	0.17	0.17	0.24	0.28	0.26	0.26	0.34	0.34	0.38	0.38		0.17	0.17	0.19 0.70	N.47
$\Delta \delta^{15} N$ (% v. air)	-0.1	0.5	1.3	2.3	2.6	2.7	3.4	2.5	4.7	1.6	4.9	-0.8	-0.3	-2.0	0.3	-0.5	-0.2	0.0	-0.1	-0.3	3.0	3.3	3.0	3.4	2.7	3.5	4.0	3.0	2.4	-2.0	1.6	0.2	2.7	2.1	4.4	4.1	4.0	3.6	4.1	4.1	3.0	2.7	0.5	-0.3	-0.3	1.1	0.0
Surface δ ¹⁵ N (‰ v. air)	2	2	3.5	3.5	3.5	3.5	6.5	6.7	6.7	7.1	7.1	3.4	3.4	2.14	2.14	2.14	2.14	2.14	2.14	3.4	8.2	7.6	7.6	7.6	6.1	9.6	9.4	8.7	5.2	5.2	7.1	7.1	7.0	0.7	9.6	9.6	9.8	9.8	10.5	10.5	10.0	10.0	8.2	6. 6.	ω υ.	0.7	11.4
Flux Weighted $\delta^{15}N$ (‰ v. air)	2.1	1.5	2.2	1.2	0.9	0.8	3.1	4.2	2.0	5.5	2.2	4.2	3.7	4.18	1.89	2.6	2.33	2.16	2.23	3.7	5.2	4.3	4.6	4.2	3.4	6.1	5.4	5.7	2.8	7.2	5.5	6.9	4.9 6.9	4.5 5.5	1.0	0.0	5.8	6.2	6.4	6.4	7.1	7.3	7.7	3.6	3.6	2.0 20 20	0.1
Trap Depth (m)	1300	477	3720	2560	1508	2689	700	500	3000	006	3800	225	1200	15	40	80	120	160	175	140	1097	953	859	853	984	598	948	1068	614	3196	599	1648	2250	006	3000	006	2800	1400	3090	1090	2080	1080	590	1900	1000	730	00/
Bottom Depth (m)	2055	2220	3750	3600	3620	3620	966	3610	3610	4327	4327	1400	1400	180	180	180	180	180	180	240	4141	4399	4522	4481	4524	3450	3490	3906	3750	3750	2196	2196	3271	3271	4020	4020	3770	3770	4075	4075	4020	4020	1100	2700	2700	2000	0060
Longitude	37.57	34.01	26.1792	26.19	26.193	26.193	346.85	344.55	344.55	342.05	342.05	295.33	295.33	6.75	6.75	6.75	6.75	6.75	6.75	20.24	349.23	348.75	348.86	348.87	348.84	350.58	349.74	349.91	5.83	5.83	9.15	9.15	84.33	84.33	80.92	86.92	68.67	68.67	67.13	67.13	60.50	60.50	65.82	88.83	88.83	89.11	04.//
Latitude	42.35	43.00	34.43	34.44	34.44	34.44	28.72	29.13	29.13	29.77	29.77	10.08	10.08	58.13	58.13	58.13	58.13	58.13	58.13	57.30	-0.08	1.78	1.79	1.79	3.17	-5.79	-4.19	-2.19	-50.13	-50.13	-20.05	-20.05	13.1	13.1	0.c	6.0	15.5	15.5	13.2	13.2	16.3	16.3	24.8	15.5	16.5	د.ئ ج 11	L4.U
Region	Black Sea	Black Sea	Mediterranean Sea	Mediterranean Sea	Mediterranean Sea	Mediterranean Sea	Canary Island region	Cariaco Basin	Cariaco Basin	Framvaren Fjord, Norway	Gotland Sea (Baltic)	Eq. Guinea Basin	N Guinea Basin	N Guinea Basin	N Guinea Basin	N Guinea Basin	S Guinea Basin	S Guinea Basin	S Guinea Basin	South Atlantic	South Atlantic	Walvis Ridge	Walvis Ridge	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ocean	Indian Ucean													

Table 1. Tabulated Data From Sediment Trap and Surface Sediment $\delta^{15}N$ Comparison With References

Source ^a	Gave-Haake et al. [2005]; Sirocko et al. [2000]	Gave et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	Gaye et al. [2009]; Wang et al. [1995] (SR)	<i>Altabet</i> [2001]; <i>Murray et al.</i> [2000] (SR)	Altabet [2001]; Murray et al. [2000] (SR)	Altabet [2001]; Mutray et al. [2000] (SK) Altabet et al [1990]: Thunell et al [1994]	Altabet et al. [1999]: Thunell et al. [1994]	Altabet et al. [1999]	Altabet et al. [1999]; Lewis et al. [2002]	Silverberg et al. [2004]; R. Thunell (personal communication, 2011)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Kienast et al. [2002] (SR)	S. Kienast (unpublished data, 2002); Lyle et al. [1992] (SR)	S. Kienast (unpublished data, 2002); Lyle et al. [1992] (SR)	S. Kienast (unpublished data, 2002); Lyle et al. [1992] (SR)	Kienast et al. [2002]	Kienast et al. [2002] Kienast et al. [2002]										
BW [O2] (mg/L)	3.30	2.15	2.15	2.40	2.40	2.60	2.60	2.60	2.58	2.58	2.50	2.40	2.40	2.58	2.58	3.93	3.93	3.84	3.87	3.81	3.81	3.81	3.77	3.78	3.78	3.78	3.99	96.5 070	0.00		0.57	0.32	2.51	2.51	2.51	2.51	2.51	2.51	2.51	2.51	2.75	2.75	2.75	2.10	2.10
Sed Rates (cm/kyr)	2.38	2.15	2.15	2.40	2.40	2.60	2.60	2.60	2.58	2.58	2.40	2.40	2.40	2.58	2.58	3.93	3.93	3.84	3.87	3.81	3.81	3.81	3.77	3.77	3.77	3.77	3.99	5.99 260.00	118.00	100.00	200.00	108.00	9.40	9.40	9.40	9.40	9.40	9.40	9.40	9.40	1.30	1.30	1.30	27.00	27.00 27.00
Chl a (mg/L)	0.29	0.12	0.12	0.11	0.11	0.10	0.10	0.10	0.11	0.11	0.11	0.11	0.11	0.13	0.13	0.11	0.11	0.13	0.33	0.36	0.36	0.36	0.23	0.15	0.15	0.15	0.11	0.80	1.03	1.10		0.35	0.31	0.31	0.31	0.31	0.31	0.31	0.31	0.31	0.20	0.20	0.20	0.39	0.38 0.39
Δδ ¹⁵ N (‱ v. air)	4.2	1.2	0.5	1.4	2.5	2.0	2.2	1.8	2.3	2.9	0.6	1.5	1.4	1.6	2.2	1.9	2.9	3.8	4.2	4.8	4.6	4.6	5.8	2.9	2.8	3.2	6.5	0.0	1.7	0.5	-0.7	0.4	-0.7	2.6	1.3	1.8	-0.2	0.0	-0.3	0.3	2.8	1.6	3.7	-1.5	-0.8 0.5
Surface δ ¹⁵ N (‰ v. air)	11.4	4.5	4.5	5.9	5.9	4.9	4.9	4.9	5.6	5.6	4.2	4.2	4.2	5.4	5.4	15.5	15.5	7.7	7	9.4	9.4	9.4	12.3	11.3	11.3	11.3	13.9	13.9 104	11.1	8.5	7.1	10.6	8.03	8.03	8.03	8.03	8.03	8.03	8.03	8.03	11.1	11.1	11.1	6.46	6.40 6.46
Flux Weighted $\delta^{15}N$ (% v. air)	7.2	3.3	4.0	4.5	3.4	2.9	2.7	3.1	3.3	2.7	3.6	2.7	2.8	3.8	3.2	13.64	12.63	3.92	2.80	4.55	4.85	4.78	6.50	8.42	8.46 2.25	8.07	7.35	1 <i>6.1</i>	9.4	×	7.8	10.2	8.75	5.43	6.78	6.25	8.24	8.01	8.31	7.72	8.26	9.47	7.4	7.93	7.25 5.93
Trap Depth (m)	3020	009	1200	900	1700	1200	2000	3750	1465	3500	1225	1765	2287	1000	3350	1292	3594	2099	3593	880	3618	2284	2203	1200	2200	3800	2150	4400 665	485	500	450	330	500	1500	1750	2330	1000	1500	1750	2350	3218	1500	1750	1500	1750 1
Bottom Depth (m)	3900	1730	1730	2300	2300	4300	4300	4300	4038	4038	2826	2628	2628	3766	3677	4294	4294	4198	4293	4385	4385	4385	4397	4493	4493	4493	5100	001 C	665 665	850	950	450	3133	3133	3133	3133	3133	3133	3133	3133	3700	3700	3700	2700	2700 2700
Longitude	64.77	109.72	109.72	111.28	111.28	115.12	115.12	115.12	117.18	117.18	119.52	119.52	119.52	116.02	116.02	224.97	224.97	220.27	220.27	220.18	220.18	220.18	219.87	220.22	220.22	220.22	220.02	20.022 249 08	248.33	241.50	237.95	292.7	-127.578	-127.578	-127.578	-127.578	-127.578	-127.578	-127.578	-127.578	-131.99	-131.99	-131.99	-125.761	-125.762 -125.832
Latitude	14.5	9.15	9.15	11.40	11.40	14.62	14.62	14.62	17.07	17.07	17.27	17.27	17.27	18.47	18.47	-11.97	-11.97	-4.95	-1.95	0.07	0.07	0.07	2.00	5.02	5.02	5.02 2.02	00.6	9.00 26.03	27.88	33.55	36.75	25.2	42.192	42.192	42.192	42.192	42.192	42.192	42.192	42.192	41.564	41.564	41.564	42.084	42.049 42.108
Region	Indian Ocean	South China Sea	Equatorial Pacific	Equatorial Pacific	Equatorial Pacific	Equatorial Pacific	Equatorial Pacific	Equatorial Pacific California Current: Carmen B	California Current: Guavmas B	California Current; San Pedro B	California Current; Monterey Bay	California Current; Soledad B	California Current	California Current	California Current	California Current	California Current	California Current California Current																											

Table 1. (continued)

					Flux						
			Bottom	Trap	Weighted	15	- 1 <i>5</i>				
			Depth	Depth	$N_{c1}g$	Surface $\delta^{12}N$	$\Lambda^{c_1} \delta \Sigma$	Chl a	Sed Rates	BW [02]	
Region	Latitude	Longitude	(m)	(m) (‰ v. air)	(‰ v. air)	(%o v. air)	(mg/L)	(cm/kyr)	(mg/L)	Source ^a
apan Sea	42.73	138.28	3665	1090	4.2	6.8	2.6	0.49	4.52	5.19	Nakanishi and Minagawa [2003]; Crusius et al. [1999]
apan Sea	42.73	138.28	3665	1600	5.1	6.8	1.7	0.49	4.52	5.19	Nakanishi and Minagawa [2003]; Crusius et al. [1999]
apan Sea	42.73	138.28	3665	3110	4.0	6.8	2.8	0.49	4.52	5.19	Nakanishi and Minagawa [2003]; Crusius et al. [1999]
apan Trench	34.17	142.0	9200	4789	2.9	5.8	2.9	0.22	23.00		Nakatsuka et al. [1997]; Boggs [1984]
apan Trench	34.17	142.0	9200	8789	3.5	5.8	2.3	0.22	23.00		Nakatsuka et al. [1997]; Boggs [1984]
apan Trench	34.17	142.0	9200	1674	3.3	5.8	2.5	0.22	23.00		Nakatsuka et al. [1997]; Boggs [1984]
apan Trench	34.17	142.0	9200	4180	2.0	5.8	3.8	0.22	23.00		Nakatsuka et al. [1997]; Boggs [1984]
apan Trench	34.17	142.0	9200	5687	3.0	5.8	2.8	0.22	23.00		Nakatsuka et al. [1997]; Boggs [1984]
apan Trench	34.17	142.0	9200	8688	3.3	5.8	2.5	0.22	23.00		Nakatsuka et al. [1997]; Boggs [1984]
^a "SR" following a citation in	dicates that c	itation is the	source o	f the dat	ta in the Se	dimentation R:	ate column.				

Table 1. (continued)

1:1 line (Figure 3a). The remaining data tend to lie above the 1:1 line and these data are broadly distributed geographically, without an apparent pattern of variation. A reduced major axis linear regression (which accounts for error in both the x and y data sets) yields a slope close to 1(1.2) and a y-intercept of 1.1%. If sites with depths <1000 m are excluded (this includes all of the sites on the 1:1 line), the slope is the same, 1.2, and the intercept is 1.4 ‰. In both cases, the relationship is significant, with a p value <0.001 at 95% confidence limit. This suggests that the offset between bulk and sinking $\delta^{15}N_{PN}$ values is relatively consistent across a range of depositional environments but that there is an increase in the offset with increasing water depth (Figures 3a). The $\Delta \delta^{15}$ N values increase with water depth by 0.5%/km using the entire data set and by $\sim 1\%$ /km if the samples from the Japan trench, which fall off of the trend, are excluded (Figure 3c). This is similar to the increase in δ^{15} N values observed with water depth for a larger, surface sediment database (0.4%/km) (Galbraith et al., submitted manuscript, 2012). No such linear trend is observed if we compare $\Delta \delta^{15}$ N to sinking depth (water depth - trap depth) (Figure 3b). Nor is any systematic change observed when one examines data from locations with trap data from multiple depths; in some cases it is unchanged, in others it decreases and in still others it increases. This suggests that while there is isotopic alteration in the water column, the systematic increase that is observed is likely a product of early burial diagenesis at the seafloor [Altabet and Francois, 1994], rather than in the water column between the trap and the seafloor. This is consistent with the observations from the South China Sea fluff-sediment comparison (see above). The variability observed when comparing $\delta^{15}N$ values between traps also suggests that the controls in the water column are more complex.

[14] The discrepancy between the $\delta^{15}N$ values in trap material and sediments could have several causes. It may simply be due to differences in the sources of organic matter to traps and to the seafloor. This is caused by differences in the timescale of deposition between traps and sediments. where short-term perturbations, either natural such as blooms or dust storms or anthropogenic including plumes and agricultural runoff, have the potential to heavily influence what is captured in traps but may not appear in the seafloor signal. It can also be due to "swimmers" in the traps or lateral transport at the seafloor. Transport at the seafloor has the potential to fractionate the sediments by size, which in turn may alter the isotopic signal. Transport-related processes in particular may be important in this data set, given its bias toward the margins, where organic matter may be moved downslope beneath surface waters with a strong gradient in $\delta^{15}N_{\text{nitrate}}$ values [e.g., Freudenthal et al., 2001a] and toward open ocean sites located in regions with active sediment focusing (e.g., Equatorial Pacific, Southern Ocean) [Francois et al., 2004; Mollenhauer et al., 2005]. However, we compiled as large a data set as possible in order to highlight first order trends in the data and to avoid bias by site specific effects, and despite the broad scatter in the data, the observed trends are significant. We attribute the majority of the isotopic change between traps and surface sediments to early diagenesis of the isotopic signal.

[15] Preservation of organic matter, while not perfectly understood, is likely a function of the length of time that



Figure 5. Crossplots of (left) $\Delta \delta^{15}$ N versus bottom water [O₂], (middle) Chl-a concentration, and (right) sedimentation rate for the sites in the sediment trap database. Sites close to the margins were excluded from the Chl-a comparison because of the large error associated with nearshore-site Chl-a estimates from satellites. In all cases but Chl-a, the correlations are statistically significant at a 95% confidence interval.

organic matter is exposed to molecular O₂ in pore waters [Hartnett et al., 1998], which is a function of bottom water oxygen concentrations and total sediment and organic carbon accumulation rates. For comparison, we have plotted our $\Delta \delta^{15}$ N values against first order estimates of these parameters (Figure 5). In each case, values are estimated from gridded data sets; bottom water oxygen estimates are from WOA09 [Garcia et al., 2010] and chlorophyll-a concentrations (Global SeaWIFS chlorophyll mean Sept. 1997-Dec. 2004). Nearshore sites were omitted from the chlorophyll-a comparison because the data are unreliable. Sedimentation rate estimates are based on published age models (see Table 1 for references). Oxvgen concentrations, sedimentation rates, and chlorophyll-a concentrations are not completely independent from one another and all are related to $\Delta \delta^{15}$ N for the trap sites. Oxygen is linearly related while the relation to sedimentation rates is log linear. In both cases, the correlations are statistically significant at a 95% confidence interval. The linear correlation between $\Delta \delta^{15}$ N and Chl-*a* is weaker and is not statistically significant, although this may be related to the omission of the highly productive margin sites from this analysis (Figure 5). In addition, all are linearly related to water depth. The water depth versus $\Delta \delta^{15}$ N comparison gives the best fit, with an r^2 value of 0.46, suggesting that the degree of sedimentary $\Delta \delta^{15}$ N alteration is a function of oxygen exposure time, consistent with the suggestion of *Möbius et al.* [2010, 2011]. Water depth is a robust proxy for exposure time because early burial, or the period in which sediments are exposed to oxygen-rich bottom waters and active microbial processes, is generally prolonged in the deep sea due to slower sediment accumulation and microbial metabolic rates [D'Hondt et al., 2009; Roy et al., 2012]. However, these are not strictly depth related processes, but are also a function of distance from land, sediment type, productivity in the overlying surface water and the chemistry of the water bathing the seafloor at a given site [Alkhatib et al., 2012]. The Japan Trench samples serve as a good example of an exception to the water depth generalization, in that they do not fall along the trend of $\sim 1\%$ of alteration per km of water depth, but rather more like 0.5‰ [Nakatsuka et al., 1997]. The lower degree of alteration for their given depth (9200 m) is likely due to the sites proximity to land and relatively rapid sediment accumulation rates when compared to the deep gyres.

[16] If we take depth as a robust proxy for the controls on alteration and consider significant changes in burial depth at a given site unlikely on orbital timescales or without major changes in the site location (e.g., tectonic transport away from a spreading ridge), then significant temporal changes in the degree of alteration at a given site are unlikely as well. Furthermore, regionally coherent downcore records of $\delta^{15}N$ changes, across a range of sedimentation rates and sediment compositions, from the continental margin to the central/ western Equatorial Pacific argue against significant changes in preservation as a function of realistic changes in accumulation rates at a single site [Galbraith et al., 2008a; Kao et al., 2008; Martinez et al., 2006]. The data suggest that spatial comparisons using absolute $\delta^{15}N$ values should consider the potential impact of alteration at deeper/more distal sites. However, when comparing the average offset of 2.3% to the total range of surface sedimentary δ^{15} N values compiled in the NICOPP database of 15‰, this seems minor (Galbraith et al., submitted manuscript, 2012). Finally, the $\sim 2.3\%$ average offset gleaned from the sediment trap compilation is, interestingly, approximately equal to the 2.5% mean offset between predicted surface sediment $\delta^{15}N$ values derived from simulations with a global ocean biogeochemical model incorporating nitrogen isotopes [Somes et al., 2010] and the global compilation of observed surface sediment δ^{15} N values (Galbraith et al., submitted manuscript, 2012).

6. Avoiding Alteration in Paleo Reconstructions

[17] In order to avoid alteration, work with sedimentary N isotopes proceeded with a two-pronged approach, using bulk δ^{15} N measurements on the margins where fidelity was fairly certain and developing new N archives that were less prone to alteration for the study of relatively slowly accumulating sediments, such as the Southern Ocean and the oligotrophic regions [*Altabet and Curry*, 1989; *Higgins et al.*, 2009; *Ren et al.*, 2009; *Robinson et al.*, 2004; *Sachs and Repeta*, 1999; *Shemesh et al.*, 1993]. These include microfossil bound N isotope proxies, developed to tap into a mineral bound

organic fraction, and compound specific measurements of chlorophyll and chlorophyll degradation products such as those discussed in the Mediterranean Sea example. Both proxy types assume that the δ^{15} N value being measured is 1) a reflection of water column variability, rather than species- or molecule-specific biases, and 2) protected from alteration during sinking and burial.

[18] These alternative δ^{15} N proxies all have the potential to allow a fresh look at the nitrogen cycle by opening up the field of reconstructions to include the margins and open ocean pelagic settings where preservation of bulk organic nitrogen is questionable or the organic matter content is so low that it precludes a robust measurement by EA-IRMS. Both culture work and downcore reconstructions that compare bulk δ^{15} N values to the δ^{15} N values of the novel N isotope proxies themselves allow for added investigations into the integrity of the two signals.

[19] The predominance of sedimentary $\delta^{15}N$ records from the margins and a potentially unjustified skepticism of downcore records from low productivity regions have impeded our ability to study the global marine nitrogen cycle in the past. Bulk analyses are inexpensive and rapid and appear more robust than previously perceived. Testing regional patterns of alteration by comparing the $\delta^{15}N$ of nitrate, sinking particles and surface sediments is useful but not always feasible. Moreover, it does not exclude the possibility of differences in preservation in the past. We suggest that the routine adoption of simple evaluation tools can help detect potential problems with bulk δ^{15} N in sediment cores. For example, a nonzero y-intercept in a plot of total organic carbon versus total nitrogen is indicative of the presence of inorganic nitrogen, likely NH⁴ fixed in clays [*Calvert*, 2004; Kienast et al., 2005; Schubert and Calvert, 2001] or inputs of terrestrial organics, with their distinct C/N. Given significant deviations from a 1:1 line, the inorganic N pool can be measured and the organic N δ^{15} N calculated. This correction gives a value for ON, but it does not avoid variable preservation of the ON.

7. Summary and Outlook

[20] Over the last several decades the use of sedimentary δ^{15} N as a tool to study the marine nitrogen cycle has expanded. We have an excellent picture of variability in rapidly accumulating sediments from the last \sim 50–100 kyr. However, data from the oligotrophic ocean are few, due to concern regarding alteration. Studies that have sought to unravel the exact mechanisms responsible for isotopic shifts in the bulk sedimentary organic N pool have been largely inconclusive due to the difficulty of discerning slow sediment alteration directly. However, empirical data, including the sediment trap results compared here, suggest that even in slowly accumulating regions of the ocean, bulk sedimentary δ^{15} N records will primarily reflect changes in the δ^{15} N of exported N in most cases, rather than differential alteration. This is consistent with recent comparisons between chlorin/ porphyrin $\delta^{15}N$ and bulk sedimentary $\delta^{15}N$ profiles that indicate that in sediments of poor organic matter preservation, such as the Mediterranean marls, or those millions of years old, like the Cretaceous black shales [Higgins et al., 2010, 2012], the bulk δ^{15} N presents a robust picture of biogeochemical cycling of N.

[21] The future of N isotope studies is likely to lead to an improved understanding of the proxy itself as well as the evolution of the long-term N cycle and its importance for modulating climatic and ecological changes. At present, published N isotope records largely span the last 100 kyr with only a few on the order of 3-5 Ma and older (e.g., Ocean Anoxic Events/the Archean [Garvin et al., 2009; Higgins et al., 2012; Jenkyns et al., 2007; Junium and Arther, 2007]). While there is value in examining relative changes in sedimentary δ^{15} N across an ancient interval, it is difficult to place the data in context without a continuous, robust record of δ^{15} N in the ocean. This is essential for distinguishing between global marine nitrogen cycle processes and regional phenomena. Examining N cycle processes in ancient sediments will also require further evaluation of isotopic alteration on extremely long timescales.

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